

UNIVERSIDADE DE SÃO PAULO

FACULDADE DE CIÊNCIAS FARMACÊUTICAS DE RIBEIRÃO PRETO



Phaeurus antarcticus and its endophytic fungi: Chemical diversity of a hidden pharmacy underneath the Antarctic Ocean

Phaeurus antarcticus e seus fungos endofíticos: Diversidade química de uma farmácia oculta sob o oceano Antártico

Gustavo Souza dos Santos

Ribeirão Preto 2022

GUSTAVO SOUZA DOS SANTOS

Phaeurus antarcticus and its endophytic fungi: Chemical diversity of a hidden pharmacy underneath the Antarctic Ocean

Phaeurus antarcticus e seus fungos endofíticos: Diversidade química de uma farmácia oculta sob o oceano Antártico

Doctoral thesis presented to the Graduate Program of School of Pharmaceutical Sciences of Ribeirão Preto/USP for the degree of Doctor in Sciences.

Concentration Area: Natural and Synthetic Products

Supervisor: Professor Hosana Maria Debonsi, PhD

Co supervisor: Professor RuAngelie Edrada-Ebel, PhD

Versão corrigida da Tese de Doutorado apresentada ao Programa de Pós-graduação em Ciências Farmacêuticas em 29/11/2022. A versão original encontra-se disponível na Faculdade de Ciências Farmacêuticas de Ribeirão Preto/USP.

Ribeirão Preto 2022 I AUTHORIZE THE WHOLE OR PARTIAL REPRODUCTION /DISCLOSURE OF THIS WORK, BY ANY CONVENTIONAL OR ELECTRONIC MEANS, TO STUDY AND RESEARCH RELATED PURPOSES AS LONG THE SOURCE IS CITED.

Santos, Gustavo Souza dos

Phaeurus antarcticus and its endophytic fungi: Chemical diversity of a hidden pharmacy underneath the Antarctic Ocean Ribeirão Preto, 2022.

367 p.; 30 cm.

Doctoral thesis presented to the Graduate Program of School of Pharmaceutical Sciences of Ribeirão Preto/USP for the degree of Doctor in Sciences. Concentration Area: Natural and Synthetic Products

Supervisor: Hosana Maria Debonsi, PhD

Co-supervisor: RuAngelie Edrada-Ebel, PhD

Marine natural products 2. Antarctic fungi 3. Antarctic seaweed 4.
 Photoprotection 5. Antimicrobials 6. Neglected diseases 7.
 Metabolomics 8. Immunomodulatory activity.

APPROVAL PAGE

Name: Gustavo Souza dos Santos

Title: *Phaeurus antarcticus* and its endophytic fungi: Chemical diversity of a hidden pharmacy underneath the Antarctic Ocean.

Doctoral thesis presented to the Graduate Program of School of Pharmaceutical Sciences of Ribeirão Preto/USP for the degree of Doctor in Sciences.

Concentration Area: Natural and Synthetic Products

Supervisor: Hosana Maria Debonsi, PhD

Co-supervisor: RuAngelie Edrada-Ebel, PhD

Approved on:	
Examin	ers
Prof. Dr	
Institution:	Signature:
Prof. Dr	
Institution:	Signature:
Prof. Dr	
Institution:	Signature:
Prof. Dr	
Institution:	Signature:

Dedico este trabalho aos meus pais Rainy e Geilson, que sempre tomaram minhas lutas como suas e nunca deixaram de acreditar em mim.

AGRADECIMENTOS/ACKNOWLEDGEMENTS

Aos meus pais, Rainy e Geilson e a minha irmã Raissa, obrigado pelo amor incondicional e por dividir os fardos mais pesados durante essa caminhada.

Aos meus avós, Francisca, Raimundo, Vitor e Julia por serem os alicerces da minha criação.

As minhas tias, Izabel, Aparecida, Socorro, Rosa, Carol, Rosário, Julia e Geises pelo carinho e cuidado.

Ao meu namorado, Guilherme Rocha, por todo o amor, compreensão, incentivo e paciência. Obrigado por ser meu porto seguro.

Aos meus sogros, Mauro e Lindaura e família pelo acolhimento e carinho.

Aos meus primos Ethel, Ennio, Rosainhe e Thais por fazerem parte dessa jornada.

A minha orientadora, Prof. Dra. Hosana Maria Debonsi, pelo voto de confiança e pela imensa troca de conhecimentos durante todos esses anos. Obrigado pelo acolhimento, carinho e oportunidade de realizar este trabalho.

Ao Prof. Dr. Pio Colepicolo pela oportunidade de fazer parte da Missão Antartica 36.

Aos professores colaboradores, obrigado pela atenção e dedicação dispensada, as quais foram essenciais para a finalização deste trabalho.

Aos professores e alunos do Núcleo de Pesquisas em Produtos Naturais e Sintéticos, por proporcionarem um ambiente de aprendizado e descontração.

Aos técnicos Izabel Cristina, José Carlos Tomaz, Karina Castro, Vinícius Palaretti, e em especial à Ludmila Carvalho pelas contribuições na realização deste trabalho.

A FCFRP-USP, especialmente ao Programa de Pós-graduação em Ciências Farmacêuticas.

Aos meus colegas de laboratório Ana Carolina, Márcia e Vitor pelo companheirismo e aprendizado.

Aos meus amigos e companheiros de pós-graduação e experimentos, Leandro, Allana, Isadora e Nathália por toda a dedicação.

Aos amigos de Ribeirão Preto, Thaiz, Péricles, Victor, Valdeline, Jennyfer, Adriany, Karen, Gabriel, Pedro, Grilo, Mario, Natalia, Neto e Victoria. Obrigado por se tornarem uma segunda família durante todos esses anos, e por compartilharem as alegrias, as decepções, e em ambos os casos, os litrões.

Aos meus amigos, Maura, Roberta, Higor, Soraia, Gabriel, Taynara, Elen, Sara, Lunara e Tâmille por sempre estarem presentes mesmo separados pela distância física.

Aos meus companheiros da OPERANTAR 36, Angélica R. Soares, Bruna Pacheco, Carlos Menk, Maria Beatriz Barreto e Rodrigo Mendes pela incrível aventura no continente gelado.

A todos os professores que fizeram parte da minha formação, em especial aos queridos, Adrian Pohlit, Ana Guerra, Ângela Lucia, Breno Schumaher, Camila Fabbri, Cynthia Alcantara, Cristiane Carvalho, Luciana Lopes, Marcio Martinez, Marcia Alves, Margareth Alencar, Tiago Pereira e Valdeana Linard.

To my co-supervisor Dr. RuAngelie Edrada-Ebel (Strathclyde Institute of Pharmacy and Biomedical Sciences), thank you for the opportunity to develop this work, learn from you, and for sharing amazing experiences while in Scotland. I am grateful to have worked with you and your research group.

To Dr. Louise Young for sharing her expertise in bioassays.

To my lab mate Elizabeth for the guidance and friendship during my project development at University of Strathclyde.

Ao Conselho Nacional de Desenvolvimento Científico e Tecnológico – CNPQ, pelo financiamento da bolsa de doutorado (Processo 140811/2018-4).

A Coordenação de Aperfeiçoamento de Pessoal de Nível Superior – CAPES, pelo financiamento da bolsa de doutorado sanduiche (88887.572104/2020-00).

O presente trabalho foi realizado com apoio da Coordenação de Aperfeiçoamento de Pessoal de Nível Superior – Brasil (CAPES) – Código de Financiamento 001.

"Man is the most insane species. He worships an invisible God and destroys a visible nature. Unaware that this nature he's destroying is this God he's worshiping."

- Hubert Reeve

RESUMO

SANTOS, G. S. *Phaeurus antarcticus* e seus fungos endofíticos: Diversidade química de uma farmácia oculta sob o oceano Antártico. 2022. 367f. Tese (Doutorado). Faculdade de Ciências Farmacêuticas de Ribeirão Preto – Universidade de São Paulo, Ribeirão Preto, 2022.

Capítulo I: O perfil químico e as atividades antibacteriana, antiparasitária e inseticida da alga P. antarcticus foram investigados. O extrato hexânico (HX) e suas frações exibiram atividade bacteriostática e bactericida seletiva contra Staphylococcus. aureus. As frações HX-FC e HX-FD apresentaram atividade antiparasitária frente a Leishmania amazonensis e Neospora caninum. O extrato HX e a fração HX-FC apresentaram potencial inseticida frente a larvas de Aedes aegypti. Através de análises em CG-EM, e combinação de análises estatísticas multivariadas (OPLS-DA) com a construção de redes moleculares (GNPS) foi possível anotar terpenos, esteroides ácidos graxos e álcoois nas frações bioativas. Capítulo II: Utilizando técnicas metabolômicas (CLUE-EM/RMN¹H) combinadas a análises estatísticas multivariadas (PCA, OPLS-DA) a variabilidade espacial de metabólitos secundários da alga P. antarcticus e a atividade antibiofilme dos extratos e frações foram avaliados frente a S. aureus resistente à meticilina e oxacilina (MRSA). Os resultados revelaram que as mudanças no perfil químico não foram sítio específicas e podem estar relacionadas a diferentes estágios do ciclo de vida das algas coletadas. Porfirinas, terpenos e carotenoides foram anotados como sustâncias discriminantes. O ácido linoleico (1) e o fucosterol (2) foram isolados das frações bioativas e apresentaram atividade antibiofilme frente a MRSA. Capítulo III: O extrato e frações do fungo purpurogenum foram submetidos a avaliação Penicillium das atividades antibacteriana, antiparasitária, imunomoduladora e fotoprotetora. O fracionamento bioguiado levou ao isolamento de seis policetídeos: 5,6,8-triidroxi-4-(1'-hidroxietil) isocumarina (3), aspergilumarina A (4), aspergilumarina B (5) berkeleyacetal C (6), sescandelina (7) e vermistatina (8). O potencial fotoprotetor, imunomodulador e leishmanicida das substâncias foi avaliado. As substâncias 3, 4, 7 e 8 apresentaram fotoestabilidade e atividade fotoprotetora frente a radiação UVB-UVA. As substâncias 3, 6, 7 e 8 apresentaram potencial imunomodulador, reduzindo a produção de espécies reativas de oxigênio em neutrófilos humanos estimulados pelo PMA sem efeitos citotóxicos. A substância 6 apresentou atividade leishmanicida frente a amastigotas de L. amazonensis. Capítulo IV: O perfil químico (CLUE-EM/ RMN¹H) e a atividade antibiofilme frente a MRSA de 21 linhagens fúngicas foram investigados. O fracionamento do extrato do fungo Epicoccum nigrum, levou ao isolamento das epicozarinas A – B (9-10) e C (12) e de um terpeno (11) inédito. As substâncias 9 – 12 exibiram atividade antibiofilme contra MRSA. Dentre estas, a substância 11 apresentou maior inibição da formação de pré-biofilme e ruptura do biofilme formado. Diante do contexto global, este trabalho buscou a obtenção de novos produtos biotecnológicos, considerando os Objetivos do Desenvolvimento Sustentável das Nações Unidas. Os resultados obtidos fornecem novas informações quimiotaxonômicas e potencial biotecnológico dos organismos marinhos da Antártica como produtores de biomoléculas de interesse farmacêutico, enfatizando a importância da bioprospecção para a conservação de organismos ainda pouco conhecidos.

Palavras-chave: Produtos naturais marinhos, Fungos antárticos, Macroalgas antárticas, Fotoproteção, Antimicrobianos, Doenças negligenciadas, Atividade imunomoduladora, Metabolômica

ABSTRACT

SANTOS, G. S. *Phaeurus antarcticus* and its endophytic fungi: Chemical diversity of a hidden pharmacy underneath the Antarctic Ocean. 2022. 367f. Thesis (Doctoral). Faculdade de Ciências Farmacêuticas de Ribeirão Preto – Universidade de São Paulo, Ribeirão Preto, 2022.

