



Universidade de São Paulo
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**Desenvolvimento de nanoimunossensores de microscopia de
força atômica para estudo da esclerose múltipla**

Tese apresentada ao Instituto de Medicina Tropical da
Universidade de São Paulo para obtenção do título de
Doutora em Ciências.

Área de concentração: Doenças Tropicais e Saúde
Internacional.

Orientador: Prof. Dr. Fabio de Lima Leite

Co-orientadora: Profa. Dra. Doralina G. Brum Souza

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Descritores: 1. MICROSCOPIA DE FORÇA ATÔMICA. 2. ESPECTROSCOPIA ATÔMICA. 3. ESCLEROSE MÚLTIPLA. 4. BAINHA DE MIELINA. 5. SISTEMA NERVOSO CENTRAL. 6. SENSOR.

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Somewhere, something incredible is waiting to be known."

Carl Sagan

Resumo

Garcia PS. Desenvolvimento de nanoimunossensores de microscopia de força atômica para estudo da esclerose múltipla (tese). São Paulo: Instituto de Medicina Tropical da Universidade de São Paulo; 2018.

A glicoproteína oligodendrocítica da mielina (MOG) e proteína básica da mielina (MBP) têm sido implicadas como os antígenos-alvo mais importantes nos processos desmielinizantes do sistema nervoso central (SNC), e mais importantes autoantígenos que surgiram dos estudos com o modelo animal para a esclerose múltipla (EM), a encefalomielite autoimune experimental (EAE). Os primeiros autoanticorpos detectados no soro e líquido cefalorraquidiano (LCR) de pacientes com EM foram anticorpos contra antígenos da mielina. O diagnóstico diferencial da EM inclui a presença de bandas oligoclonais (BOCs) no LCR e ausência no soro, demonstrando dessa forma síntese intratecal de imunoglobulinas G (IgG). As técnicas de detecção de anticorpos mais utilizadas atualmente são ELISA, ensaio baseado em células e *western blot* (WB). Neste contexto, o estudo da anti-MOG e anti-MBP e seu papel na EM podem ser estudados através da técnica de espectroscopia de força atômica (AFS). Esta é uma técnica altamente sensível que permite a detecção molecular, com a interação de uma ponta funcionalizada de microscópio de força atômica (AFM) com uma amostra, a qual fornece desta forma a força de adesão (Fad) específica para o sistema. Nesta pesquisa, foi inserido na ponta de AFM funcionalizada os peptídeos encefalitogênicos MOG₉₂₋₁₀₆ e MBP₈₅₋₉₉, para detectar e estudar os anticorpos específicos IgG anti- MOG₉₂₋₁₀₆ e MBP₈₅₋₉₉, no soro e LCR de pacientes na amostra, utilizando a técnica AFS. Sendo assim, este estudo foi realizado de forma inédita utilizando a AFS, auxiliando diretamente na investigação da EM e doenças desmielinizantes relacionadas.

Descritores: Microscopia de Força Atômica. Espectroscopia Atômica. Esclerose Múltipla. Bainha de Mielina. Sistema Nervoso Central. Sensor.

Abstract

Garcia PS. Development of atomic force microscopy nanoimmunosensor applied to the survey of multiple sclerosis (thesis). São Paulo: Instituto de Medicina Tropical da Universidade de São Paulo, Brazil; 2018.

Myelin oligodendrocyte glycoprotein (MOG) and myelin basic protein (MBP) have been implied as the most important target antigens in demyelinating processes of central nervous system (CNS) and the most important antigens candidates whom arised from the animal model for multiple sclerosis (MS), the experimental autoimmune encephalomyelitis (EAE). The first autoantibodies detected in serum and cerebrospinal fluid (CSF) of MS patients were antibodies against myelin antigens. Differential diagnostic to MS includes the presence of oligoclonal bands (OCBs) in CSF and absence in serum, which demonstrate intrathecal IgG synthesis. Most applied techniques to detection of antibodies nowadays are ELISA, cell-based assay and western blot (WB). In this context, the study of anti-MOG role in e disease may be supported through its detection by atomic force spectroscopy (AFS) technique. AFS is a highly sensitive technique that allows molecular detection as a functionalized atomic force microscope (AFM) tip interacts with the sample, providing the system specific adhesion force (Fad). In this research, it was attached in the functionalized AFM tip the notable encephalitogenic peptides MOG₉₂₋₁₀₆ and MBP₈₅₋₉₉ to detect and study the specific antibodies anti-MOG₉₂₋₁₀₆ and anti-MBP₈₅₋₉₉ on the sample with AFS technique. Thus, this study was applied for the first time in research with AFS, assisting directly to MS and other demyelinating diseases investigation.

Descriptors: Atomic Force Microscopy. Force Spectroscopy. Multiple Sclerosis. Myelin Sheath. Central Nervous System. Sensor.

CONCLUSÕES

Este foi o primeiro trabalho experimental realizado pelo nosso grupo, o GNN, para o estudo de doenças desmielinizantes utilizando sensores de ponta de AFM com enfoque na EM. As sequências da MOG e MBP foram escolhidas por serem amplamente estudadas na indução de EAE e os primeiros autoanticorpos detectados, tanto no soro quanto no LCR de pacientes com EM.

Nesta pesquisa, foi desenvolvido com sucesso um protocolo adequado de funcionalização de superfícies para a detecção da anti-MOG, validado pelos métodos AFS e caracterização aplicados, e corroborado pelo método *in silico* para o mesmo sistema; assim, os seguintes pontos puderam ser concluídos:

- a) Foi desenvolvido um protocolo eficiente para funcionalização química de pontas de AFM e substratos, para o estudo de autoanticorpos envolvidos na EM;
- b) A anti-MOG₉₂₋₁₀₆ e anti-MBP₈₅₋₉₉ foram detectadas com êxito utilizando o sensor desenvolvido, tanto no controle específico quanto nas IgGs purificadas dos pacientes;
- c) uma vez que os anticorpos específicos foram detectados somente nos pacientes que estavam em surto e no LCR, pôde-se concluir que o nanossensor desenvolvido é capaz de detectar síntese intratecal de IgG, e dessa forma, pacientes em processos desmielinizantes, em especial na EM no início da doença;
- d) utilizando as técnicas de caracterização, juntamente com os resultados satisfatórios de AFS, foi possível verificar que as superfícies estavam devidamente funcionalizadas;
- e) foi possível comparar os resultados teóricos obtidos por SMD para os valores de força de adesão com os resultados experimentais, dando suporte à detecção dos autoanticorpos com o nanoimunossensor desenvolvido.

Foi verificado que, com o sensor desenvolvido, é possível detectar surto (estágios iniciais da EMRR), e pode ser utilizado como um sensor de anti-MOG e anti-MBP para verificação de síntese intratecal e desmielinização, juntamente com o quadro clínico estabelecido pelo neurologista com a aplicação dos critérios de McDonald. Estudos futuros serão realizados com um n amostral maior. Sendo assim, o estudo destes anticorpos auto-reativos em pacientes portadores de doenças desmielinizantes com esta técnica de vanguarda é uma ferramenta importante para o estudo antígeno-anticorpo neste contexto, e ainda poderá em breve auxiliar a elucidar ainda outras questões da presença dos mesmos nos pacientes.

Dessa forma, os achados podem fornecer pistas importantes para a participação dos autoanticorpos estudados nas doenças desmielinizantes, bem como na EM, uma vez que o valor da anti-MOG e anti-MBP ainda são conflitantes na literatura; mas seu envolvimento nos processos neurodegenerativos é inegável.

Assim, o estudo da presença de anticorpos contra sequências da MOG e MBP, em particular das sequências MOG₉₂₋₁₀₆ e MBP₈₅₋₉₉ na EM, é substancial tanto para o entendimento da doença, quanto para o desenvolvimento de novos sensores, a busca de biomarcadores para sua detecção ou contribuição para o tratamento da EM. Neste contexto, a microscopia de força atômica pôde ser aplicada como método de detecção para estudar e entender os mecanismos moleculares envolvidos com a EM.