Chapter I: The chemical profile and antibacterial, antiparasitic, and insecticidal activities of the seaweed *P. antarcticus* were investigated. The hexane extract (HX) and its fractions exhibited selective antibacterial activity against Staphylococcus aureus. Fractions HX-FC and HX-FD showed the highest antiparasitic activity against Leishmania amazonensis and Neospora caninum. The HX extract and the HX-FC fraction presented insecticidal potential against Aedes aegypti larvae. Through GC-MS analysis and the combination of multivariate statistical analyses (OPLS-DA) and the construction of molecular networks (GNPS), it was possible to annotate terpenes, steroids, fatty acids, and alcohols in the bioactive fractions. Chapter II: Using metabolomics techniques (UPLC-HRMS/1H NMR) and multivariate statistical analysis (PCA, OPLS-DA), the spatial variability of secondary metabolites of the seaweed P. antarcticus and the antibiofilm activity of extracts and fractions were evaluated against S. aureus resistant to methicillin-oxacillin (MRSA). The results revealed that the changes in the chemical profile were not site-specific and could be related to different life cycle stages of the collected algae. Porphyrins, terpenes, and carotenoids were annotated as discriminating compounds. Linoleic acid (1) and fucosterol (2) were isolated from the bioactive fractions and showed antibiofilm activity against MRSA. **Chapter III:** The extract and fractions of the fungus *Penicillium purpurogenum* were evaluated for antibacterial, antiparasitic, immunomodulatory, and photoprotective activities. The bioquided fractionation led to the isolation of six polyketides: 5,6,8trihydroxy-4-(1'-hydroxyethyl) isocoumarin (3), aspergillumarin A (4), aspergillumarin B (5), Berkelevacetal C (6), sescandelin (7), and vermistatin (8). Compounds 3, 4, 7, and 8 presented photostability and photoprotective activities against UVB-UVA. Compounds 3, 6, 7, and 8 presented immunomodulatory activity by down-regulating the production of reactive oxygen species in PMA-stimulated human neutrophils without cytotoxic effects. Compound 6 presented leishmanicidal activity against amastigotes of *L. amazonensis*. Chapter IV: The chemical profile (UPLC-HRMS/¹H NMR) and antibiofilm activity against MRSA of 21 fungal strains were investigated. Among them, the fractionation of the extract of the fungus *Epicoccum nigrum* led to the isolation of epicorazines A-B (9-10) and C (12) as well as a new terpenoid (11). Compound 9–12 exhibited antibiofilm activity against MRSA. Among these, compound 11 showed the highest inhibition of pre-biofilm formation and rupture of the formed biofilm. Given the global context, this work sought to obtain new biotechnological products, considering the United Nations Sustainable Development Goals. The results obtained provide new information about the chemotaxonomy and biotechnological potential of Antarctic marine organisms as producers of biomolecules of pharmaceutical interest, emphasizing the importance of bioprospecting for the conservation of understudied organisms.

Keywords: Marine natural products, Antarctic fungi, Antarctic seaweed, Photoprotection, Antimicrobials, Neglected diseases, Immunomodulatory activity, Metabolomics

LIST OF FIGURES

Figure 1. Antarctic glacier near the Comandante Ferraz Brazilian Antarctic Base1
Figure 2 Antarctic fauna. A – Pygoscelis antarcticus; B – Pygoscelis adeliae; C –
Pygoscelis papua; D – Macronectes giganteus; E – Hydrurga leptonyx; F – Mirounga
leonina2
Figure 3. Oceanographic ship Prof. Wladimir Besnard4
Figure 4. Brazil in Antarctica. A-B – Old Antarctic base; C-D – New Antarctic base; E
– Polar ship Almirante Maximiano (H-41); F – Oceanographic ship Ary Rongel5
Figure 5. Compounds isolated from Antarctic seaweeds
Figure 6. Compounds isolated from Antarctic-derived fungi11
Figure 7. The seaweed <i>Phaeurus antarcticus</i> 12
Figure 8. Multivariate models and their applicability. 14
Figure 9. OPLS-DA scores scatter plot of GC-MS data of <i>P. antarcticus</i> fractions and
bioactivity data of the cytotoxicity to human fibroblasts
Figure 10. OPLS-DA loadings scatter plot of GC-MS data of <i>P. antarcticus</i> fractions
and bioactivity data of the antibacterial assays against S. aureus. The detected
compounds are colored according to their retention time in minutes
Figure 11. GC-MS molecular networking showing the cluster containing the
discriminant IDs pointed by the OPLS-DA (circled in red) as the bioactive constituents
in fraction HX-FB
Figure 12. OPLS-DA scores scatter plot of GC-MS data of <i>P. antarcticus</i> fractions and
antibacterial activity against <i>S. aureus</i>
Figure 13. OPLS-DA scores scatter plot of GC-MS data of <i>P. antarcticus</i> fractions and
antibacterial activity against <i>S. aureus</i>
Figure 14. GC-MS molecular networking showing the cluster containing the
discriminant IDs pointed by the OPLS-DA (circled in red) as the bioactive constituents
in fraction HX-FD
Figure 15. OPLS-DA scores scatter plot of GC-MS data of <i>P. antarcticus</i> fractions and
antiparasitic activity40
Figure 16. OPLS-DA scores scatter plot of GC-MS data of <i>P. antarcticus</i> fractions and
antiparasitic activity40
Figure 17. GC-MS molecular networking shows the cluster containing the discriminant
IDs pointed by the OPLS-DA (circled in red) as the bioactive constituents in fraction
HX-FC41

Figure 18. OPLS-DA scores scatter plot of GC-MS data of *P. antarcticus* fractions and Figure 19. OPLS-DA loadings scatter plot of GC-MS data of *P. antarcticus* fractions and larvicidal activity.43 Figure 20. GC-MS molecular networking shows the cluster containing the discriminant Figure 21. Collection site of *P. antarcticus* in the year of 2015. A – Antarctica; B – South Shetland Islands in Antarctica Peninsula; C – Expansion of South Shetland Figure 22. PCA score scatter plot of the ¹H NMR spectral data of *P. antarcticus* crude extracts collected in 2015; R2: 1.0, Q2: 0.99......58 Figure 23. PCA loadings scatter plot of the ¹H NMR spectral of *P. antarcticus* crude extracts collected in 2015; Variables are colored by their chemical shifts (ppm); R2: Figure 24. PCA score scatter plot of the HRMS spectral data of *P. antarcticus* crude extracts collected in 2015; R2: 1.0, Q2: 0.99......59 Figure 25. PCA loadings scatter plot of the HRMS data of *P. antarcticus* crude extracts collected in 2015; Variables are colored by their m/z values; R2: 1.0, Q2: 0.99......60 Figure 26. Putative structures of metabolites pointed out as discriminant features in samples collected in Arctowski......61 Figure 27. Putative structures of metabolites pointed out as discriminant features in samples collected in Arctowski.....61 Figure 28. Putative structures of metabolites pointed out as discriminant features in Figure 29. Putative structures of metabolites pointed out as discriminant features in samples collected in Halfmoon Island (2015).....63 Figure 30. Collection site of *P. antarcticus* in the year of 2015. A – Antarctica; B – South Shetland Islands in Antarctica Peninsula; **C** – Greenwich Island; **D** – Demay Point (green pin), Vaureal Point (yellow pin), Penguin Island (purple pin)......64 Figure 31. PCA score scatter plot of the ¹H NMR spectral data of *P. antarcticus* crude extracts collected in 2017/2018; R2: 1.0, Q2: 0.99.....65 Figure 32. PCA loadings scatter plot of the ¹H NMR spectral data of *P. antarcticus* crude extracts collected in 2017/2018; Variables are colored according to their chemical shifts (ppm) R2: 1.0, Q2: 0.99.65

Figure 33. PCA score scatter plot of the HRMS data of *P. antarcticus* crude extracts collected in 2018; R2: 1.0, Q2: 0.99.66 Figure 34. PCA loadings scatter plot of the HRMS data of *P. antarcticus* crude extracts collected in 2015; Variables are colored by their m/z values; R2: 1.0, Q2: 0.99......67 Figure 35. Putative structures of metabolites pointed out as discriminant features in samples collected in Greenwich Island (2018).68 Figure 36. Putative structures of metabolites pointed as discriminant features in samples collected in Demay Point (2018).68 Figure 37. PCA score scatter plot of the ¹H NMR spectral data of *P. antarcticus* crude Figure 38. PCA loadings scatter plot of the ¹H NMR spectral data of *P. antarcticus* crude extracts collected on Greenwich Island in 2015 and 2018. Variables are colored according to their chemical shift (ppm).....70 Figure 39. PCA score scatter plot of the HRMS spectral data of *P. antarcticus* crude extracts collected on Greenwich Island in 2015 and 2018. R2 0.78; Q2 0.44.71 Figure 40. PCA loadings scatter plot of the HRMS spectral data of *P. antarcticus* crude extracts collected on Greenwich Island in 2015 and 2018. Variables are colored Figure 41. PCA score scatter plot of the HRMS spectral data of *P. antarcticus* crude extracts collected in 2015 and 2018. R2 0.81; Q2 0.48......72 Figure 42. Heatmap showing the top 100 variables (*m/z*) in *P. antarcticus* collected in Figure 43. OPLS-DA score scatter plot of the proton NMR spectral data of P. antarcticus crude extracts according to the antibiofilm activity against S. aureus; R2 Figure 44. OPLS-DA score scatter plot of the proton NMR spectral data of P. antarcticus crude extracts according to the antibiofilm activity against S. aureus; R2 Figure 45. OPLS-DA score scatter plot of the HRMS spectral data of P. antarcticus crude extracts according to the antibiofilm activity against *S. aureus*; R2 0.99 Q2 0.90. Figure 46. OPLS-DA S-plot of the HRMS spectral data of *P. antarcticus* crude extracts

Figure 47. OPLS-DA score scatter plot of the ¹H NMR spectral data of *P. antarcticus* fractions according to the antibiofilm activity against S. aureus. R2 0.99 Q2 0.92....84 Figure 48. OPLS-DA loadings scatter of the ¹H NMR spectral data of *P. antarcticus* fractions according to the antibiofilm activity against S. aureus; R2 0.99 Q2 0.92....85 Figure 49. ¹H NMR spectra of fractions Fr4 (pink), Fr5 (navy blue), Fr6 (green) measured in DMSO-d6 (400 MHz).....85 Figure 50. OPLS-DA score scatter plot of the HRMS data of *P. antarcticus* fractions according to the antibiofilm activity against S. aureus; R2 0.98 Q2 0.69......86 Figure 51. OPLS-DA loadings scatter plot of the HRMS data of *P. antarcticus* fractions according to the antibiofilm activity against S. aureus; R2 0.99 Q2 0.77......87 Figure 52. Putative structures of compound pointed out as discriminant features in Figure 53. Overview of the isolation of compounds 1 and 2 from the MeOH extract of Figure 54. ¹H NMR spectrum of compound 2 in DMSO-d₆ measured at 400 MHz...90 Figure 55. COSY correlation spectrum of compound 1 in DMSO-d₆ measured at 400 MHz......91 Figure 56. ¹H NMR spectrum of compound 2 in acetone-d₆ measured at 400 MHz. 93 Figure 57. HSQC correlation spectrum of compound 2 in acetone-d₆ (400 MHz). ...94 Figure 58. ¹³C (J-mode) spectrum of compound 2 in acetone-d₆ (400 MHz)......94 Figure 60. Structures of dolastatin 10, and its synthetic analogs currently available in the market......102 Figure 61. Anti-inflammatory compounds isolated from Antarctic-derived marine fungi. Figure 62. Approved drugs derived from fungi......105 Figure 63. Antileishmanial compound isolated from the marine fungus *Paecilomyces* sp. and the reference drug amphotericin B.106 Figure 64. Antiviral compounds isolated from *P. purpurogenum* JS03-21.....107 Figure 65. Insecticidal and cytotoxic compounds isolated from *P. purpurogenum*. 108 Figure 66. Collection sites of the seaweed Phaeurus antarcticus in the Antarctic Peninsula; A – Antarctic Peninsula; B – South Shetlands Archipelago; C – Coppermine

Figure 67. Isolation procedures to the isolation of endophytic fungi from *P. antarcticus*. Figure 69. Extraction procedures to evaluate the changes in secondary metabolites production......117 Figure 70. Macro morphology of fungi endophytes isolated from seaweed P. antarcticus in potato agar. A) P. purpurogenum; B) P. vitícola; C) Cladosporium sp.; Figure 71. Crude extracts absorption spectra in the UVA-UVB range and photostability assay based on the absorption spectrum after UVA irradiation (dotted line) or not Figure 72. Graphical representation of *P. purpurogenum* crude extracts in natural seawater – potato dextrose media (NSW-PDB) and artificial seawater – potato dextrose media (ASW-PDB)......133 Figure 73. P. purpurogenum cultures in ASW-PD media; A – B and D. Colony macro Figure 74. Comparative TLC of crude extracts obtained in NSW and ASW potato dextrose media in 7 days cultivation.....134 Figure 75. Comparative TLC of crude extracts obtained in NSW and ASW potato dextrose media in 14 days cultivation......135 Figure 76. Comparative TLC of crude extracts obtained in NSW and ASW potato dextrose media in 21 days cultivation.....136 Figure 77. A,C,E – HPLC-DAD profiles of the EtOAc extracts from NSW-PDB media in 7, 14 and 21 days respectively. **B,D,F** – HPLC-DAD profiles of the EtOAc crude Figure 78. ¹H NMR profiles of *P. purpurogenum* crude extracts in DMSO-d6 (400 MHz) Figure 79. PLSDA scatter plot of the proton NMR spectral data of *P. purpurogenum* extracts obtained from PDB-ASW (pink) and PDB-NSW (green) media incubated for 7, 14 and 21 days. R2= 0.99; Q2= 0.90.140 Figure 80. PLSDA loadings plot of proton NMR spectral data of *P. purpurogenum* extracts obtained from PDB-ASW and PDB-NSW media incubated for 7, 14 and 21

days.....141

vii

Figure 90. P. purpurogenum extracts absorption spectra in the UVA-UVB region. A) Figure 91. Absorption spectra in the UVA-UVB region and photostability assay based on the absorption spectrum after UV-A irradiation (dotted line) or not (continuous line) Figure 92. Inhibition of ROS in human neutrophils treated with *P. purpurogenum* EtOAc extracts 7 days; NSW-PDB (green-left); ASW-PDB (pink-right). Results are expressed mean values ± S.D from two independent experiments in triplicate......152 Figure 93. Inhibition of ROS in human neutrophils treated with P. purpurogenum EtOAc extracts 14 days; NSW-PDB (green-left); ASW-PDB (pink-right). Results are expressed mean values ± S.D from two independent experiments in triplicate......152 Figure 94. Inhibition of ROS in human neutrophils treated with *P. purpurogenum* EtOAc extracts 21 days NSW-PDB (green-left); ASW-PDB (pink-right). Results are expressed mean values ± S.D from two independent experiments in triplicate......153 Figure 95. Schematic representation of VLC fractionation of *P. purpurogenum* crude extract. Numbers represent the yield in grams. Orange arrows and boxes summarize the bioactivity results of each fraction......155 Figure 96. Chromatographic profile of fractions A-I analyzed by HPLC-DAD. Fraction A (A), Fraction (B), Fraction (C) and Fraction D (D) chromatograms in 225 nm. Fraction E (E), Fraction F (F), Fraction G (G), Fractio H (H) and Fraction I (I) chromatograms Figure 98. Chromatogram of subfraction FrE-6, elution in a polarity gradient: 0 min -60% H₂O + 40% CH₃CN; 7 min - 50% H₂O + 50% CH₃CN; 10 min -12 min 100% CH₃CN; 16 min 40 % CH₃CN, monitoring in 329 nm. Compound 3 (Rt = 5.55 min) and **4** (Rt= 10.43 min)......161 Figure 99. ¹H NMR spectra of compound 3 in DMSO-d₆ measured at 500 MHz ... 162 Figure 100. COSY correlation spectrum of compound 3 in DMSO-d₆, 500 MHz ... 163 Figure 101. HSQC spectra of compound 3 in DMSO-d6 measured at 500 MHz ...163 Figure 102. ¹H NMR of compound 3 in DMSO-d₆ measured at 500 MHz . A – Before Figure 103. HMQC spectra of compound 3 in DMSO-d₆ measured at 500 MHz ...164 Figure 104. Structure of 5,6,8- trihydroxy-4-(1'-hydroxyethyl) isocoumarin (3)......165 Figure 105. ¹H NMR spectra of compound 4 in CDCl₃ measured at 500 MHz166

Figure 106. HSQC spectrum of compound 4 in CDCl₃ measured at 500 MHz 167 Figure 107. HMBC spectrum of compound 4 in CDCI₃ measured at 500 MHz. 167 Figure 109. Chromatogram of subfraction FrF-2. Elution in a polarity gradient: 0 min -70% H₂O + 30% CH₃CN; 20 min - 22 min - 30% H₂O + 70% CH₃CN; 25 min - 70% $H_2O + 30\%$ CH₃CN, monitoring in 300 nm. Compound **5** (Rt = 14.45 min), and **6** (Rt = Figure 110. ¹H NMR spectrum of compound 5 in CDCl₃ measured at 400 MHz170 Figure 111. COSY correlation spectrum of compound 5 in CDCI₃ (400 MHz).171 Figure 112. HSQC spectrum of compound 5 in CDCl₃ (400 MHz)172 Figure 113. HMBC spectrum of compound 5 in CDCI₃ (400 MHz)172 Figure 114. Structure of aspergillumarin B (5).....173 Figure 115. ¹H NMR spectrum of compound 6 in CDCl₃ (400 MHz)174 Figure 116. COSY correlation spectrum of compound 6 in CDCI₃ (400 MHz) 175 Figure 117. HSQC correlation spectrum of compound 6 in CDCl₃ (400 MHz)175 Figure 118. HMBC correlation spectra of compound 6, in CDCl₃ (400 MHz)176 Figure 119. Chromatogram of subfraction FrF-4, elution in a polarity gradient: 0 min -65% H₂O + 35% CH₃CN; 10 min – 22 min - 25% H₂O + 75% CH₃CN; 25 min – 65% Figure 121. COSY correlation spectrum of compound 7 in MeOD₄ (500 MHz), showing Figure 122. HSQC correlation spectrum of compound 7 in MeOD₄ (500 MHz) 180 Figure 123. HMBC correlation spectrum of compound 7 in MeOD₄ (500 MHz)181 Figure 124. ¹H NMR spectrum of compound 8 in CDCI₃ measured at 500 MHz.....182 Figure 125. COSY correlation spectrum of compound 8 in CDCl₃ (500 MHz)......183 Figure 126. HSQC correlation spectrum of compound 8 in CDCl₃ (500 MHz) 183 Figure 127. HMBC correlation spectrum of compound 8 in CDCl₃ (500 MHz)184 Figure 129. Absorption spectra in the UVA-UVB region and photostability assay based on the absorption spectrum after UVA irradiation (dotted line) or not (continuous line). A – Fraction E; B – Fraction F; C – Fraction G; D – Fraction H; E – Fraction I. 187 Figure 132. HaCaT cell viability after treatment with the isolated compounds (200, 100 and 50 µg/mL) and ethylhexyl methoxycinnamate (MTX; 100 and 200 µg/mL) followed by UVB irradiation (300 mJ/cm²). Untreated non-irradiated (NT – UV); untreated Figure 133. Preliminary cytotoxicity of sescandelin and vermistatin to HaCaT cells. The results are expressed as % of cellular viability. Different letters indicate statistically significant differences among the groups. The results are presented as mean ± standard error of the mean of three independent experiments in triplicates (n = 3).193Figure 134. Quantification of intracellular ROS induced by UVA radiation in HaCaT cells after the pretreatment with the test samples for 1 h. The results are expressed as % fluorescence. The cells were untreated irradiated (+UV) or non-irradiated (-UV) and pretreated with quercetin (Q: 10 µg/mL), norfloxacin (N: 100 µg/mL), sescandelin and vermistatin 4 (200, 100 and 50 µg/mL).....194 Figure 135. Evaluation of the irritant potential of sescandelin (7) and vermistatin (8) (0.2%) by the HET-CAM assay. The vascular effects were monitored for 5 min after Figure 137. Scheme of seaweed disinfection to the isolation of endophytic fungi. .214 Figure 140. Colonies of fungal endophytes emerging from inner tissues of P.

Figure 145. OPLS-DA scatter plot of the NMR spectral data of the fungal extracts grouped according to their cultivation media; RX2: 0.75, R2Y: 0.96; Q2: 0.75. .232 Figure 146. OPLS-DA scatter plot of the NMR spectral data of the fungal extracts grouped according to their cultivation media. Variables are colored according to their chemical shifts 0 – 13 ppm.233 **Figure 147.** Heatmap showing the distribution of the top eighty variables (chemical shifts). A - Variables in each extract and media; B - Average of variables in all malt (pink) and marine (green) media......233 Figure 148. Fungal crude extracts yields (mg) in marine and malt agar......235 Figure 149. PCA score scatter plot of the ¹H NMR spectral data of fungal crude Figure 150. PCA loadings scatter plot of the ¹H NMR spectral data of fungal crude extracts LMC1001 to LMC1029; Variables are colored by their chemical shifts Figure 151. PCA score scatter plot of the HRMS data of fungal crude extracts Figure 152. PCA loadings scatter plot of the HRMS data of fungal crude extracts LMC1001 to LMC1029; Expansion of discriminant variables in the Penidiella sp. (pink box) and Cladosporium sp. LMC1001 and 1026 (green); R2: 0.818, Q2: Figure 153. Putative structures of the dereplicated discriminant metabolites present in the *Cladosporium* sp. (LMC1001 and 1026) and *Penidiella* sp. (LMC1010/1012). Figure 154. Phylogenetic tree of fungi Cladosporium sp. isolated from the seaweed P. Figure 155. PLS-DA score scatter plot of the ¹H NMR spectral data of *Cladosporium* sp. crude extracts; R2X 0.96, R2Y 0.91, Q2 0.68......245 Figure 156. PLS-DA loadings scatter plot of the ¹H NMR spectral data of Figure 157. PLS-DA scatter plot of the LC-HRMS spectral data of fungal Figure 158. PLS-DA loadings scatter plot of the HRMS data of Cladosporium sp. crude extracts; Chemogroup 1 (C. halotolerans), Chemogroup 2 (C. velox, C. perangustum, C. pseudocladosporioides) and Chemogroup 3 (C. tenuissimum, C. cladosporioides);

Figure 161. OPLS-DA scores scatter plot the NMR ¹H spectral data of fungal crude extracts (LMC 1001 – 1029) grouped according to their antimicrobial activity results against biofilm-forming S. aureus (ATCC 43300). R2X 0.85 R2Y 0.92 Q2 0.83.....253 Figure 162. OPLS-DA S-plot scatter plot the NMR ¹H spectral data of fungal crude extracts (LMC 1001 – 1029) grouped according to their antimicrobial activity results against biofilm-forming S. aureus (ATCC 43300); Variables are colored according to their chemical shifts 0 – 13 ppm; R2X 0.85 R2Y 0.92 Q2 0.83......253 Figure 163. OPLS-DA scores scatter plot the HRMS data of fungal crude extracts (LMC 1001 – 1029) grouped according to their antimicrobial activity results against biofilm-forming S. aureus (ATCC 43300). R2X 0.45 R2Y 0.93 Q2 0.88.254 Figure 164. OPLS-DA loadings scatter plot the HRMS data of fungal crude extracts (LMC 1001 – 1029) grouped according to their antimicrobial activity results against biofilm-forming S. aureus (ATCC 43300). R2X 0.45 R2Y 0.93 Q2 0.88.255 Figure 165. OPLS-DA plot list plot the HRMS data of fungal crude extracts showing the distribution of the discriminant variables in the active extracts; R2X 0.45 R2Y 0.93 Figure 166. Comparative TLC of C. perangustum (A – green), P. terrigenum (B –

Figure 168. Different macromorphology of <i>E. nigrum</i> grown in malt broth – sea salt
after 14 days of cultivation258
Figure 169. TLC chemical profile of <i>E. nigrum</i> extracts; A – Extract from the small
scale and screening steps; $\mathbf{B} - \mathbf{D}$ Extracts from the 3 batches of scale up fermenation;
E – Extract from fermentation in 2 L flaks259
Figure 170. Comparation of <i>E. nigrum</i> extracts by ¹ H NMR (DMSO-d ₆ , 400 MHz).
A - C Extracts from the 3 batches of scale up fermenation; D – Extract from
fermentation in 2 L flaks
Figure 171. MPLC fractions of the E. nigrum crude extracts (A); Precipitation of
compounds during the fractionation process (B)261
Figure 172. Summary TLC plate of fractions E.N-1 to E.N-25. Elution system DCM:
MeOH 9:1
Figure 173. Extract yields of fractions E.N-1 to E.N-25
Figure 174. Stacked proton NMR spectra of fractions E.N-1 to E.N-15. Spectra was
colored according to the elution solvent in the MPLC fractionantion. Boxes
represent predominant signals in each fraction group
Figure 175. OPLS-DA scores scatter plot the NMR ¹ H spectral data of <i>E. nigrum</i>
fractions $(1 - 25)$ grouped according to their antimicrobial activity results against
biofilm-forming S. aureus (ATCC 43300). R2X 0.94 R2Y 0.99 Q2 0.74265
Figure 176. OPLS-DA loadings scatter plot the NMR ¹ H spectral data of <i>E. nigrum</i>
fractions (1 - 25) grouped according to their antimicrobial activity results against
biofilm-forming S. aureus (ATCC 43300). R2X 0.94 R2Y 0.99 Q2 0.74266
Figure 177. OPLS-DA score scatter plot the HRMS data of <i>E. nigrum</i> fractions (1 –
25) grouped according to their antimicrobial activity results against biofilm-forming S.
aureus (ATCC 43300). R2X 0.22 R2Y 0.94 Q2 0.77
Figure 178. OPLS-DA loadings scatter plot the HRMS data of <i>E. nigrum</i> fractions (1
- 25) grouped according to their antimicrobial activity results against biofilm-forming
S. aureus (ATCC 43300). R2X 0.22 R2Y 0.94 Q2 0.77
Figure 179. OPLS-DA S-plot the HRMS data of <i>E. nigrum</i> fractions $(1 - 25)$ grouped
according to their antimicrobial activity results against biofilm-forming S. aureus (ATCC
43300). R2X 0.22 R2Y 0.94 Q2 0.77
Figure 180. Putative structures of the dereplicated discriminant metabolites present
in the <i>E. nigrum</i> active fractions. Compounds circled in pink were isolated in this work.