REFERÊNCIAS BIBLIOGRÁFICAS

- 1 Kipnis J, Filiano AJ. Neuroimmunology in 2017: The central nervous system: privileged by immune connections. *Nat Rev Immunol*. 2018 Feb;18(2):83-84.
- 2 Leite FL, Hausen M, Oliveira GS, Brum DG, Oliveira ON. Nanoneurobiophysics: new challenges for diagnosis and therapy of neurologic disorders. *Nanomed* 2015;10(23):3417-9.
- 3 Drolle E, Hane F, Lee B, Leonenko Z. Atomic force microscopy to study molecular mechanisms of amyloid fibril formation and toxicity in Alzheimer's disease. *Drug Metab Rev*. 2014 May;46(2):207-23.
- 4 Steffens C, Leite FL, Bueno CC, Manzoli A, Herrmann PSDP. Atomic force microscopy as a tool applied to nano/biosensors. *Sensors*. 2012;12(6):8278-300.
- 5 Leite F. Theoretical Models for Surface Forces and Adhesion and Their Measurement Using Atomic Force Microscopy. *Int J Mol Sci*. 2012 Oct 8;13(10):12773-856.
- 6 Deda DK, Bueno C de C, Ribeiro GA, Moraes A de S, Garcia PS, Brito B, *et al*. Atomic force microscopy-based molecular recognition: a promising alternative to environmental contaminants detection. *Curr. Microsc. Contrib Adv Sci Technol* 2012; (5): 1–30.
- 7 Soto Garcia P, Moreau ALD, Magalhaes Ierich JC, Araujo Vig AC, Higa AM, Oliveira GS, *et al*. A Nanobiosensor Based on 4-Hydroxyphenylpyruvate Dioxygenase Enzyme for Mesotrione Detection. *IEEE Sens J*. 2015;(15):2106–13.
- 8 Deda DK, Pereira BBS, Bueno CC, Silva AN da, Ribeiro GA, Amarante AM, *et al*. The use of functionalized AFM tips as molecular sensors in the detection of pesticides. *Mater Res*. 2013;(16):683–7.
- 9 da Silva ACN, Deda DK, Bueno CC, Moraes AS, Da Roz AL, Yamaji FM, *et al*. Nanobiosensors Exploiting Specific Interactions Between an Enzyme and Herbicides in Atomic Force Spectroscopy. *J Nanosci Nanotechnol*. 2014;(14):6678–84.
- 10 Leite FL, Herrmann PSP. Application of atomic force spectroscopy (AFS) to studies of adhesion phenomena: a review. *J Adhes Sci Technol*. 2005;(19):365–405.
- 11 Janissen R, Oberbarnscheidt L, Oesterhelt F. Optimized straight forward procedure for covalent surface immobilization of different biomolecules for single molecule applications. *Colloids Surf B Biointerfaces*. 2009;(71):200–7.

- 12 Etchegaray A, Bueno C de C, Teschke O. Identification of microcistin LR at the molecular level using atomic force microscopy. *Quím Nova*. 2010; (33):1843–8.
- 13 Castro Bueno C, Moraes Amarante A, Oliveira GS, Kotra Deda D, Teschke O, de Faria Franca E, *et al*. Nanobiosensor for Diclofop Detection Based on Chemically Modified AFM Probes. *IEEE Sens J*. 2014;(14):1467–75.
- 14 da Silva ACN, Deda DK, da Róz AL, Prado RA, Carvalho CC, Viviani V, *et al*. Nanobiosensors Based on Chemically Modified AFM Probes: A Useful Tool for Metsulfuron-Methyl Detection. *Sensors*. 2013;(13):1477–89.
- 15 Amarante AM, Oliveira GS, Bueno CC, Cunha RA, Ierich JCM, Freitas LCG, *et al*. Modeling the coverage of an AFM tip by enzymes and its application in nanobiosensors. *J Mol Graph Model*. 2014;(53):100–4.
- 16 Goldenberg MM. Multiple Sclerosis Review. *Pharm Ther* 2012;(37):175–84.
- 17 Gama PD da, Machado L, Ramos SD, Livramento J, Antonio, Gomes H, *et al*. Oligoclonal Bands in Cerebrospinal Fluid of Black Patients with Multiple Sclerosis, Oligoclonal Bands in Cerebrospinal Fluid of Black Patients with Multiple Sclerosis. *BioMed Res Int BioMed Res Int*. 2015; (2015): ID 217961.
- 18 Olival GS do, Lima LCP, Lima GPS, Tilbery CP. Clinical predictors of response to immunomodulators for multiple sclerosis. *Arq Neuropsiquiatr*. 2012; (70):12–6.
- 19 Lassmann H. Mechanisms of white matter damage in multiple sclerosis. *Glia* 2014; (62):1816–30.
- 20 Correale J, Farez MF. The Role of Astrocytes in Multiple Sclerosis Progression. *Front Neurol*. 2015 Aug 18; (6):180.
- 21 Gama PD. Estudo de bandas oligoclonais restritas ao líquido cefalorraquidiano em pacientes com esclerose múltipla na cidade de São Paulo [tese]. São Paulo: Faculdade de Medicina, Universidade de São Paulo; 2009.
- 22 Mirshafiey A, Kianiaslani M. Autoantigens and autoantibodies in multiple sclerosis. *Iran J Allergy Asthma Immunol*. 2013;(12):292–303.
- 23 Egg R, Reindl M, Deisenhammer F, Linington C, Berger T. Anti-MOG and anti-MBP antibody subclasses in multiple sclerosis. *Mult Scler Houndmills Basingstoke Engl*. 2001;(7):285–9.
- 24 Miller SD, Karpus WJ, Davidson TS. Experimental Autoimmune Encephalomyelitis in the Mouse. *Curr Protoc Immunol Ed John E Coligan Al*. 2007 May;Chapter 15:Unit 15.1.

- 25 Constantinescu CS, Farooqi N, O'Brien K, Gran B. Experimental autoimmune encephalomyelitis (EAE) as a model for multiple sclerosis (MS). *Br J Pharmacol*. 2011;(164):1079–106.
- 26 Robinson AP, Harp CT, Noronha A, Miller SD. The experimental autoimmune encephalomyelitis (EAE) model of MS: utility for understanding disease pathophysiology and treatment. *Handb Clin Neurol*. 2014;(122):173–89.
- 27 Tsunoda I, Kuang LQ, Theil DJ, Fujinami RS. Antibody association with a novel model for primary progressive multiple sclerosis: induction of relapsing-remitting and progressive forms of EAE in H2s mouse strains. *Brain Pathol Zurich Switz*. 2000;(10):402–18.
- 28 Lima E, Guimarães J, Pereira A, Bodas A, Delgado L, Sá MJ. Determinação de Anticorpos Anti-Mielina na Esclerose Múltipla. *Arq Med*. 2008;(22):107–11.
- 29 Baumann N, Pham-Dinh D. Biology of oligodendrocyte and myelin in the mammalian central nervous system. *Physiol Rev*. 2001;(81):871–927.
- 30 Woolsey TA, Hanaway J, Gado MH. The Brain Atlas: A Visual Guide to the Human Central Nervous System. *Am. Journ. of Neuror*. 2008; 29(5):e38.
- 31 Silver J, Schwab ME, Popovich PG. Central Nervous System Regenerative Failure: Role of Oligodendrocytes, Astrocytes, and Microglia. *Cold Spring Harb Perspect Biol*. 2015;(7):a020602.
- 32 Reindl M, Di Pauli F, Rostásy K, Berger T. The spectrum of MOG autoantibody-associated demyelinating diseases. *Nat Rev Neurol*. 2013;(9):455–61.
- 33 Ramanathan S, Dale RC, Brilot F. Anti-MOG antibody: The history, clinical phenotype, and pathogenicity of a serum biomarker for demyelination. *Autoimmun Rev*. 2016;(15):307–24.
- 34 Warsi N, Moore C, Antel J, Bar-Or A. Distinct Immune Cell Subsets Influence Astrocytes To Express Factors Relevant for Myelin Repair (P05.169). *Neurology*. 2013;(80):05-169.
- 35 Kinzel S, Weber MS. The Role of Peripheral CNS-Directed Antibodies in Promoting Inflammatory CNS Demyelination. *Brain Sci*. 2017; Jun 22;7(7). pii: E70.
- 36 Passos D *et al*. MOG-IgG-Associated Optic Neuritis, Encephalitis, and Myelitis: Lessons Learned From Neuromyelitis Optica Spectrum Disorder. *Front Neurol*. 2018; 9: 217.
- 37 Brum DG, Comini-Frota ER, Vasconcelos CCF, Dias-Tosta E, Brum DG, Comini-Frota ER, *et al*. Supplementation and therapeutic use of vitamin D in patients

with multiple sclerosis: Consensus of the Scientific Department of Neuroimmunology of the Brazilian Academy of Neurology. *Arq Neuropsiquiatr.* 2014;(72):152–6.