Figure 181. Chemical profile by ¹H NMR of fractions E.N-10 to E.N-13 (DMSO-d6, 400 MHz); E.N-10 (blue line); E.N-11 (orange line); E.N-12 (pink line); E.N-13 Figure 182. Chemical profile by ¹H NMR of fractions E.N-10 to E.N-13 (DMSO-d6, 400 MHz); E.N-14 (purple line); E.N-11 (orange line); E.N-12 (pink line); E.N-13 Figure 183. TLC summary plate of subfractions E.1-S1 to E.1-S-20. Mobile phase: dichloromethane, and methanol 9:1, staining with anisaldehyde......273 Figure 184. TLC summary plate of subfractions E.2-S1 to E.2-S-12. Mobile phase: dichloromethane, and methanol 9:1, staining with anisaldehyde......273 Figure 185. Flowchart of the isolation of compounds from the fungus E. nigrum. ..274 Figure 186. ¹H NMR spectra of compound 11 in DMSO-d₆ (400 MHz).275 Figure 187. HSQC correlation spectrum of compound 9 in DMSO-d₆ (400 MHz)...276 Figure 188. COSY correlation spectrum of compound 9 in DMSO-d₆ (400 MHz)...276 **Figure 189.** HMBC correlation spectrum of compound **9** in DMSO-d₆ (400 MHz)...277 Figure 190. ¹³C NMR spectrum of compound 9 in DMSO-d₆ (400 MHz).277 Figure 192. COSY correlation spectrum of compound 10 in DMSO-d₆ (400 MHz). 280 Figure 193. HSQC correlation spectrum of compound 10 in DMSO-d₆ (400 MHz). 281 Figure 194. HMBC correlation spectrum of compound 10 in DMSO-d₆ (400 MHz).281 Figure 195. ROESY correlation spectrum of compound 10 in DMSO-d₆ (400 MHz).

LIST OF TABLES

Table 1. Collection sites of P. antarcticus in the South Shetland Islands, Antarctica.
Table 2. Elution gradient used in the VLC fractionation of P. antarcticus extracts25
Table 3. Antibacterial activity of <i>P. antarcticus</i> crude extracts and fractions against
pathogenic bacteria32
Table 4. Antileishmanial activity of P. antarcticus crude extracts and fractions against
L. amazonensis
Table 5. Anti-Neospora activity of P. antarcticus n-hexane extract and fractions34
Table 6. Annotated compounds in P. antarcticus bioactive fractions. 41
Table 7. Larvicidal activity of P. antarcticus crude extracts and fractions. 42
Table 8. Annotated compounds in <i>P. antarcticus</i> fractions with larvicidal activity44
Table 9. Collection sites of P. antarcticus in the South Shetland Islands, Antarctica. 51
Table 10 MzMine parameters used in the deconvolution of fundal crude extracts and
fractions
Table 11 Elution gradient used in the MPLC fractionation of <i>P</i> antarcticus MoOH
crude extract
Table 12 Elution gradient used in the sub-fractionation of fraction $PA = Fr^{2}_{0}$
Table 12. Elution gradient used in the sub-fractionation of fraction $A = 110$
loadings plot (Po) polarity $P = positive mode N = pegative mode: Pt retention time in$
minutes: MW molecular weight: ME molecular formula: DBE double bond equivalent:
^a Arctowski ^b Groopwich 2015 ^c Halfmoon ^d Groopwich 2018 ^e Domay Point ^f Vauroal
Point 9 Ponguin
Table 14 Antibiofilm activity of <i>P</i> antarcticus crude extracts against S aurous (ATCC
42200) at a concentration of 100 µg/ml
Table 15 Antibiofilm activity of <i>D</i> anterational MoOH and a avtract and fractional
Table 15. Antibionini activity of <i>P. antarcticus</i> MeOH crude extract and fractions
Table 10. Devention of motobalitae predicted on bioactive veriables in the ODL C
Table 16. Dereplication of metabolites predicted as bloactive variables in the OPLS-
DA loadings plot. (Po) polarity, $P = positive mode$, $N = negative mode$; Rt retention
time in minutes; IVIV molecular weight; IVIF molecular formula; DBE double bond
Table 17. 'H NMR experimental data of compound 1 in comparation to literature data
of linoleic acid

Table 18. ¹ H and ¹³ C NMR experimental data of compound 2 in comparation to
literature data of fucosterol95
Table 19. Mobile phase used for the VLC fraction of the crude extract119
Table 20. Mobile phase used to analyze the crude extract and fractions chemical
profiles120
Table 21. Mobile phase used in the purification of sub fraction FrE-6120
Table 22. Mobile phase used in the subfractionation of FrF121
Table 23. Mobile phase used in the purification of sub fraction FrF-1 – FrF-4121
Table 24. Identification of fungi, collection site and disinfection method. 129
Table 25. Remaining percentage of the area under the curve of the irradiated samples
compared to non-irradiated samples considered as 100% in the UVA and UVB range.
Table 26. Annotated compounds and their main neighboring nodes indicated as the
discriminant features in the OPLS-DA loadings plot. Compounds in blue were pointed
in the as the discriminant features149
Table 27. Cytotoxic effects of P. purpurogenum crude extracts to human neutrophils.
Table 28. % ROS inhibition in human neutrophils treated with P. purpurogenum153
Table 28 . % ROS inhibition in human neutrophils treated with P. purpurogenum153 Table 29. Antileishmanial activity of P. purpurogenum ASW-PDC EtOAc extracts
Table 28. % ROS inhibition in human neutrophils treated with P. purpurogenum153Table 29. Antileishmanial activity of P. purpurogenum ASW-PDC EtOAc extractsagainst L. amazonensis promastigotes.154
 Table 28. % ROS inhibition in human neutrophils treated with P. purpurogenum153 Table 29. Antileishmanial activity of P. purpurogenum ASW-PDC EtOAc extracts against L. amazonensis promastigotes
Table 28. % ROS inhibition in human neutrophils treated with P. purpurogenum153Table 29. Antileishmanial activity of P. purpurogenum ASW-PDC EtOAc extractsagainst L. amazonensis promastigotes
Table 28. % ROS inhibition in human neutrophils treated with P. purpurogenum153Table 29. Antileishmanial activity of P. purpurogenum ASW-PDC EtOAc extractsagainst L. amazonensis promastigotes
Table 28. % ROS inhibition in human neutrophils treated with P. purpurogenum153Table 29. Antileishmanial activity of P. purpurogenum ASW-PDC EtOAc extractsagainst L. amazonensis promastigotes
Table 28. % ROS inhibition in human neutrophils treated with P. purpurogenum153Table 29. Antileishmanial activity of P. purpurogenum ASW-PDC EtOAc extractsagainst L. amazonensis promastigotes
Table 28. % ROS inhibition in human neutrophils treated with P. purpurogenum153Table 29. Antileishmanial activity of P. purpurogenum ASW-PDC EtOAc extractsagainst L. amazonensis promastigotes
Table 28. % ROS inhibition in human neutrophils treated with P. purpurogenum153Table 29. Antileishmanial activity of P. purpurogenum ASW-PDC EtOAc extractsagainst L. amazonensis promastigotes
Table 28. % ROS inhibition in human neutrophils treated with P. purpurogenum153Table 29. Antileishmanial activity of P. purpurogenum ASW-PDC EtOAc extractsagainst L. amazonensis promastigotes
Table 28. % ROS inhibition in human neutrophils treated with P. purpurogenum153Table 29. Antileishmanial activity of P. purpurogenum ASW-PDC EtOAc extractsagainst L. amazonensis promastigotes
Table 28. % ROS inhibition in human neutrophils treated with P. purpurogenum153Table 29. Antileishmanial activity of P. purpurogenum ASW-PDC EtOAc extractsagainst L. amazonensis promastigotes154Table 30. 1H and 13C NMR experimental data of compound 3 in comparation toliterature data of 5,6,8- trihydroxy-4-(1'-hydroxyethyl) isocoumarin165Table 31. 1H and 13C NMR experimental data of compound 4 in comparation toliterature data of aspergillumarin A168Table 32. 1H and 13C NMR experimental data of compound 5 in comparation toliterature data of aspergillumarin B173Table 33. 1H and 13C NMR experimental data of compound 6 in comparation toliterature data of Berkeleyacetal C177Table 34. 1H and 13C NMR experimental data of compound 7 in comparation to
Table 28. % ROS inhibition in human neutrophils treated with P. purpurogenum153Table 29. Antileishmanial activity of P. purpurogenum ASW-PDC EtOAc extractsagainst L. amazonensis promastigotes154Table 30. ¹ H and ¹³ C NMR experimental data of compound 3 in comparation toliterature data of 5,6,8- trihydroxy-4-(1'-hydroxyethyl) isocoumarin165Table 31. ¹ H and ¹³ C NMR experimental data of compound 4 in comparation toliterature data of aspergillumarin A168Table 32. ¹ H and ¹³ C NMR experimental data of compound 5 in comparation toliterature data of aspergillumarin B173Table 33. ¹ H and ¹³ C NMR experimental data of compound 6 in comparation toliterature data of Berkeleyacetal C177Table 34. ¹ H and ¹³ C NMR experimental data of compound 7 in comparation toliterature data of Sescandelin
Table 28. % ROS inhibition in human neutrophils treated with P. purpurogenum153Table 29. Antileishmanial activity of P. purpurogenum ASW-PDC EtOAc extractsagainst L. amazonensis promastigotes154Table 30. 1H and 13C NMR experimental data of compound 3 in comparation toliterature data of 5,6,8- trihydroxy-4-(1'-hydroxyethyl) isocoumarin165Table 31. 1H and 13C NMR experimental data of compound 4 in comparation toliterature data of aspergillumarin A168Table 32. 1H and 13C NMR experimental data of compound 5 in comparation toliterature data of aspergillumarin B173Table 33. 1H and 13C NMR experimental data of compound 6 in comparation toliterature data of Berkeleyacetal C177Table 34. 1H and 13C NMR experimental data of compound 6 in comparation toliterature data of Berkeleyacetal C177Table 34. 1H and 13C NMR experimental data of compound 7 in comparation toliterature data of Sescandelin.181Table 35. 1H and 13C NMR experimental data of compound 7 in comparation toliterature data of Sescandelin.181Table 35. 1H and 13C NMR experimental data of compound 7 in comparation to
Table 28. % ROS inhibition in human neutrophils treated with P. purpurogenum153Table 29. Antileishmanial activity of P. purpurogenum ASW-PDC EtOAc extractsagainst L. amazonensis promastigotes.Table 30. 1H and 13C NMR experimental data of compound 3 in comparation toliterature data of 5,6,8- trihydroxy-4-(1'-hydroxyethyl) isocoumarin.165Table 31. 1H and 13C NMR experimental data of compound 4 in comparation toliterature data of aspergillumarin A.168Table 32. 1H and 13C NMR experimental data of compound 5 in comparation toliterature data of aspergillumarin B.173Table 33. 1H and 13C NMR experimental data of compound 6 in comparation toliterature data of Berkeleyacetal C.177Table 34. 1H and 13C NMR experimental data of compound 6 in comparation toliterature data of Sescandelin181Table 35. 1H and 13C NMR experimental data of compound 7 in comparation toliterature data of Sescandelin181Table 35. 1H and 13C NMR experimental data of compound 8 in comparation toliterature data of Sescandelin181Table 35. 1H and 13C NMR experimental data of compound 7 in comparation toliterature data of Sescandelin181Table 35. 1H and 13C NMR experimental data of compound 8 in comparation toliterature data of Sescandelin181Table 36. Remaining percentage of the area under the curve of the irradiated samples

Table 39. Irritation scores, reported as mean \pm standard errors of the mean (n = 4),and classification of the effects of compounds in the HET-CAM assay. The sodiumlauryl sulfate (SDS) and sodium chloride (NaCl) solutions were used as positive andnegative controls, respectively.196

 Table 40. Cytotoxicity of fractions FrA – FrD to human neutrophils expressed as % of cellular viability. Hanks's solution was used as negative control.
 197

 Table 41. Cytotoxicity of fractions FrE – FrI to human neutrophils expressed as % of cellular viability. Hanks's solution was used as negative control.
 197

 Table 42. Cytotoxicity of isolated compounds to human neutrophils expressed as % of cellular viability. Hanks's solution and DMSO 1% were used as controls.
 198

 Table 43. Immunomodulation effects of the VLC fractions on ROS production by human neutrophils oxidative metabolism expressed as % percentage of ROS inhibition. Hanks's solution and DMSO 1% were used as controls.
 198

 Table 44. Immunomodulation effects of isolated compounds on ROS production by human neutrophils expressed as half maximal inhibitory concentration IC₅₀. Hanks's solution and DMSO 1% were used as controls.
 198

 Table 44. Immunomodulation effects of isolated compounds in on ROS production by human neutrophils expressed as half maximal inhibitory concentration IC₅₀. Hanks's solution and DMSO 1% were used as controls.
 199

 Table 45. Immunomodulation effects of isolated compounds in on ROS production by human neutrophils expressed as half maximal inhibitory concentration IC₅₀. Hanks's solution and DMSO 1% were used as controls.
 199

Table 47. Antileishmanial activity of subfractions from FrF against *L. amazonensis*

 promastigotes. Antileishmanial activity is expressed as half-maximal inhibitory

 concentrations (IC_{50-PRO}).

 202

Table 48. Antileishmanial activity of isolated compounds against *L. amazonensis* and cytotoxicity to murine macrophages. Antileishmanial activity is expressed as half-maximal inhibitory concentrations IC_{50-PRO} for promastigotes and IC_{50-AMA} for

amastigotes. Cytotoxic activity is expressed as half-maximal cytotoxic concentration
 Table 49. SWBG-11 media composition for stock solution and media preparation.213

 Table 50. Collection locations of the seaweed P. antarcticus
 214
 Table 51. MzMine parameters used in the deconvolution of fungal crude extracts and **Table 52.** Elution gradient used in the MPLC fractionation of *E. nigrum* crude extract. Table 54. Elution gradient used in the sub-fractionation of fraction E.N-13-17.223
 Table 55. Molecular identification of endophytic fungi recovered from the inner tissues
 Table 56. Antibiofilm activity of fungi crude extracts against S. aureus at a concentration of 100 µg/mL......234 Table 57. Dereplication of metabolites predicted as discriminant variables in the OPLS-DA loadings plot. (Po) polarity, P = positive mode, N = negative mode; Rt retention time in minutes; MW molecular weight; MF molecular formula; DBE double **Table 58.** Molecular identification of endophytic *Cladosporium* sp. isolated from of *P*. Table 59. Molecular identification of endophytic *Cladosporium* sp. isolated from P. antarcticus by ACT and EF sequencing.243 Table 60. Dereplication of metabolites predicted as discriminant variables in the PLS-DA loadings plot. (Po) polarity, P = positive mode, N = negative mode; Rt retention time in minutes; MW molecular weight; MF molecular formula; DBE double bond equivalent......248 Table 61. Antibiofilm activity of fungi crude extracts against S. aureus at a concentration of 100 µg/mL.....252 Table 62. Dereplication of metabolites predicted as discriminant variables in the PLS-DA loadings plot. (Po) polarity, P = positive mode, N = negative mode; Rt retention time in minutes; MW molecular weight; MF molecular formula; DBE double bond equivalent......256 Table 63. Antibiofilm activity of E. nigrum fractions against S. aureus at a concentration of 100 µg/mL......264

Table 64. Dereplication of metabolites predicted as discriminant variables in the OPLS-
DA loadings plot. (Po) polarity, P = positive mode, N = negative mode; Rt retention
time in minutes; MW molecular weight; MF molecular formula; DBE double bond
equivalent269
Table 65. ¹ H and ¹³ C NMR experimental data of compound 9 in comparation to
literature data of epicorazine A278
Table 66. ¹ H and ¹³ C NMR experimental data of compound 10 in comparation to
literature data of epicorazine B283
Table 67. ¹ H and ¹³ C NMR experimental data of compound 11
Table 68. ¹ H and ¹³ C NMR experimental data of compound 12 in comparation to
literature data of epicorazine C292
Table 69. Antibiofilm activity of isolated compounds against S. aureus. 293

LIST OF ABREVIATIONS

ASW	Artificial seawater
COSY	Correlation spectroscopy
EtOAc	Ethyl acetate
GC-MS	Gas chromatography mass spectrometry
h	hour
HMBC	Heteronuclear Multiple Bound Correlation
HRMS	High resolution mass spectrometry
HSQC	Heteronuclear Single Quantum Correlation
ΗХ	Hexane
m/z	mass over charge
MeOH	Methanol
min	minutes
MPLC	Medium pressure liquid chromatography
NMR	Nuclear magnetic resonance
NSW	Natural seawater
OPLS-DA	Orthogonal partial least squares discriminant analysis
PCA	Principal component analysis
R <i>t</i>	Retention time
TLC	Thin layer chromatography
UVR	Ultraviolet radiation
VLC	Vacuum liquid chromatography

TABLE OF CONTENTS

RESUMO	I
ABSTRACT	II
LIST OF FIGURES	
LIST OF TABLES	XVI
LIST OF ABREVIATIONS	XXI
1. INTRODUCTION	1
1. 1 The Antarctic continent	1
1. 2 The Brazilian Antarctic Program and the University of São Paulo	4
1. 3 Bioprospecting to Protect	5
1. 4 Antarctic Natural products	6
1. 4. 1 Natural products from Antarctic seaweed	8
1. 4. 2 Natural products from Antarctic marine fungi	9
1. 5 The seaweed Phaeurus antarcticus	12
1. 6 Metabolomic and chemometrics approaches	12
1. 7 Investigated biological activities	14
1.7.1 Photoprotection	14
1. 7. 2 Neutrophils as new drugs target for chronic inflammatory diseases	s 15
1. 7. 3 Leishmaniasis	16
1. 7. 4 Neosporosis	16
1. 7. 5 Antibacterial and antibiofilm	17
1. 7. 6 Aedes aegypti control	18
1. 8 Preface	19
2. CHAPTER I – Chemical profiling and bioactivity potential of the seawed	€d
Phaeurus antarcticus	21
2.1 INTRODUCTION	22
2. 2 OBJECTIVES	23
2. 2. 1 General objective	23
2. 2. 2 Specific objectives	23
2. 3 MATERIAL AND METHODS	24

2. 3. 1 Material and equipment	24
2. 3. 2 Collection of the seaweed P. antarcticus	24
2. 3. 3 Extracts preparation	25
2. 3. 4 GC-MS molecular networking and multivariate analysis	26
2. 3. 5 Antibacterial activity	27
2. 3. 6 Antileishmanial activity	27
2. 3. 7 Anti-Neospora activity	29
2. 3. 8 Cytotoxicity to human fibroblasts	29
2. 3. 9 Larvicidal activity (Aedes aegypti)	
2. 4 RESULTS AND DISCUSSION	31
2. 4. 1 Antibacterial activity	31
2. 4. 2 Antileishmanial activity	32
2. 4. 3 Anti-Neospora activity	34
2. 4. 4 GC-MS molecular networking and multivariate analysis	35
2. 4. 5 Larvicidal activity (Aedes aegypti)	42
2.5 CONCLUSION	45
3 CHAPTER II – Spatial variability in secondary metabolites and antibio	ofilm
activity of the seaweed Phaeurus antarcticus	47
activity of the seaweed <i>Phaeurus antarcticus</i>	47 48
activity of the seaweed <i>Phaeurus antarcticus</i>	47 48 49
activity of the seaweed <i>Phaeurus antarcticus</i>	47 48 49 49
 activity of the seaweed <i>Phaeurus antarcticus</i>	47 48 49 49 49 49
 activity of the seaweed <i>Phaeurus antarcticus</i>	47 48 49 49 49 49 50
 activity of the seaweed <i>Phaeurus antarcticus</i>	47 48 49 49 49 50 50
 activity of the seaweed <i>Phaeurus antarcticus</i>	47 48 49 49 49 49 50 50 50
 activity of the seaweed <i>Phaeurus antarcticus</i>	47 48 49 49 49 50 50 51 51
activity of the seaweed <i>Phaeurus antarcticus</i>	47 48 49 49 49 50 50 50 51 52 52
activity of the seaweed <i>Phaeurus antarcticus</i>	47 48 49 49 49 50 50 50 51 52 52 52
activity of the seaweed <i>Phaeurus antarcticus</i>	47 48 49 49 49 49 50 50 51 52 52 52 52 52
activity of the seaweed <i>Phaeurus antarcticus</i> 3. 1 INTRODUCTION 3. 2 OBJECTIVES 3. 2. 1 General objective 3. 2. 2 Specific objectives 3. 3 MATERIAL AND METHODS 3. 3. 1 Material and equipment 3. 3. 2 Seaweed collection 3. 3. 2 Seaweed collection 3. 3. 3 Extracts preparation. 3. 3. 4 Chemical profiling by UHPLC-HRMS 3. 3. 5 ¹ H NMR spectral analysis 3. 3. 6 Chemometrics and multivariate analysis. 3. 3. 6. 1 HRMS data processing.	47 48 49 49 49 50 50 50 51 52 52 52 52 52 52 52 52 52 52 52 52 52
activity of the seaweed <i>Phaeurus antarcticus</i>	47 48 49 49 49 50 50 50 51 52 52 52 52 52 52 53 53 54
activity of the seaweed <i>Phaeurus antarcticus</i>	47 48 49 49 49 50 50 50 50 51 52 52 52 52 52 52 52 52 52 52 52 52 52

3. 3. 8 Antimicrobial activity	55
3. 3. 8. 1 Pre-culture	55
3. 3. 8. 2 Main bacterial suspension for biofilm assay	55
3. 3. 8. 3 Pre-biofilm inhibition assay	56
3. 3. 8. 4 Post-biofilm inhibition assay	56
3. 3. 8. 5 MIC and Minimum biofilm eradication concentration (MBEC)	57
3. 4 RESULTS AND DISCUSSION	57
3. 4. 1 Spatial variability in <i>Phaeurus antarcticus</i> secondary metabolites	57
3. 4. 2 Antibiofilm activity of <i>P. antarcticus</i> collected in different locations	79
3. 4. 3 Antibiofilm activity of <i>P. antarcticus</i> methanolic extract	82
3. 4. 4 Isolated compounds	89
3. 4. 4. 1 Structure elucidation of compound 1	90
3. 4. 2 Structural elucidation of compound 2	93
3. 4. 5 Antibiofilm activity of isolated compounds	96
3. 5 CONCLUSION	97
4. CHAPTER III – Bioguided isolation of bioactive metabolites from the fung	gus
Penicillium purpurogenum	99
4.1 INTRODUCTION	100
4. 1. 1 Photoprotective compounds from marine and polar microorganisms	.100
4. 1. 2 Immunomodulatory compounds from the sea	102
4. 1. 3 Antileishmanial compounds isolated from fungi	105
4. 2 OBJECTIVES	109
4. 2. 1 General objective	109
4. 2. 2 Specific objectives	109
4. 3 MATERIAL AND METHODS	110
4. 3. 1 Material and equipment	110
4. 3. 2 Collection of the seaweed P. antarcticus	112
4. 3. 3 Isolation of endophytic fungi	113
4. 3. 3. 1 Culture media preparation	. 113
4. 3. 3. 1 Culture media preparation4. 3. 3. 2 Disinfection procedures	. 113 . 113
 4. 3. 3. 1 Culture media preparation	. 113 . 113 . 114
 4. 3. 3. 1 Culture media preparation	. 113 . 113 . 114 115