38 Briggs FBS, Acuna B, Shen L, Ramsay P, Quach H, Bernstein A, *et al.* Smoking and risk of multiple sclerosis: evidence of modification by NAT1 variants. *Epidemiol Camb Mass.* 2014;(25):605–14.

39 Brum DG, Luizon MR, Santos AC, Lana-Peixoto MA, Rocha CF, Brito ML, *et al.* European Ancestry Predominates in Neuromyelitis Optica and Multiple Sclerosis Patients from Brazil. *Plos One.* 2013;(8):e58925.

40 Fuchs KL. Relapsing–Remitting Multiple Sclerosis. In: Kreutzer JS, DeLuca J, Caplan B, editors. *Encycl. Clin. Neuropsychol.*, Springer New York; 2011, 2145–9.

41 Bertotti AP, Lenzi MCR, Portes JRM. O portador de Esclerose Múltipla e suas formas de enfrentamento frente à doença. *Barbaroi* 2011:101–24.

42 Lublin FD, Reingold SC, Cohen JA, Cutter GR, Sørensen PS, Thompson AJ, *et al.* Defining the clinical course of multiple sclerosis The 2013 revisions. *Neurology.* 2014; (10):1212.

43 Fitzner D, Simons M. Chronic Progressive Multiple Sclerosis – Pathogenesis of Neurodegeneration and Therapeutic Strategies. *Curr Neuropharmacol.* 2010;(8):305–15.

44 Fitzner B, Hecker M, Zettl UK. Molecular biomarkers in cerebrospinal fluid of multiple sclerosis patients. *Autoimmun Rev.* 2015;(14):903–13.

45 Kurtzke JF. Rating neurologic impairment in multiple sclerosis: an expanded disability status scale (EDSS). *Neurology.* 1983;(33):1444–52.

46 Winger RC, Zamvil SS. Antibodies in multiple sclerosis oligoclonal bands target debris. *Proc Natl Acad Sci USA.* 2016;(113):7696–8.

47 Kolb-Mäurer A, Goebeler M, Mäurer M. Cutaneous Adverse Events Associated with Interferon- β Treatment of Multiple Sclerosis. *Int J Mol Sci.* 2015;(16):14951–60.

48 Normann B, Sørgaard KW, Salvesen R, Moe S. Contextualized perceptions of movement as a source of expanded insight: People with multiple sclerosis' experience with physiotherapy. *Physiother Theory Pract.* 2013;(29):19–30.

49 Silva VM, Silva DF. Esclerose Múltipla: imunopatologia, diagnóstico e tratamento – Artigo de revisão. *Interf Cient - Saúde e Ambiente.* 2014;(2):81–90.

50 Lehmann-Horn K, Kronsbein HC, Weber MS. Targeting B cells in the treatment of multiple sclerosis: recent advances and remaining challenges. *Ther Adv Neurol Disord.* 2013;(6):161–73.

- 51 Ziemssen T, Schrempf W. Glatiramer acetate: mechanisms of action in multiple sclerosis. *Int Rev Neurobiol.* 2007;(79):537–70.
- 52 Cohen J, Belova A, Selmaj K, *et al.* Equivalence of generic glatiramer acetate in multiple sclerosis: A randomized clinical trial. *JAMA Neurol.* 2015:1–9.
- 53 Martinelli Boneschi F, Vacchi L, Rovaris M, Capra R, Comi G. Mitoxantrone for multiple sclerosis. *Cochrane Database Syst Rev.* 2013;(5):CD002127.
- 54 Steinman L. Blocking adhesion molecules as therapy for multiple sclerosis: natalizumab. *Nat Rev Drug Discov.* 2005;(4):510–8.
- 55 Steinman L. Immunology of Relapse and Remission in Multiple Sclerosis. *Annu Rev Immunol.* 2014;(32):257–81.
- 56 Pelletier D, Hafler DA. Fingolimod for Multiple Sclerosis. *N Engl J Med.* 2012;(366):339–47.
- 57 Friese MA, Schattling B, Fugger L. Mechanisms of neurodegeneration and axonal dysfunction in multiple sclerosis. *Nat Rev Neurol.* 2014;(10):225–38.
- 58 Shetty A, Gupta SG, Varrin-Doyer M, Weber MS, Prod'homme T, Molnarfi N, *et al.* Immunodominant T-cell epitopes of MOG reside in its transmembrane and cytoplasmic domains in EAE. *Neurol Neuroimmunol Neuroinflammation.* 2014;(1):e22.
- 59 Weber MS, Hemmer B, Cepok S. The role of antibodies in multiple sclerosis. *Biochim Biophys Acta BBA - Mol Basis Dis.* 2011;(1812):239–45.
- 60 Sospedra M, Martin R. Immunology of Multiple Sclerosis. *Annu Rev Immunol.* 2005;(23):683–747.
- 61 Marta CB, Oliver AR, Sweet RA, Pfeiffer SE, Ruddle NH. Pathogenic myelin oligodendrocyte glycoprotein antibodies recognize glycosylated epitopes and perturb oligodendrocyte physiology. *Proc Natl Acad Sci U S A.* 2005;(102):13992–7.
- 62 Carvalho A, Sant'anna G, Santos CC, Frugulhetti IP, Leon SA, Quirico-Santos T. Determination of autoantibody for myelin antigens in the serum of patients HLA - DQB1*0602 with multiple sclerosis. *Arq Neuropsiquiatr.* 2003;(61):968–73.
- 63 Gerdoni E, Gallo B, Casazza S, Musio S, Bonanni I, Pedemonte E, *et al.* Mesenchymal stem cells effectively modulate pathogenic immune response in experimental autoimmune encephalomyelitis. *Ann Neurol.* 2007;(61):219–27.
- 64 Ercolini AM, Miller SD. Mechanisms of immunopathology in murine models of central nervous system demyelinating disease. *J Immunol Baltim Md.* 2006 Mar 15;176(6):3293-8.

- 65 Reindl M, Linington C, Brehm U, Egg R, Dilitz E, Deisenhammer F, *et al.* Antibodies against the myelin oligodendrocyte glycoprotein and the myelin basic protein in multiple sclerosis and other neurological diseases: a comparative study. *Brain J Neurol.* 1999;122 (11):2047–56.
- 66 Degano AL, Ditamo Y, Roth GA. Neuronal antigens modulate the immune response associated with experimental autoimmune encephalomyelitis. *Immunol Cell Biol.* 2004;(82):17–23.
- 67 Frohman EM, Racke MK, Raine CS. Multiple Sclerosis — The Plaque and Its Pathogenesis. 2006:942–55.
- 68 Polman CH, Reingold SC, Banwell B, Clanet M, Cohen JA, Filippi M, *et al.* Diagnostic criteria for multiple sclerosis: 2010 revisions to the McDonald criteria. *Ann Neurol.* 2011;(69):292–302.
- 69 McDonald WI, Compston A, Edan G, Goodkin D, Hartung HP, Lublin FD, *et al.* Recommended diagnostic criteria for multiple sclerosis: guidelines from the International Panel on the diagnosis of multiple sclerosis. *Ann Neurol.* 2001;(50):121–7.
- 70 Minguetti G. Magnetic resonance imaging in multiple sclerosis: analysis of 270 cases. *Arq Neuropsiquiatr.* 2001;(59):563–9.
- 71 Marzia Puccioni-Sohler *et al.* Esclerose múltipla: correlação clínico-laboratorial. *Arq. Neuro-Psiquiatr.* 2001; (59):1678.
- 72 Aidar RC, Suzuki FA. Potencial evocado miogênico vestibular: novas perspectivas diagnósticas em esclerose múltipla. *Braz J Otorhinolaryngol.* 2005;(71):7299.
- 73 Katsavos S, Anagnostouli M, Katsavos S, Anagnostouli M. Biomarkers in Multiple Sclerosis: An Up-to-Date Overview, Biomarkers in Multiple Sclerosis: An Up-to-Date Overview. *Mult Scler Int Mult Scler Int.* 2013; (2013):e340508.
- 74 Katrin Paap B. Molecular Biomarkers in Multiple Sclerosis. *J Clin Cell Immunol.* 2013; (10):009.
- 75 Lesko LJ, AJ Atkinson J. Use of biomarkers and surrogate endpoints in drug development and regulatory decision making: Criteria, Validation, Strategies. *Annu Rev Pharmacol Toxicol.* 2001;(41):347–66.
- 76 Filippi M *et al.* KIR4.1: another misleading expectation in multiple sclerosis? *The Lancet Neurol.* 2014; (13):753-755,.
- 77 Cantó E, Comabella M. Biomarcadores en la esclerosis múltiple: estado actual. *Rev Esp Mult Escl.* 2012; (23): 20-29.

- 78 Tanaka M, Tanaka K. Anti-MOG antibodies in adult patients with demyelinating disorders of the central nervous system. *J Neuroimmunol*. 2014;(270):98–9.
- 79 Hohlfeld R, Dornmair K, Meinl E, Wekerle H. The search for the target antigens of multiple sclerosis, part 2: CD8+ T cells, B cells, and antibodies in the focus of reverse-translational research. *Lancet Neurol*. 2016;(15):317–31.
- 80 Kursula P. Structural properties of proteins specific to the myelin sheath. *Amino Acids*. 2006;(34):175–85.
- 81 Mantegazza R, Cristaldini P, Bernasconi P, Baggi F, Pedotti R, Piccini I, *et al*. Anti-MOG autoantibodies in Italian multiple sclerosis patients: specificity, sensitivity and clinical association. *Int Immunol*. 2004;(16):559–65.
- 82 Spadaro M, Meinl E. Detection of Autoantibodies Against Myelin Oligodendrocyte Glycoprotein in Multiple Sclerosis and Related Diseases. In: Weissert R, editor. *Mult. Scler*. Springer. New York; 2016, 99–104.
- 83 Havla J, Kümpfel T, Schinner R, Spadaro M, Schuh E, Meinl E, *et al*. Myelin-oligodendrocyte-glycoprotein (MOG) autoantibodies as potential markers of severe optic neuritis and subclinical retinal axonal degeneration. *J Neurol*. 2017;(264):139–51.
- 84 Spadaro M, Gerdes LA, Krumbholz M, Ertl-Wagner B, Thaler FS, Schuh E, *et al*. Autoantibodies to MOG in a distinct subgroup of adult multiple sclerosis. *Neurol Neuroimmunol Neuroinflammation*. Oct 2016; (3):5.
- 85 Hecker M, Fitzner B, Wendt M, Lorenz P, Flechtner K, Steinbeck F, *et al*. High-Density Peptide Microarray Analysis of IgG Autoantibody Reactivities in Serum and Cerebrospinal Fluid of Multiple Sclerosis Patients. *Mol Cell Proteomics MCP*. 2016;(15):1360–80.
- 86 Hemmer B, Archelos JJ, Hartung H-P. New concepts in the immunopathogenesis of multiple sclerosis. *Nat Rev Neurosci*. 2002;(3):291–301.
- 87 Sherman DL, Brophy PJ. Mechanisms of axon ensheathment and myelin growth. *Nat Rev Neurosci*. 2005;(6):683–90.
- 88 Minagar A, Alexander JS. Blood-brain barrier disruption in multiple sclerosis. *Mult Scler Houndmills Basingstoke Engl*. 2003;(9):540–9.
- 89 Dorovini-Zis K. *The Blood-Brain Barrier in Health and Disease, Volume Two: Pathophysiology and Pathology*. CRC Press; 2015.
- 90 Montagne, A *et al*. Blood-Brain Barrier Breakdown in the Aging Human Hippocampus *Neuron*. 2015; 85(2): 296–302..

- 91 Mahad DH, Trapp BD, Lassmann H. Pathological mechanisms in progressive multiple sclerosis. *Lancet Neurol.* 2015;(14):183–93.
- 92 Steinman L, Zamvil S. Transcriptional analysis of targets in multiple sclerosis. *Nat Rev Immunol.* 2003;(3):483–92.
- 93 Fraussen J, Claes N, de Bock L, Somers V. Targets of the humoral autoimmune response in multiple sclerosis. *Autoimmun Rev.* 2014;(13):1126–37.
- 94 Mayer MC, Meinl E. Glycoproteins as targets of autoantibodies in CNS inflammation: MOG and more. *Ther Adv Neurol Disord.* 2012;(5):147–59.
- 95 Goverman J. Autoimmune T cell responses in the central nervous system. *Nat Rev Immunol.* 2009;(9):393–407.
- 96 McLaughlin KA, Wucherpfennig KW. B cells and autoantibodies in the pathogenesis of multiple sclerosis and related inflammatory demyelinating diseases. *Adv Immunol.* 2008;(98):121–49.
- 97 Nylander A, Hafler DA. Multiple sclerosis. *J Clin Invest.* 2012;(122):1180–8.
- 98 Elong Ngonu A, Pettré S, Salou M, Bahbouhi B, Soullillou J-P, Brouard S, *et al.* Frequency of circulating autoreactive T cells committed to myelin determinants in relapsing-remitting multiple sclerosis patients. *Clin Immunol Orlando Fla.* 2012;(144):117–26.
- 99 Platten M, Steinman L. Multiple sclerosis: trapped in deadly glue. *Nat Med.* 2005;(11):252–3.
- 100 Calabrese M, Magliozzi R, Ciccarelli O, Geurts JGG, Reynolds R, Martin R. Exploring the origins of grey matter damage in multiple sclerosis. *Nat Rev Neurosci.* 2015;(16):147–58.
- 101 Xiao BG, Linington C, Link H. Antibodies to myelin-oligodendrocyte glycoprotein in cerebrospinal fluid from patients with multiple sclerosis and controls. *J Neuroimmunol.* 1991;(31):91–6.
- 102 Dendrou CA, Fugger L, Friese MA. Immunopathology of multiple sclerosis. *Nat Rev Immunol.* 2015;(15):545–58.
- 103 Berger T, Rubner P, Schautzer F, Egg R, Ulmer H, Mayringer I, *et al.* Antimyelin Antibodies as a Predictor of Clinically Definite Multiple Sclerosis after a First Demyelinating Event. *N Engl J Med.* 2003;(349):139–45.
- 104 Tomassini V, De Giglio L, Reindl M, Russo P, Pestalozza I, Pantano P, *et al.* Anti-myelin antibodies predict the clinical outcome after a first episode suggestive of MS. *Mult Scler Houndmills Basingstoke Engl.* 2007;(13):1086–94.