4. 3. 4. 2 Extraction procedures	115
4. 3. 4. 3 Determination of the UV absorption spectrum and photostability test	116
4. 3. 4. 4 Optimization of secondary metabolites production	116
<i>4. 3. 4. 5 ¹H NMR analysis</i>	117
4. 3. 4. 6 Gas chromatography mass spectrometry – GC-MS	117
4. 3. 4. 7 Thin layer chromatography analysis (TLC)	118
4. 3. 5 Scale up fermentation of <i>P. purpurogenum</i>	119
4. 3. 5. 1 Crude extract fractionation	119
4. 3. 5. 2 Chemical profile of VLC fractions FrA – FrI	120
4.3.5.3 Isolation of compounds from fraction FrE	120
4. 3. 5. 4 Isolation of compounds from fraction FrF	120
4. 3. 6 Evaluation of the photoprotective potential	121
4. 3. 6. 1 Determination of the UV absorption spectrum	122
4. 3. 6. 2 Evaluation of Photostability by UV Spectrometry	122
4. 3. 6. 3 Photoreactivity studies – ROS generation assay	122
4. 3. 6. 4 Phototoxicity test in 3T3 mouse fibroblast (3T3 NRU PT)	122
4. 3. 6. 5 UVB and UVA photoprotection assays	123
4. 3. 6. 5. 1 Cytotoxicity to HaCaT cells	123
4. 3. 6. 5. 2 UVB photoprotection assay	123
4. 3. 6. 5. 3 Photoprotection against UVA-induced ROS production	124
4. 3. 6. 6 Evaluation of ocular irritation potential (HET-CAM)	124
4. 3. 6. 7 Antibacterial activity against Cutibacterium acnes	125
4. 3. 7 Evaluation of the immunomodulatory potential	126
4. 3. 7. 1 Cytotoxicity to human neutrophils	126
4. 3. 7. 2 Effect on ROS production by human neutrophils	126
4. 3. 8 Evaluation of the antileishmanial activity	127
4. 3. 8. 1 Anti-promastigote assay	127
4. 3. 8. 2 Anti-amastigote assay	128
4. 3. 9 Evaluation of the antibacterial activity	128
4. 4 RESULTS AND DISCUSSION	129
4. 4. 1 Identification of endophytic fungi	129
4. 4. 2 Screening for UVB-UVA absorbers	131
4. 4. 3 Optimization of secondary metabolites production	132
4. 4. 3. 1 Extraction yields and TLC chemical profile	132

4. 4. 3. 2 Chemical profile by TLC	134
4. 4. 3. 3 Chemical profile by GC-MS	142
4. 4. 3. 4 Determination of the UV absorption spectrum and photostability .	150
4. 4. 3. 5 Evaluation of the immunomodulatory activity	151
4. 4. 3. 6 Evaluation of the antibacterial activity	153
4. 4. 3. 7 Evaluation antileishmanial activity	154
4. 4. 4 Bioguided isolation of compounds	155
4. 4. 5 Compounds isolated from the fungus Penicillium purpurogenum.	159
4. 4. 5. 1 Isolation of compounds from Fraction FrE-6	161
4. 4. 5. 2 Structure elucidation of compound 3	162
4. 4. 5. 3 Structure elucidation of compound 4	166
4. 4. 6 Isolation of compounds from fraction FrF	169
4. 4. 6. 1 Isolation and identification of compounds from subfraction FrF-2	169
4. 4. 6. 2 Structure elucidation of compound 5	170
4. 4. 6. 3 Structure elucidation of compound 6	174
4. 4. 6. 4 Isolation of compounds from subfraction FrF-4	178
4. 4. 6. 5 Structure elucidation of compound 7	179
4. 4. 6. 6 Structure elucidation of compound 8	
4. 4. 7 Photoprotective potential of <i>P. purpurogenum</i> secondary metabol	ites 186
4. 4. 7. 1 Determination of the UV absorption spectrum and photostabilit	y assay
	186
4. 4. 7. 2 Photoreactivity analysis – ROS assay (OECD TC 495)	188
4. 4. 7. 3 Phototoxicity test in 3T3 mouse fibroblast (3T3 NRU PT)	189
4. 4. 7. 4 UV-B photoprotective assay	191
4. 4. 7. 5 Photoprotection against UVA-induced ROS production	193
4. 4. 7. 6 Antibacterial activity against C. acnes	194
4. 4. 7. 7 Evaluation of ocular irritation potential (HET-CAM)	195
4. 4. 8 Immunomodulatory potential of <i>P. purpurogenum</i>	197
4. 4. 8. 1 Cytotoxicity to human neutrophils	197
4. 4. 8. 2 Evaluation of the immunomodulatory activity on ROS production by	human
neutrophils	198
4. 4. 9 Antileishmanial potential of <i>P. purpurogenum</i>	201
4.5 CONCLUSION	204

5. CHAPTER IV – Metabolomic approaches to the identification of an	ntibiofilm
compounds from Phaeurus antarcticus endophytic fungi	207
5.1 INTRODUCTION	208
5. 1. 1 Antibiofilm compounds from marine derived fungi	208
5. 2 OBJECTIVES	210
5. 2. 1 General objective	210
5. 2. 2 Specific objectives	210
5. 3 MATERIAL AND METHODS	211
5. 3. 1 Materials and equipment	211
5. 3. 2 Isolation and identification of endophytic fungi	213
5. 3. 2. 1 Preparation of potato dextrose-agar – SWBG11	213
5. 3. 2. 2 Disinfection of seaweed surface to target the isolation of endop	hytes213
5. 3. 2. 3 Classical identification of endophytic fungi	214
5. 3. 2. 4 Identification of endophytic fungi by ITS gene sequencing	215
5. 3. 2. 5 Identification of Cladosporium sp. by EF-1 and ACT gene seque	<i>encing</i> 215
5. 3. 2. 6 Construction of phylogenetic tree using ITS, EF-1 and ACT seq	uences of
Cladosporium sp	216
5. 3. 3 Preparation of fungal crude extracts for screening	216
5. 3. 3. 1 Preparation of malt agar with sea salt	216
5. 3. 3. 2 Preparation of marine agar	216
5. 3. 3. 3 Preparation of malt broth with sea salt	217
5.3. 3. 4 Extraction procedures for solid media cultures	217
5. 3. 3. 5 Extraction procedures for liquid media cultures	217
5. 3. 4 Scale up fermentation and stability of <i>Epicoccum nigrum</i>	218
5. 3. 5 Chemical profiling and multivariate analysis	218
5. 3. 5. 1 UHPLC-HRMS	218
5. 3. 5. 2 Chemical profiling by ¹ H NMR	220
5. 3. 5. 3 Multivariate analysis	220
5. 3. 6 Fractionation of <i>Epicoccum nigrum</i> crude extract	221
5. 3. 7 Compounds isolation	221
5. 3. 7.1 Crystallization of E.N-Fr7	221
5. 3. 7. 2 Purification of fraction E.N-S1	222
5. 3. 7. 3 Purification of fraction E.N-S2	

5. 3. 8 Antimicrobial activity	223
5. 3. 8. 1 Pre-culture	223
5. 4. 8. 2 Main bacterial suspension for biofilm assay	224
5. 3. 8. 3 Pre-biofilm inhibition assay	224
5. 3. 8. 4 Post-biofilm inhibition assay	225
5. 3. 8. 5 MIC and MBEC	225
5. 4 RESULTS AND DISCUSSION	226
5. 4. 1 Fungal isolation and identification by ITS region sequencing	226
5. 4. 2 Chemodiversity and antibiofilm activity	231
5. 4. 2. 1 Preliminary media screening	231
5. 4. 2. 2 Chemical profiling of endophytic fungi	235
5. 4. 2. 3 Insights into the chemotaxonomy of Antarctic derived Cladospori	um sp.
	240
5. 4. 2. 3. 1 Identification of Cladosporium sp. by ACT and EF-1α gene seque	encing
	240
5. 4. 3. 2 Specificity of secondary metabolites in Antarctic derived Cladospori	um sp.
5. 4. 3. 2 Specificity of secondary metabolites in Antarctic derived Cladospori	um sp. 244
 5. 4. 3. 2 Specificity of secondary metabolites in Antarctic derived Cladospori 5. 4. 3 Antibiofilm activity of <i>P. antarcticus</i> endophytes 	um sp. 244 251
 5. 4. 3. 2 Specificity of secondary metabolites in Antarctic derived Cladospori 5. 4. 3 Antibiofilm activity of <i>P. antarcticus</i> endophytes 5. 4. 4 Antimicrobial and antibiofilm activity of <i>Epicoccum nigrum</i> 	um sp. 244 251 258
 5. 4. 3. 2 Specificity of secondary metabolites in Antarctic derived Cladospori 5. 4. 3 Antibiofilm activity of <i>P. antarcticus</i> endophytes 5. 4. 4 Antimicrobial and antibiofilm activity of <i>Epicoccum nigrum</i> 5. 4. 1 Crude extract fractionation 	um sp. 244 251 258 258
 5. 4. 3. 2 Specificity of secondary metabolites in Antarctic derived Cladospori 5. 4. 3 Antibiofilm activity of <i>P. antarcticus</i> endophytes 5. 4. 4 Antimicrobial and antibiofilm activity of <i>Epicoccum nigrum</i> 5. 4. 1 Crude extract fractionation 5. 4. 2 Chemical profiling and antibiofilm activity of <i>E. nigrum</i> fractions 	um sp. 244 251 258 258 258 262
 5. 4. 3. 2 Specificity of secondary metabolites in Antarctic derived Cladospori 5. 4. 3 Antibiofilm activity of <i>P. antarcticus</i> endophytes 5. 4. 4 Antimicrobial and antibiofilm activity of <i>Epicoccum nigrum</i> 5. 4. 4. 1 Crude extract fractionation 5. 4. 2 Chemical profiling and antibiofilm activity of <i>E. nigrum</i> fractions 5. 4. 3 Isolation of antibiofilm compounds from the fungus <i>E. nigrum</i> 	um sp. 244 251 258 258 262 271
 5. 4. 3. 2 Specificity of secondary metabolites in Antarctic derived Cladospori. 5. 4. 3 Antibiofilm activity of <i>P. antarcticus</i> endophytes	um sp. 244 251 258 258 262 262 271 275
 5. 4. 3. 2 Specificity of secondary metabolites in Antarctic derived Cladospori. 5. 4. 3 Antibiofilm activity of <i>P. antarcticus</i> endophytes	um sp. 244 251 258 258 262 262 271 275 279
 5. 4. 3. 2 Specificity of secondary metabolites in Antarctic derived Cladosporie 5. 4. 3 Antibiofilm activity of <i>P. antarcticus</i> endophytes 5. 4. 4 Antimicrobial and antibiofilm activity of <i>Epicoccum nigrum</i> 5. 4. 4. 1 Crude extract fractionation 5. 4. 4. 2 Chemical profiling and antibiofilm activity of <i>E. nigrum</i> fractions. 5. 4. 3 Isolation of antibiofilm compounds from the fungus <i>E. nigrum</i> 5. 4. 4. 5 Structural elucidation of compound 10. 5. 4. 4. 6 Structural elucidation of compound 11. 	um sp. 244 251 258 258 262 271 275 279 279 284
 5. 4. 3. 2 Specificity of secondary metabolites in Antarctic derived Cladospori 5. 4. 3 Antibiofilm activity of <i>P. antarcticus</i> endophytes 5. 4. 4 Antimicrobial and antibiofilm activity of <i>Epicoccum nigrum</i> 5. 4. 4 Antimicrobial and antibiofilm activity of <i>E. nigrum nigrum</i> 5. 4. 4. 2 Chemical profiling and antibiofilm activity of <i>E. nigrum fractions</i> 5. 4. 3 Isolation of antibiofilm compounds from the fungus <i>E. nigrum</i> 5. 4. 4. 5 Structural elucidation of compound 9 5. 4. 6 Structural elucidation of compound 11 5. 4. 7 Structural elucidation of compound 12 	um sp. 244 251 258 258 262 262 271 275 279 279 284 288
 5. 4. 3. 2 Specificity of secondary metabolites in Antarctic derived Cladospori. 5. 4. 3 Antibiofilm activity of <i>P. antarcticus</i> endophytes	um sp. 244 251 258 258 258 262 271 275 279 279 279 284 288 293
 5. 4. 3. 2 Specificity of secondary metabolites in Antarctic derived Cladospori. 5. 4. 3 Antibiofilm activity of <i>P. antarcticus</i> endophytes	um sp. 244 251 258 258 258 262 271 275 279 279 279 284 288 293 295
1. INTRODUCTION

1.1 The Antarctic continent

Antartica is known as the "Continent of Superlatives" and represents one of the world's last frontiers. The continental landscape is characterized by rash climate conditions, such as extremely low temperatures, strong seasonal shifts in solar radiation, high ultraviolet exposure in the summer and darkness in winter, and heavy seasonal changes in ice cover (DINASQUET et al., 2018; WENZEL et al., 2016).

The continent is home to remote locations with pristine and little-studied ecosystems, each with their own distinct characteristics (Fig. 1). Surrounded by an ocean, the world's highest, driest, windiest, and coldest continent possess very distinctive environmental conditions that play an important role in the distribution and development of life (WENZEL et al., 2016).



Figure 1. Antarctic glacier near the Comandante Ferraz Brazilian Antarctic Base.

Source: G. S. SANTOS (2017).

The Antarctic biodiversity is mainly dominated by marine organisms, seabirds and microorganisms and lacks terrestrial mammalian predators (Fig. 2). In fact, until the establishment of scientific bases, even humans were absent from the continent (MELTOFTE, 2019).