- 105 Khalil M, Reindl M, Lutterotti A, Kuenz B, Ehling R, Gneiss C, Lackner P, Deisenhammer F, Berger T. Epitope specificity of serum antibodies directed against the extracellular domain of myelin oligodendrocyte glycoprotein: Influence of relapses and immunomodulatory treatments. *J Neuroimmunol.* 2006;(174):147–56.
- 106 Lampasona V, Franciotta D, Furlan R, Zanaboni S, Fazio R, Bonifacio E, *et al.* Similar low frequency of anti-MOG IgG and IgM in MS patients and healthy subjects. *Neurology.* 2004;(62):2092–4.
- 107 Haase CG, Guggenmos J, Brehm U, Andersson M, Olsson T, Reindl M, *et al.* The fine specificity of the myelin oligodendrocyte glycoprotein autoantibody response in patients with multiple sclerosis and normal healthy controls. *J Neuroimmunol.* 2001;(114):220–5.
- 108 Pröbstel A-K, Rudolf G, Dornmair K, Collongues N, Chanson J-B, Sanderson NS, *et al.* Anti-MOG antibodies are present in a subgroup of patients with a neuromyelitis optica phenotype. *J Neuroinflammation.* 2015;(12):46.
- 109 Körtvélyessy P *et al.* ADEM-like presentation, anti-MOG antibodies, and MS pathology: TWO case reports. *Neurol Neuroimmunol Neuroinflamm.* 2017; 4(3): e335.
- 110 Brownlee WJ, Ciccarelli O. Is the early course of multiple sclerosis the same in adults and children? *Europ Journ of Neurol.* 2017; (24):235-236.
- 111 Waldman A, Ghezzi A, Bar-Or A, Mikaeloff Y, Tardieu M, Banwell B. Multiple sclerosis in children: an update on clinical diagnosis, therapeutic strategies, and research. *Lancet Neurol.* 2014;(13):936–48.
- 112 Ketelslegers IA, Van Pelt DE, Bryde S, Neuteboom RF, Catsman-Berrevoets CE, Hamann D, *et al.* Anti-MOG antibodies plead against MS diagnosis in an Acquired Demyelinating Syndromes cohort. *Mult Scler Houndmills Basingstoke Engl.* 2015;(21):1513–20.
- 113 Nakamura Y, Nakajima H, Tani H, Hosokawa T, Ishida S, Kimura F, *et al.* Anti-MOG antibody-positive ADEM following infectious mononucleosis due to a primary EBV infection: a case report. *BMC Neurol.* 2017;(17):76.
- 114 Alper G, Heyman R, Wang L. Multiple sclerosis and acute disseminated encephalomyelitis diagnosed in children after long-term follow-up: comparison of presenting features. *Dev Med Child Neurol.* 2009;(51):480–6.
- 115 Reindl M, Jarius S, Rostasy K, Berger T. Myelin oligodendrocyte glycoprotein antibodies: How clinically useful are they? *Curr Opin Neurol.* 2017;(30):295–301.

- 116 Zhou L, Huang Y, Li H, Fan J, Zhangbao J, Yu H, *et al.* MOG-antibody associated demyelinating disease of the CNS: A clinical and pathological study in Chinese Han patients. *J Neuroimmunol.* 2017;(305):19–28.
- 117 Hyun J-W, Woodhall MR, Kim S-H, Jeong IH, Kong B, Kim G, *et al.* Longitudinal analysis of myelin oligodendrocyte glycoprotein antibodies in CNS inflammatory diseases. *J Neurol Neurosurg Psychiatry.* 2017 Oct;88(10):811-817
- 118 Siritho S, Sato DK, Kaneko K, Fujihara K, Prayoonwiwat N. The clinical spectrum associated with myelin oligodendrocyte glycoprotein antibodies (anti-MOG-Ab) in Thai patients. *Mult Scler Houndmills Basingstoke Engl.* 2016;(22):964–8.
- 119 Kim S-M, Woodhall MR, Kim J-S, Kim S-J, Park KS, Vincent A, *et al.* Antibodies to MOG in adults with inflammatory demyelinating disease of the CNS. *Neurol Neuroimmunol Neuroinflammation.* 2015;(2):e163.
- 120 Olsson T. White matter disease: Roles of anti-MOG antibodies in demyelinating diseases : *Nature Reviews Neurology, Nature Reviews Neurology.* 2011;(5):248-9.
- 121 Gori F, Mulinacci B, Massai L, Avolio C, Caragnano M, Peroni E, *et al.* IgG and IgM antibodies to the refolded MOG(1-125) extracellular domain in humans. *J Neuroimmunol.* 2011;(233):216–20.
- 122 Zadro I, Brinar V, Horvat G, Brinar M. Clinical relevance of antibodies against myelin oligodendrocyte glycoprotein in different clinical types of multiple sclerosis. *Clin Neurol Neurosurg.* 2007;(109):23–6.
- 123 Bischof F, Bins A, Dürr M, Zevering Y, Melms A, Kruisbeek AM. A structurally available encephalitogenic epitope of myelin oligodendrocyte glycoprotein specifically induces a diversified pathogenic autoimmune response. *J Immunol Baltim Md.* 2004;(173):600–6.
- 124 Khare P, Challa DK, Devanaboyina SC, Velmurugan R, Hughes S, Greenberg BM, *et al.* Myelin oligodendrocyte glycoprotein-specific antibodies from multiple sclerosis patients exacerbate disease in a humanized mouse model. *J Autoimmun.* 2018;(86):104–15.
- 125 Belogurov A, Kudriaeva A, Kuzina E, Smirnov I, Bobik T, Ponomarenko N, *et al.* Multiple sclerosis autoantigen myelin basic protein escapes control by ubiquitination during proteasomal degradation. *J Biol Chem.* 2014;(289):17758–66.
- 126 Boggs JM. Myelin basic protein: a multifunctional protein. *Cell Mol Life Sci CMLS.* 2006;(63):1945–61.

- 127 Friedrich MG, Hancock SE, Raftery MJ, Truscott RJW. Isoaspartic acid is present at specific sites in myelin basic protein from multiple sclerosis patients: could this represent a trigger for disease onset? *Acta Neuropathol Commun.* 2016;(4):83.
- 128 Derkus B, Emregul E, Yucesan C, Cebesoy Emregul K. Myelin basic protein immunosensor for multiple sclerosis detection based upon label-free electrochemical impedance spectroscopy. *Biosens Bioelectron.* 2013;(46):53–60.
- 129 Krogsgaard M, Wucherpfennig KW, Canella B, Hansen BE, Svejgaard A, Pyrdol J, *et al.* Visualization of Myelin Basic Protein (Mbp) T Cell Epitopes in Multiple Sclerosis Lesions Using a Monoclonal Antibody Specific for the Human Histocompatibility Leukocyte Antigen (Hla)-Dr2–Mbp 85–99 Complex. *J Exp Med.* 2000;(191):1395–412.
- 130 Fridkis-Hareli M, Santambrogio L, Stern JNH, Fugger L, Brosnan C, Strominger JL. Novel synthetic amino acid copolymers that inhibit autoantigen-specific T cell responses and suppress experimental autoimmune encephalomyelitis. *J Clin Invest.* 2002;(109):1635–43.
- 131 Fridkis-Hareli M, Stern JNH, Fugger L, Strominger JL. Synthetic peptides that inhibit binding of the myelin basic protein 85-99 epitope to multiple sclerosis-associated HLA-DR2 molecules and MBP-specific T-cell responses. 2001 Aug;62(8):753-63.
- 132 Yang L, Tan D, Piao H. Myelin Basic Protein Citrullination in Multiple Sclerosis: A Potential Therapeutic Target for the Pathology. *Neurochem Res.* 2016;(41):1845–56.
- 133 Panitch HS, Hooper CJ, Johnson KP. CSF antibody to myelin basic protein. Measurement in patients with multiple sclerosis and subacute sclerosing panencephalitis. *Arch Neurol.* 1980;(37):206–9.
- 134 Derkus B, Acar Bozkurt P, Tulu M, Emregul KC, Yucesan C, Emregul E. Simultaneous quantification of Myelin Basic Protein and Tau proteins in cerebrospinal fluid and serum of Multiple Sclerosis patients using nanoimmunosensor. *Biosens Bioelectron.* 2017;(89):781–8.
- 135 Bielekova B, Goodwin B, Richert N, Cortese I, Kondo T, Afshar G, *et al.* Encephalitogenic potential of the myelin basic protein peptide (amino acids 83-99) in multiple sclerosis: results of a phase II clinical trial with an altered peptide ligand. *Nat Med.* 2000;(6):1167–75.