Figure 2. Antarctic fauna. **A** – *Pygoscelis antarcticus*; **B** – *Pygoscelis adeliae*; **C** – *Pygoscelis papua*; **D** – *Macronectes giganteus*; **E** – *Hydrurga leptonyx*; **F** – *Mirounga leonina.*



Source: M. JATOBÁ – SeCom - UNB (2017).

Despite the geographical isolation, Antarctica is not protected from the impacts of anthropogenic activities and climate change effects. Human activities such as oil prospection, overfishing, pollution, and the constant release of greenhouse gases all exert pressure on the Antarctic ecosystems (DINASQUET et al., 2018; WAUCHOPE; SHAW; TERAUDS, 2019). Between 1908 and 1943, seven countries claimed Antarctic territories, to avoid conflicts and further territorial claims, the Antarctic Treaty was created in 1959. The Treaty sets aside disputes over territorial claims and promotes peaceful use and scientific cooperation in the region (DODDS, 2010). Later, the Protocol on Environmental Protection to the Antarctic Treaty was signed in Madrid on October 4, 1991. The protocol established the continent as an International Natural Reserve devoted to peace and science. The protocol provided environmental protection, including a ban on mining and mineral exploration; the intentional introduction of nonnative species; strict regulations on disturbance to native species; waste management; and environmental impact assessment requirements (WAUCHOPE; SHAW; TERAUDS, 2019).

The severe environmental conditions are the bottleneck for studying organisms derived from polar regions. The cost of logistics is high, and specialized equipment is required to explore the poles. The climate conditions make it difficult to access the poles but do not limit biological communities' development (SOLDATOU; BAKER, 2017). However, behind the white and frozen landscapes, the Antarctic marine environment harbors a rich and underexplored biodiversity with unmeasurable biotechnological potential. To survive and overcome abiotic and ecological pressures such as food and space competition, predation, high UV ray exposure and low temperatures, polar organisms evolved different adaptation strategies, including the production of secondary metabolites (LEBAR; HEIMBEGNER; BAKER, 2007; RANGEL et al., 2020; TEIXEIRA et al., 2021).

Recent numbers demonstrate the growing interest of natural products scientists in marine-derived molecules. For example, only in 2020, 1407 new compounds were isolated from marine organisms while only 332 were reported in 1984 (CARROL et al., 2021). According to a literature review, until 2016, about 30 000 publications regarding marine natural products have been listed in the MarinLit database, but only 3% of the total reported natural products from cold environments (SOLDATOU; BAKER, 2017). (BLUNT et al., 2017; CARROLL et al., 2021). Despite the lower number of studies when compared to tropical regions, Antarctica represents a hotspot of bioprospection. In 2013, when then last survey was made there were patents registered for 439 Antarctic species highlighting the biotechnological potential of polar organisms (NÚÑEZ-PONS et al., 2020).

1. 2 The Brazilian Antarctic Program and the University of São Paulo

Brazil signed the Antarctic Treaty in 1975, and with the establishment of the Brazilian Antarctic Program (PROANTAR) the Brazilian Antarctic Missions began in 1982. Later, in 1983, these efforts enabled the country to be considered as a consultant member of the Antarctic Treaty. Since then, Brazil participates in the decision-making and policies regarding the future of the continent (MARINHA DO BRASIL, 2016).

The first Brazilian Antarctic expedition (OPERANTAR I) was carried out on board of the oceanographic survey ship Barão de Tefé (Marinha do Brasil) and on the oceanographic ship Prof. Wladimir Besnard from the Institute of Oceanography of the University of São Paulo (MARINHA DO BRASIL, 2016). The oceanographic ship Prof. Wladimir Besnard (Fig. 3) was built with funds provided by the Brazilian Federal agencies and State government of São Paulo. Thus, the University of São Paulo was pioneer in the Antarctic expeditions and until now has contributed to the advance of Antarctic science.



Figure 3. Oceanographic ship Prof. Wladimir Besnard.

Source: https://www.io.usp.br/index.php/embarcacoes/n-oc-prof-w-besnard.html

The Brazilian Antarctic Base, Comandante Ferraz (Fig. 4A-B), was established in Admiralty Bay on King's George Island in 1984 and served as support for the expeditions until February 2012, when the structure was consumed by fire. After eight years since the accident, a new base was inaugurated in 2020 (Fig. 4C-D).

The Laboratory of Organic Chemistry of the Marine Environment has integrated two field expeditions to the continent, the first in 2015, when Dr. Hosana Maria Debonsi initiated the study of natural products derived from Antarctic seaweeds and their endophytic fungi, and the second in 2017, when samples of *P. antarcticus* used in this work were collected. The seaweed collection and isolation of fungi were conducted on board the polar ship Almirante Maximiano H-41 (Fig. 4E) with the logistics support of the oceanographic ship Ary Rongel H-44 (Fig. 4F).

Figure 4. Brazil in Antarctica. **A-B** – Old Antarctic base; **C-D** – New Antarctic base; **E** – Polar ship Almirante Maximiano (H-41); **F** – Oceanographic ship Ary Rongel.



Source: SeCom – CIRM (2017); SeCIRM: https://www.marinha.mil.br/secirm/ptbr/proantar/nova-estacao (2019).

1. 3 Bioprospecting to Protect

The Antarctic Peninsula is one of the world's fastest warming regions. This environmental change has an impact on the planet's climate and will result in the loss of unknown biodiversity and imbalance in the continent's biological communities (SIEGERT et al., 2019). In this scenario, bioprospecting of polar organisms is essential to their conservation. Thus, continuous research is required to investigate Antarctic life forms and their biotechnological potential. The majority of Antarctic derived natural products are extracted from marine organisms and, despite the growing number of reports, cold-water organisms are still underexplored (DOS SANTOS et al., 2021; SOLDATOU; BAKER, 2017).

The year 2021 started the United Nations Decade of Ocean Science for Sustainable Development (2021 – 2030) which targets the creation of a cooperative framework to benefit both ocean ecosystems and society through the achievements of ocean science. In addition, on July 8^{th,} 2021, the National Geographic Society recognized the surrounding waters of the Antarctic continent as the fifth ocean on Earth. In the present global context, there is no better opportunity to deepen our knowledge of the polar waters and gather resources from governments, industry, and academia to explore the hidden chemistry and biotechnological potential of polar marine organisms (DOS SANTOS et al., 2021).

This work aims to contribute to the University of São Paulo commitment to the United Nations Sustainable Development Goals (SDG's) agenda. Among the 17 SDG's, this work will contribute to numbers 3 ("Good Health and Well-Being") and 14 ("Life Below Water").

1. 4 Antarctic Natural products

Although the oceans occupy two-thirds of the earth's surface, marine biodiversity remains an untapped source of new natural products. Systematic investigations of marine natural products began only 50 years ago. However, marine natural products have demonstrated high diversity and structural complexity, which are not found in terrestrial organisms (CARROLL et al., 2021; IOANNOU; ROUSSIS, 2009).

Seaweeds have been used in food, traditional medicine, and agriculture since ancient times, and they were among the first marine organisms to be chemically analyzed (IOANNOU; ROUSSIS, 2009; MILLEDGE; NIELSEN; BAILEY, 2016). Secondary metabolite production is directly related to abiotic factors in both the marine and terrestrial environments (TEIXEIRA et al., 2019a). In this sense, the seas and oceans present a number of adverse conditions, such as changes in salinity, acidity, temperature, ultraviolet radiation incidence, and nutrient competition, that make this environment an excellent trigger for the production of differentiated metabolites (CONTE et al., 2021). The polar regions, which include the Arctic and Antarctica, are considered the most extreme on earth (LIU et al., 2013). As a result, there is a limitation in the development of biological communities, and their biodiversity is dominated by marine organisms and microorganisms (GODINHO et al., 2013).

The Antarctic marine environment is characterized by high biological activity and biogeochemical cycling (BALDI et al., 2010). The intensity of biological activity and nutrient cycling in this environment is directly related to the high primary production that occurs during the summer (DUCKLOW et al., 2008; FONDA UMANI et al., 2005). Due to the isolation caused by the Antarctic Circumpolar Current, the benthic communities in the region are characterized by a high degree of endemism and the presence of cold-adapted species (WIENCKE et al., 2007).

In the Antarctic marine environment, seaweeds are important primary producers and plays an essential role in carbon cycling (NĘDZAREK, A., RAKUSA-SUSZCZEWSKI, 2004). In addition to primary production, seaweeds shelter a variety of associated organisms, including fish, invertebrates and microorganisms such as bacteria and fungi (LOQUE et al., 2010).

To survive the harsh conditions of the marine environment, seaweeds have developed/improved several defense strategies. Regarding secondary metabolites, these adaptations resulted in the production of diverse molecules biosynthesized by different metabolic pathways (IOANNOU; ROUSSIS, 2009). For example, to modulate microbial colonization on its surface and avoid herbivory, seaweeds produce compounds capable of balancing these ecological relationships and ensuring their survival (PERSSON et al., 2011). Abiotic factors such as increasing or decreasing temperature can also influence the metabolic profile displayed by macroalgae (FARIMAN; SHASTAN; ZAHEDI, 2016). For example, previous studies indicated the relationship between the decrease in water temperature and the increase in the production of polyunsaturated fatty acids (FARIMAN; SHASTAN; ZAHEDI, 2016; NOMURA et al., 2013).

In addition to the production of secondary metabolites, the association between seaweeds and microorganisms might characterize another form of defense and adaptation (SURYANARAYANAN, 2012). Marine fungi have been recognized as a prolific source of bioactive compounds. In the marine environment, they occur as spores, fragments of hyphae and mycelia and are classified into two distinct groups: obligate marine fungi and facultative marine fungi (JONES; KOHLMEYER; KOHLMEYER, 1980). Obligate marine fungi are those that grow and sporulate exclusively in a marine or estuarine habitat, while marine facultative fungi are those found in freshwater or terrestrial environments, which are able to grow and possibly also sporulate in the marine environment (PANG et al., 2016).

In polar regions such as Antarctica, fungi are further classified as psychrophilic and psychrotolerant (HARDING et al., 2011). The endemic Antarctic fungi are classified as true psychrophilic because they are capable of growing and sporulate exclusively in the Antarctic environment. The so-called psychrotolerant are cosmopolitan ecotypes that resulted from an adaptation to the climatic conditions of Antarctica, thus being able to develop in the region (ROSA et al., 2019b).

Macroalgae are the second largest source of fungi in the marine environment, evidencing the ecological relationship between these two groups of organisms (BUGNI; IRELAND, 2004). The fungi that colonize the algal surface are called epiphytic and those capable of colonizing the tissues and internal parts of the algae are then called endophytic (SURYANARAYANAN, 2012). By definition, endophytic fungi are those that colonize or spend part of their life cycle inside the host tissue, in a relationship that can range from symbiotic-mutualistic to latent-pathogenic (BUGNI; IRELAND, 2004; JONES; KOHLMEYER; KOHLMEYER, 1980; SURYANARAYANAN, 2012).

Studies related to Antarctic seaweeds and their symbionts are limited by logistics and safety in the face of extreme conditions in these environments. A few studies describe the diversity and biotechnological potential of the fungal community associated with the Antarctic seaweeds *Acrosiphonia arcta*, *Adenocystis utricularis*, *Monostroma hariotii*, *Palmaria decipiens*, *Ulva intestinalis*, *Desmarestia menziesii* and *Phaeurus antarcticus*, however, the methodologies used in these works did not target the isolation of endophytic fungi. Furthermore, only the potential of crude extracts was investigated with no records of isolated compounds (FURBINO et al., 2014; GODINHO et al., 2013).

1. 4. 1 Natural products from Antarctic seaweed

Seaweeds are a rich source of secondary metabolites, including antibacterial, antifungal, antiparasitic and photoprotective compounds (FALKENBERG et al., 2019; SHANNON; ABU-GHANNAM, 2019). *Plocamium cartilagineum* is a red macroalga species widely distributed on the Antarctic Peninsula. Previous chemical investigations

of this macroalga led to the identification of different chemogroups (YOUNG et al., 2013). In a recent report, Shilling and co-workers (2019) isolated nine monoterpenes from three chemogroups of this alga, including four new compounds named anverenes B–E (Fig. 5). All isolated compounds were submitted to clonogenic survival assay using a human cervical cancer cell line (HeLa). Anverene D was the most active compound with IC₅₀ value of 1.19 (SHILLING et al., 2019a).

Tavares and co-workers (2020) described the isolation of fucoxanthin from *Desmarestia anceps*. Fucoxanthin presented UVA and UVB absorption, photostability when incorporated into a sunscreen formulation, and antioxidant activity after UVA induction of ROS production on reconstructed human skin. This compound presented photoprotective properties and potential to be used as a UV-booster on sunscreens (TAVARES et al., 2020a).