- 136 Smith KJ, Pyrdol J, Gauthier L, Wiley DC, Wucherpfennig KW. Crystal structure of HLA-DR2 (DRA*0101, DRB1*1501) complexed with a peptide from human myelin basic protein. *J Exp Med*. 1998;(188):1511–20.
- 137 Li Y, Li H, Martin R, Mariuzza RA. Structural basis for the binding of an immunodominant peptide from myelin basic protein in different registers by two HLA-DR2 proteins. *J Mol Biol*. 2000;(304):177–88.
- 138 Stern JNH, Illés Z, Reddy J, Keskin DB, Fridkis-Hareli M, Kuchroo VK, *et al*. Peptide 15-mers of defined sequence that substitute for random amino acid copolymers in amelioration of experimental autoimmune encephalomyelitis. *Proc Natl Acad Sci U S A*. 2005;(102):1620–5.
- 139 Triantafyllopoulou M, Ercolini AM, DeGutes M, Miller SD. Differential Induction of Experimental Autoimmune Encephalomyelitis by Myelin Basic Protein Molecular Mimics in Mice Humanized for HLA-DR2 and an MBP85-99-Specific T Cell Receptor. *J Autoimmun*. 2008;(31):399–407.
- 140 Nielsen CH, Börnsen L, Sellebjerg F, Brimnes MK. Myelin Basic Protein-Induced Production of Tumor Necrosis Factor- α and Interleukin-6, and Presentation of the Immunodominant Peptide MBP85-99 by B Cells from Patients with Relapsing-Remitting Multiple Sclerosis. *PLoS One*. 2016;(11):e0146971.
- 141 Greene MT, Ercolini AM, DeGutes M, Miller SD. Differential induction of experimental autoimmune encephalomyelitis by myelin basic protein molecular mimics in mice humanized for HLA-DR2 and an MBP(85-99)-specific T cell receptor. *J Autoimmun*. 2008;(31):399–407.
- 142 Binnig G, Quate CF, Gerber C. Atomic force microscope. *Phys Rev Lett*. 1986;56(9):930-933.
- 143 Binnig G, Rohrer H, Gerber C, Weibel E. Surface Studies by Scanning Tunneling Microscopy. *Phys. Rev. Lett*. 1982;(49):57.
- 144 Zavala G. Atomic force microscopy, a tool for characterization, synthesis and chemical processes. *Colloid Polym Sci*. 2008;(286):85–95.
- 145 Sugimoto Y, Pou P, Abe M, Jelinek P, Pérez R, Morita S, *et al*. Chemical identification of individual surface atoms by atomic force microscopy. *Nature*. 2007;(446):64–7.
- 146 Dufrêne YF. Using nanotechniques to explore microbial surfaces. *Nat Rev Microbiol*. 2004;(2):451–60.

- 147 Vock S, Hengst C, Wolf M, Tschulik K, Uhlemann M, Sasvári Z, *et al.* Magnetic vortex observation in FeCo nanowires by quantitative magnetic force microscopy. *Appl Phys Lett.* 2014;105.
- 148 Stannard A, Sweetman AM. A Considered Approach to Force Extraction from Dynamic Force Microscopy Measurements. In: Moriarty P, Gauthier S, editors. *Imaging Manip. Adsorbates Using Dyn. Force Microsc.*, Springer International Publishing. 2015; 63–79.
- 149 Ebner A, Wildling L, Zhu R, Rankl C, Haselgrübler T, Hinterdorfer P, *et al.* Functionalization of probe tips and supports for single-molecule recognition force microscopy. *Top Curr Chem.* 2008;(285):29–76.
- 150 Blanchette CD, Loui A, Ratto TV. Tip Functionalization: Applications to Chemical Force Spectroscopy. In: Noy A. *Handbook of Molecular Force Spectroscopy.* Boston: Springer; 2008. 185–203.
- 151 Limanskii AP. Functionalization of amino-modified probes for atomic force microscopy. *Biophysics.* 2006;(51):186–95.
- 152 Li M, Xiao X, Liu L, Xi N, Wang Y, Dong Z, *et al.* Atomic force microscopy study of the antigen-antibody binding force on patient cancer cells based on ROR1 fluorescence recognition. *J Mol Recognit.* 2013;(26):432–8.
- 153 Moraes AS *et al.* Evidências de Detecção do Herbicida Atrazina por Espectroscopia de Força Atômica: Uma Ferramenta Promissora para Sensoriamento Ambiental. *Acta Microsc.* 2015;(24):53–63.
- 154 Li M, Dang D, Liu L, Xi N, Wang Y. Imaging and Force Recognition of Single Molecular Behaviors Using Atomic Force Microscopy. *Sensors.* 2017;(17).
- 155 Puchner EM, Gaub HE. Force and function: probing proteins with AFM-based force spectroscopy. *Curr Opin Struct Biol.* 2009;(19):605–14.
- 156 Zimmermann JL, Nicolaus T, Neuert G, Blank K. Thiol-based, site-specific and covalent immobilization of biomolecules for single-molecule experiments. *Nat Protoc.* 2010;(5):975–85.
- 157 Trilling AK, Beekwilder J, Zuilhof H. Antibody orientation on biosensor surfaces: a minireview. *The Analyst.* 2013;(138):1619.
- 158 Starodub NF, Pirogova LV, Demchenko A, Nabok AV. Antibody immobilisation on the metal and silicon surfaces. The use of self-assembled layers and specific receptors. *Bioelectrochemistry Amst Neth.* 2005;(66):111–5.

- 159 Caruso F, Rodda E, Furlong DN. Orientational Aspects of Antibody Immobilization and Immunological Activity on Quartz Crystal Microbalance Electrodes. *J Colloid Interface Sci.* 1996;(178):104–15.
- 160 Ierich JCM, Oliveira GS, Vig ACA, Amarante AM, Franca EF, Leite FL, *et al.* A Computational Protein Structure Refinement of the Yeast Acetohydroxyacid Synthase. *J Braz Chem Soc.* 2015;(26):1702–9.
- 161 Shahin V, Ludwig Y, Schafer C, Nikova D, Oberleithner H. Glucocorticoids remodel nuclear envelope structure and permeability. *J Cell Sci.* 2005;(118):2881–9.
- 162 Butt H-J, Cappella B, Kappl M. Force measurements with the atomic force microscope: Technique, interpretation and applications. *Surf Sci Rep.* 2005;(59):1–152.
- 163 Hughes ML, Dougan L. The physics of pulling polyproteins: a review of single molecule force spectroscopy using the AFM to study protein unfolding. *Rep Prog Phys Phys Soc G B.* 2016;(79):076601.
- 164 Fisher TE, Marszalek PE, Fernandez JM. Stretching single molecules into novel conformations using the atomic force microscope. *Nat Struct Biol.* 2000;(7):719–24.
- 165 Hoffmann T, Dougan L. Single molecule force spectroscopy using polyproteins. *Chem Soc Rev.*, 2012;(41):4781-4796.
- 166 Ott W, Jobst MA, Schoeler C, Gaub HE, Nash MA. Single-molecule force spectroscopy on polyproteins and receptor–ligand complexes: The current toolbox. *J Struct Biol.* 2017;(197):3–12.
- 167 JPK Application Note:Structural Investigation of Single Biomolecules and Molecular Stretching Using Atomic Force Microscopy. JPK Instruments AG - all rights reserved (1-6).
- 168 Bizzarri A, Cannistraro S. Atomic Force Spectroscopy in Biological Complex Formation: Strategies and Perspectives. *J. Phys. Chem. B.* 2009, 113 (52):16449–16464
- 169 Bizzarri A, Cannistraro S. The application of atomic force spectroscopy to the study of biological complexes undergoing a biorecognition process. *Chem Soc Rev.* 2010;(39):734–49.
- 170 Dumitru AC, Herruzo ET, Rausell E, Ceña V, Garcia R. Unbinding forces and energies between a siRNA molecule and a dendrimer measured by force spectroscopy. *Nanoscale.* 2015;(7):20267–76.
- 171 Weisstein EW. Gamma Function. From MathWorld-A Wolfram Web Resource, 2004.

- 172 Thom HCS. A note on the gamma distribution. *Mon Weather Rev.* 1958;(86):117–22.
- 173 Mitnitski AB, Mogilner AJ, Rockwood K. Accumulation of Deficits as a Proxy Measure of Aging. *Sci. World J. Scientific World Journal.* 2001; 8;1:323-36.
- 174 Kataoka K, Harada A, Nagasaki Y. Block copolymer micelles for drug delivery: Design, characterization and biological significance. *Adv Drug Deliv Rev.* 2012; 64: 37–48.
- 175 Othmer HG, Dunbar SR, Alt W. Models of dispersal in biological systems. *J Math Biol.* 1988; 26: 263–298.
- 176 Isukapalli SS *et al.* Stochastic Response Surface Methods (SRSMs) for Uncertainty Propagation: Application to Environmental and Biological Systems. *Risk Anal.* 1998;18(3):351-63.
- 177 Wilkinson DJ. *Stochastic Modelling for Systems Biology.* 3rd ed. London: Chapman and Hall/CRC. 2011.
- 178 Edwards EM *et al.* Revisiting Lévy flight search patterns of wandering albatrosses, bumblebees and deer. *Nature.* 2007. 449(7165):1044-8.
- 179 Choi EJ, Dimitriadis EK. Cytochrome c Adsorption to Supported, Anionic Lipid Bilayers Studied via Atomic Force Microscopy. *Biophys J* 2004; 87: 3234–3241.
- 180 Perez-Jimenez R, Li J, Kosuri P, Sanchez-Romero I, Wiita AP, Rodriguez-Larrea D *et al.* Diversity of chemical mechanisms in thioredoxin catalysis revealed by single-molecule force spectroscopy. *Nat Struct Mol Biol* 2009; 16: 890–896.
- 181 Borlinhas F, Nogueira L, Brandão S, Nunes RG, Loureiro J, Ramos I, *et al.* Gamma distribution model in breast cancer diffusion-weighted imaging. 2015 IEEE 4th Port. Meet. Bioeng. Enbeng. 2015, 1–1.
- 182 Krieg M, Dunn AR, Goodman MB. Mechanical control of the sense of touch by β -spectrin. *Nat Cell Biol.* 2014;(16):224–33.
- 183 Girardin G, Huvier C, Delafosse D, Feaugas X. Correlation between dislocation organization and slip bands: TEM and AFM investigations in hydrogen-containing nickel and nickel–chromium. *Acta Mater* 2015;(91):141–51.
- 184 Goncharov VK, Kozadaev KV, Mikitchuk AP. Diagnostics Of The Monolayer Silver Nanostructures On A Solid Substrate Using The Bifactorial Analysis Of The Spr Band. *High Temp Mater Process Int Q High-Technol Plasma Process* 2014;18.

- 185 Faidra Angelero MG, Sabri A, Creasey R, Angelero P, Marlow M, Zelzer M. Surface-directed modulation of supramolecular gel properties. *Chem Commun.* 2016;(52):4298–300.
- 186 Williamson DF, Parker RA, Kendrick JS. The box plot: a simple visual method to interpret data. *Ann Intern Med.* 1989;(110):916–21.
- 187 Corbin EA, Kong F, Lim CT, King WP, Bashir R. Biophysical properties of human breast cancer cells measured using silicon MEMS resonators and atomic force microscopy. *Lab Chip.* 2015;(15):839–47.
- 188 Ivanov IE, Boyd CD, Newell PD, Schwartz ME, Turnbull L, Johnson MS, *et al.* Atomic force and super-resolution microscopy support a role for LapA as a cell-surface biofilm adhesin of *Pseudomonas fluorescens*. *Res Microbiol.* 2012;(163):685–91.
- 189 Migliorini E, Ban J, Greci G, Andolfi L, Pozzato A, Tormen M *et al.* Nanomechanics controls neuronal precursors adhesion and differentiation. *Biotechnol Bioeng* 2013; 110: 2301–2310.
- 190 Lanzicher T *et al.* AFM single-cell force spectroscopy links altered nuclear and cytoskeletal mechanics to defective cell adhesion in cardiac myocytes with a nuclear lamin mutation. *Nucleus.* 2015;6(5):394-407.
- 191 Sbaizero O *et al.* Analysis of long- and short-range contribution to adhesion work in cardiac fibroblasts: An atomic force microscopy study. *Materials Science and Engineering: C.* 2015 49: 217-224.
- 192 Milovanovic P, Potocnik J, Djonic D, Nikolic S, Zivkovic V, Djuric M *et al.* Age-related deterioration in trabecular bone mechanical properties at material level: nanoindentation study of the femoral neck in women by using AFM. *Exp Gerontol* 2012; 47: 154–159.
- 193 Walkiewicz MP *et al.* CENP-A octamers do not confer a reduction in nucleosome height by AFM | *Nature Structural & Molecular Biology.* *Nature Structural & Molecular Biology.* 2014. 21:2–3.
- 194 Christenson W, Yermolenko I, Plochberger B, Camacho-Alanis F, Ros A, Ugarova TP *et al.* Combined single cell AFM manipulation and TIRFM for probing the molecular stability of multilayer fibrinogen matrices. *Ultramicroscopy.* 2014; 136: 211–215.
- 195 Li Z, Persson H, Adolfsson K, Abariute L, T. Borgström M, Hessman D *et al.* Cellular traction forces: a useful parameter in cancer research. *Nanoscale.* 2017; 9: 19039–19044.

- 196 Mostowy S, Janel S, Forestier C, Roduit C, Kasas S, Pizarro-Cerdá J *et al.* A Role for Septins in the Interaction between the *Listeria monocytogenes* Invasion Protein InlB and the Met Receptor. *Biophys J.* 2011; 100: 1949–1959.
- 197 Baker MJ, Trevisan J, Bassan P, Bhargava R, Butler HJ, Dorling KM, *et al.* Using Fourier transform IR spectroscopy to analyze biological materials. *Nat Protoc.* 2014;(9):1771–91.
- 198 Pattnaik P. Surface plasmon resonance. *Appl Biochem Biotechnol.* 2005;(126):79–92.
- 199 Paddock SW, Eliceiri KW. Laser scanning confocal microscopy: history, applications, and related optical sectioning techniques. *Methods Mol Biol Clifton NJ.* 2014;(1075):9–47.
- 200 Duarte, LC *et al.* Aplicações de Microscopia Eletrônica de Varredura (MEV) e Sistema de Energia Dispersiva (EDS) no Estudo de Gemas: Exemplos brasileiros. *Pesquisas em Geociências.* 2003;30(2):3-15.
- 201 McCluskey MD. Local vibrational modes of impurities in semiconductors. *J Appl Phys* 2000;(87):3593–617.
- 202 Duygy DY *et al.* Fourier Transform Infrared (FT-IR) Spectroscopy for Biological Studies. *Journ of Scienc.* 2009;22(3)117-121.
- 203 Coates J. *Interpretation of Infrared Spectra, A Practical Approach.* New Jersey: John Wiley & Sons, Ltd. 2006.
- 204 Paddock SW. Principles and practices of laser scanning confocal microscopy. *Mol Biotechnol.* 2000;(16):127–49.
- 205 Herman B, Lemasters JJ. *Optical Microscopy: Emerging Methods and Applications.* London: academic press limited. 1993.
- 206 Periasamy A. *Methods in Cellular Imaging.* New York: Osford – American Physiological Society. 2001.
- 207 Haupt BJ, Pelling AE, Horton MA. Integrated confocal and scanning probe microscopy for biomedical research. *ScientificWorldJournal.* 2006;(6):1609–18.
- 208 Li A, Lim TS, Shi H, Yin J, Tan SJ, Li Z, *et al.* Molecular Mechanistic Insights into the Endothelial Receptor Mediated Cytoadherence of *Plasmodium falciparum*-Infected Erythrocytes. *PLoS ONE.* 2011;6.
- 209 Bogner A, Jouneau P-H, Thollet G, Basset D, Gauthier C. A history of scanning electron microscopy developments: towards “wet-STEM” imaging. *Micron Oxf Engl.* 1993 2007;(38):390–401.

- 210 Goldstein JI, Newbury DE, Echlin P, Joy DC, Jr ADR, Lyman CE, *et al.* Electron-Specimen Interactions. Scanning Electron Microsc. X-Ray Microanal., Springer US. 1992, 69–147.
- 211 Goldstein JI *et al.* X-Ray Spectral Measurement: WDS and EDS. Scanning Electron Microscopy and X-ray Microanal. Springer US. 2003. 297-353.
- 212 Chen L, Xu J, Chen J. Applications of scanning electron microscopy in earth sciences. *Sci China Earth Sci.* 2015;(58):1768–78.
- 213 Orloff J. Handbook of Charged Particle Optics, 2nd Edition. Boca Raton: CRC Press. 2009.
- 214 Mukhopadhyay A. Measurement of magnetic hysteresis loops in continuous and patterned ferromagnetic nanostructures by static magneto-optical kerr effect magnetometer. Guwahati: Satyendra Nath Bose National Centre for Basic Sciences; 2015.
- 215 Varanda LM, Miranda MT. Solid-phase peptide synthesis at elevated temperatures: a search for and optimized synthesis condition of unsulfated cholecystokinin-12. *J Pept Res Off J Am Pept Soc.* 1997;(50):102–8.
- 216 Remuzgo C, Oewel TS, Daffre S, Lopes TRS, Dyszy FH, Schreier S, *et al.* Chemical synthesis, structure-activity relationship, and properties of shepherin I: a fungicidal peptide enriched in glycine-glycine-histidine motifs. *Amino Acids.* 2014;(46):2573–86.
- 217 Machado A, Fázio MA, Miranda A, Daffre S, Machini MT. Synthesis and properties of cyclic gomesin and analogues. *J Pept Sci Off Publ Eur Pept Soc.* 2012;(18):588–98.
- 218 Loffredo C, Assunção NA, Gerhardt J, Miranda MTM. Microwave-assisted solid-phase peptide synthesis at 60 degrees C: alternative conditions with low enantiomerization. *J Pept Sci Off Publ Eur Pept Soc.* 2009;(15):808–17.
- 219 Lingyan L *et al.* In Situ Single-Molecule Detection of Antibody–Antigen Binding by Tapping-Mode Atomic Force Microscopy; *Anal. Chem.* 2002, 74 (23), 6017–6022.
- 220 Silva ACN, Deda DK, da Róz AL, Prado RA, Carvalho CC, Viviani V, *et al.* Nanobiosensors Based on Chemically Modified AFM Probes: A Useful Tool for Metsulfuron-Methyl Detection. *Sensors.* 2013;(13):1477–89.

- 221 Deda DK, Pereira BBS, Bueno CC, Silva AN da, Ribeiro GA, Amarante AM, *et al.* The use of functionalized AFM tips as molecular sensors in the detection of pesticides. *Mater Res.* 2013;(16):683–7.
- 222 Hinterdorfer P, Dufrêne YF. Detection and localization of single molecular recognition events using atomic force microscopy. *Nat Methods.* 2006;(3):347–55.
- 223 Hinterdorfer P, Baumgartner W, Gruber HJ, Schilcher K, Schindler H. Detection and localization of individual antibody-antigen recognition events by atomic force microscopy. *Proc Natl Acad Sci U S A.* 1996;(93):3477–81.
- 224 Okuda-Shinagawa NM, Moskalenko YE, Junqueira HC, Baptista MS, Marques CM, Machini MT. Fluorescent and Photosensitizing Conjugates of Cell-Penetrating Peptide TAT(47-57): Design, Microwave-Assisted Synthesis at 60 °C, and Properties. *ACS Omega.* 2017;(2):8156–66.
- 225 Arndt P, Garratty G. Evaluation of the optimal incubation temperature for detecting certain IgG antibodies with potential clinical significance. *Transfusion (Paris).* 1988;(28):210–3.
- 226 Sidney J, Southwood S, Moore C, Oseroff C, Pinilla C, Grey HM, *et al.* Measurement of MHC/peptide interactions by gel filtration or monoclonal antibody capture. *Curr Protoc Immunol Ed John E Coligan Al.* 2013; (18):18.3.
- 227 Majoul N, Aouida S, Bessaïs B. Progress of porous silicon APTES-functionalization by FTIR investigations. *Applied Surface Science.* 2015; (331):388–391.
- 228 Öztürk T *et al.* Synthesis and Characterization of Poly(methyl methacrylate-block-ethylene glycol-block-methyl methacrylate) Block Copolymers by Reversible Addition-Fragmentation Chain Transfer Polymerization. *Journal of Macromolecular Science, Part A.* 2010; (48):65-70.
- 229] Miller LM, Bourassa MW, Smith RJ. FTIR spectroscopic imaging of protein aggregation in living cells. *Biochim Biophys Acta.* 2013; (1828): 2339–2346.
- 230 Dazzi A, Prater CB. AFM-IR: Technology and Applications in Nanoscale Infrared Spectroscopy and Chemical Imaging. *Chem Rev* 2017; (117): 5146–5173.
- 231 Nguyen HH *et al.* Surface plasmon resonance: a versatile technique for biosensor applications. *Sensors (Basel).* 2015;15(5):10481-510.
- 232 Lamprou DA, Smith JR, Nevell TG, Barbu E, Willis CR, Tsibouklis J. Self-assembled structures of alkanethiols on gold-coated cantilever tips and substrates for

atomic force microscopy: Molecular organisation and conditions for reproducible deposition. *Appl Surf Sci.* 2010;(256):1961–8.

233 Lau PCY, Dutcher JR, Beveridge TJ, Lam JS. Absolute Quantitation of Bacterial Biofilm Adhesion and Viscoelasticity by Microbead Force Spectroscopy. *Biophys J.* 2009;(96):2935–48.

234 Sirghi L, Kylián O, Gilliland D, Ceccone G, Rossi F. Cleaning and Hydrophilization of Atomic Force Microscopy Silicon Probes *J Phys Chem B.* 2006;110(51):25975-81.

235 Goldstein JI et al. *Scanning Electron Microscopy and X-Ray Microanalysis.* 3rd edition. Springer US. 2003.

236 Klawiter EC, Piccio L, Lyons J-A, Mikesell R, O'Connor KC, Cross AH. Intrathecal Anti-MOG Antibody Production is Elevated in Multiple Sclerosis. *Arch Neurol.* 2010;(67):1102–8.

237 Zeman AZ, Keir G, Luxton R, Thompson EJ. Serum oligoclonal IgG is a common and persistent finding in multiple sclerosis, and has a systemic source. *QJM Mon J Assoc Physicians.* 1996;(89):187–93.

238 Arrambide G, Tintore M, Espejo C, Auger C, Castillo M, Ríó J, *et al.* The value of oligoclonal bands in the multiple sclerosis diagnostic criteria. *Brain J Neurol.* 2018;(141):1075–84.

239 Domingues RB, Fernandes GBP, Leite FBV de M, Tilbery CP, Thomaz RB, Silva GS, *et al.* The cerebrospinal fluid in multiple sclerosis: far beyond the bands. *Einstein. São Paulo* 2017;(15):100–4.

240 Breithaupt C, Schubart A, Zander H, Skerra A, Huber R, Linington C, *et al.* Structural insights into the antigenicity of myelin oligodendrocyte glycoprotein. *Proc Natl Acad Sci.* 2003;(100):9446–51.

241 Huang J, MacKerell AD. CHARMM36 all-atom additive protein force field: validation based on comparison to NMR data. *J Comput Chem.* 2013;(34):2135–45.

242 Phillips JC, Braun R, Wang W, Gumbart J, Tajkhorshid E, Villa E, *et al.* Scalable molecular dynamics with NAMD. *J Comput Chem.* 2005;(26):1781–802.