1. 4. 2 Natural products from Antarctic marine fungi

In the period ranging from January 2018 to December 2021, fungi were the second most prospected Antarctic organism, highlighting the interest of natural products researchers in this new reservoir of bioactive molecules. Herein examples of metabolites isolated from Antarctic derived fungi and their bioactivity reported in the

review article Natural Products from Poles: Structural diversity and biological activities are described (DOS SANTOS et al., 2021).

The chemical investigation of the Antarctic fungus *Aspergillus sydowii* SP-1 led to the isolation of the new alkaloid acremolin C together with the four known compounds cyclo-(*L*-Trp-*L*-Phe), (4-hydroxyphenylacetic acid, (7*S*)-(+)-hydroxysydonic acid, and (7*S*,11*S*)-(+)-12-hydroxysydonic acid (Fig. 6). These compounds were screened against *Staphylococcus aureus*, *Staphylococcus epidermidis*, methicillin resistant *S. aureus* (MRSA), and methicillin-resistant *S. epidermidis* (MRSE). Acremolin C presented inhibition activities against MRSA and MRSE with MIC values of 32 and 16 µg/mL, respectively, while compounds cyclo-(*L*-Trp-*L*-Phe), 4-hydroxyphenylacetic acid, and (7*S*,11*S*)-(+)-12-hydroxysydonic acid presented higher inhibition effects against MRSA and MRSE with MIC values ranging from 0.5 to 1 µg/mL (LI et al., 2018).

A fungal isolate from marine sediment collected from Prydz Bay, Antarctica was identified as a *Penicillium crustosum* strain which produced the new polyene compound (2E,4E,6E,8E)-10-hydroxyundeca-2,4,6,8-tetraenoic acid, and a new diketopiperazine named fusaperazine F, together with the known compounds *cis*-bis-(methylthio)-silvatin and xylariolide D (Fig. 6). Fusaperazine F exhibited bioactivity against K562 cells, with an IC₅₀ value of 12.7 μ M (LIU et al., 2019).

A strain of *Aspergillus insulicola* HDN151418 was isolated from an unidentified sponge sample collected 410 m deep from Prydz Bay, Antarctica. Using a UV-HPLC guided approach, three new tripeptides named sclerotiotide M, sclerotiotide N, and sclerotiotide O were isolated from this fungus together with the known compounds, sclerotiotide L, sclerotiotide F and sclerotiotide B (Fig. 6). Sclerotiotide M presented potent antimicrobial activity towards *Bacillus cereus, Proteus mirabilis, Mycobacterium phlei, Edwardsiella tarda* and *B. subtilis* with MIC values ranging from 1.56–6.25 μ M. Sclerotiotide N was most active against *E. tarda* with MIC value of 1.56 μ M (SUN et al., 2020).

A new polyketide named 9-*O*-methylneuchromenin, together with the four known compounds neuchromenin, asterric acid, myxotrichin C, and deoxyfunicone (Fig. 6) were isolated from the fungus *Penicillium glabrum* (SF-7123). The compounds 9-*O*-methylneuchromenin, asterric acid and myxotrichin C presented inhibitory activity of nitric oxide (NO) in lipopolysaccharide (LPS)-stimulated BV2 microglial cells, with IC_{50} values of 2.7 μ M, 28.1 μ M, and 10.6 μ M respectively. These compounds also

downregulated the production of NO in LPS-stimulated RAW264.7 macrophages. Additionally, 9-O-methylneuchromenin downregulated NO synthase and cyclooxygenase-2 and inflammatory pathways dependent on nuclear factor kappa B and protein kinase in BV2 and RAW264.7 cells. Myxotrichin C and deoxyfunicone were found to inhibit the protein tyrosine phosphate 1B with IC₅₀ values of 19.2 μ M and 24.3 μ M (HA et al., 2020).





asterric acid

myxotrichin C

deoxyfunicone

1. 5 The seaweed Phaeurus antarcticus

Phaeurus antarcticus Skottsberg (Fig. 7) is a brown seaweed endemic to the Antarctic Peninsula and adjacent islands south of the Antarctic convergence. It occurs in the coastal and sub-coastal areas at depths that vary between 8 and 10 meters. It belongs to the order *Desmarestiales*, family Desmarestiaceae, and genus *Phaeurus*. *P. antarcticus* is considered one of the most primitive species in its family due to the simple structure of its thallus (CLAYTON; WIENCKE, 1990).

Currently, there are few works describing the chemical composition of *P. antarcticus* (IKEN et al., 2009a; TEIXEIRA et al., 2019b), and the biological activity of isolated molecules from this alga have not been reported. Additionally, reports addressing the diversity of endophytic fungi associated with this seaweed and their biotechnological potential are scarce (FURBINO et al., 2014; GODINHO et al., 2013), highlighting the relevance of the chemical investigation and bioactivities of *P. antarcticus* and its endophytic fungi.

Figure 7. The seaweed Phaeurus antarcticus.



Source: GODINHO et al., (2013); ALGAEBASE: https://www.algaebase.org/search/genus/detail/?genus_id=42681.

1. 6 Metabolomic and chemometrics approaches

In natural products research metabolomics has been defined as the comprehensive and quantitative analysis of all metabolites produced by a given organism. The metabolomic approach is often used to target distinct chemical profiles and avoid the isolation of known compounds with previously described bioactivities (ZHAO; ZHANG; LI, 2018).

Metabolomic analysis is divided into targeted and non-targeted studies. In targeted studies, the search is focused on predefined metabolic pathways or

compound classes. On the other hand, in non-targeted studies, all the metabolites present in extracts/fractions will be recorded and often large datasets are generated. Data acquisition in natural products metabolomic studies commonly relies in hyphenated techniques such as gas chromatography coupled to mass spectrometry (GC-MS) and liquid chromatography coupled to mass spectrometry (LC-MS) or nuclear magnetic resonance (NMR) (ZHAO; ZHANG; LI, 2018).

Usually, non-targeted studies needed to be coupled to chemometric approaches such as multivariate analysis, or computational techniques such as the Global Natural Products Social Molecular Networking workflows to allow data mining and a comprehensive understanding of metabolic patterns (DEMARQUE et al., 2020; GRIFFITHS et al., 2010; YANG et al., 2013).

Chemometrics aims to extract chemically relevant information from data produced in chemical experiments relying on mathematical models. For example, a measurement of the chemodiversity of molecules produced by different organisms can be obtained by evaluating the size of the chemical space visualized by a multivariate analyses (MVA) such as the principal component analysis (PCA) (WOLFENDER et al., 2019).

Multivariate statistical analysis of the data generated from GC/LC-MS and NMR can be performed to visualize data patterns through unsupervised clustering such as PCA, using soft independent modelling by class analogy (SIMCA), or supervised clustering such as partial least squares (PLS), partial least squares discriminant analysis (PLS-DA) and orthogonal partial least square discriminant analysis (OPLS-DA). Additionally, multivariate models can find relations among correlated variables, as is often the case in metabolomics studies, and possess the ability to distinguish systematic variation from noise (ROBOTTI; MARENGO, 2016; WOLD; ESBENSEN; GELADI, 1987).

Supervised analysis such as OPLS-DA can be useful to discriminate between two classes, such as healthy versus diseased, allowing us to target disease biomarkers or drug targets for treatments. In natural products research, this approach can be used to discriminate between bioactive and inactive fractions and, thus, allows prioritizing the isolation of specific metabolites responsible for bioactivity (WOLD; ESBENSEN; GELADI, 1987).

An illustration of the possible applications for each multivariate models is shown in figure 8.

PCA:	SIMCA:	PLS-DA /OPLS-DA:	O2-PLS
Overview	Classification	Discrimination	Regression
Trends	Pattern recognition	Discriminating	Comparing blocks of omics data
Outliers	Diagnostics	between groups	
Quality control	Heath/diseased	Biomarker candidates	Metabolomic vs proteomic vs genomic
Biological diversity	Toxicity mechanisms	Comparing studies of instrumentation	Correlation spectroscopy
Patient monitoring	Disease progression		
* * **		**************************************	

Figure 8. Multivariate models and their applicability.

Adapted from (WOLD; ESBENSEN; GELADI, 1987).

1. 7 Investigated biological activities

1.7.1 Photoprotection

The skin is the primary body protection against external damages such as mechanical lesions and microbial infections. When exposed to sunlight human skin triggers the production of vitamin D and β -endorphin which display beneficial health effects (MEAD, 2008).

However, the deleterious effect of continuous skin exposure to ultraviolet radiation (UVR) has been known since 1960. Unprotected or prolonged exposure to UVR leads to numerous changes in the skin, such as actinic keratosis, which has been identified as a precursor of non-melanoma skin cancer and squamous cell carcinoma. These conditions are related with UVA radiation, because of its ability to penetrate deeper in the skin (RAFFA et al., 2019).

Concerns regarding the effects of global warming have also been discussed in the context of skin related diseases since there is decrease of the stratospheric ozone layer and consequently an increase of UVR reaching the Earth's surface. The UV portion of the magnetic spectrum includes long wave UVA radiation (320-400 nm), medium wave UVB (280-320) and short wave UVC radiation (100-280 nm) (MU et al., 2021; SLOMINSKI; PAWELEK, 1998; URBAN et al., 2016).

In this context, sunscreens formulations represent an additional protection against UVR damage. However, recent studies have demonstrated the toxic potential of UV filters to human health, including DNA damage, penetration into placenta, alteration of spermatozoids and endocrine functions (JESUS et al., 2022).

Additionally, UV filters such as benzophenone-3 and octyl methoxycinnamate are harmful to marine organisms, especially corals. Benzophenone-3 was considered toxic to larval forms of the coral *Stylophora pistillata* and cytotoxicity was observed to other six coral species (DOWNS et al., 2016).

Compounds derived from natural sources have been studied to as photoprotective components against the damage produced by UV radiation. Marine derived compounds such as mycosporins, alkaloids, isocoumarins (present in fungi), mycosporin-like amino acids (MAAs, found in cyanobacteria, algae, and animals) have stood out as new candidates for sunscreens formulations and represent a source of new photoprotective compounds (MACIEL et al., 2018; SINGH et al., 2017; THIESEN et al., 2017).

1. 7. 2 Neutrophils as new drugs target for chronic inflammatory diseases

Neutrophils are polymorphonuclear and phagocytic leukocytes that function as the first line of defense against pathogenic organisms. In fact, they are the most abundant leukocytes in blood, and, from the circulation, they are mobilized to sites of inflammation and/or infection (ROSALES, 2020). When their receptors interact with antigens in the pathogen's cell wall, mechanisms such as the production of reactive oxygen (ROS) species and the release of neutrophil extracellular traps are activated (NETs)(HIRSCHFELD et al., 2017).

However, in the chronic inflammatory process hyperactivity of neutrophils leads to increased ROS production, resulting in an imbalance in redox homeostasis. The deregulation in neutrophils functions promotes exacerbated oxidative stress and consequently irreversible cellular and tissue damage (AMULIC et al., 2012; MORTAZ et al., 2018).

In fact, both inefficiency and exacerbation of neutrophil responses contribute to the pathophysiology of several diseases. In particular, chronic inflammatory diseases such as systemic lupus erythematosus (SLE) (TOLLER-KAWAHISA et al., 2015). In this context, molecules with the ability to regulate neutrophil function such as ROS production might represent an alternative to the development of new therapeutic agents for this disease.

1.7.3 Leishmaniasis

Leishmaniasis is caused by more than twenty species of Leishmania and is transmitted to humans by the bite of infected female phlebotomine sandflies (TORRES-GUERRERO et al., 2017). The disease affects humans and animals, causing public health problems especially in underdeveloped and developing countries (DOS SANTOS VARJÃO et al., 2022).

There are three main forms of the disease: cutaneous leishmaniasis (CL), visceral leishmaniasis (VL), and mucocutaneous leishmaniasis (MCL). The most common form of leishmaniasis is CL, which is caused by the *L. amazonensis* species. Currently, CL affects 12 million people globally, and 2 million new cases occur each year. CL is endemic in almost 100 countries 350 million people are at risk of contracting the disease (DE VRIES; SCHALLIG, 2022).

Pharmacotherapy for the treatment of leishmaniasis is represented by a few chemotherapeutic agents, such as pentavalent antimonials, amphotericin B, paromomycin, and miltefosine, which have several limitations regarding toxicity and lack of efficacy in endemic areas (TORRES-GUERRERO et al. 2017; FALKENBERG et al. 2019).

Due to the toxicity of existing therapies, as well as the emergence of resistant forms of *Leishmania*, there is an increased need for the development of more effective and less toxic antileishmanial drugs.

1.7.4 Neosporosis

Neosporosis is caused by the etiological agent *Neospora caninum*. This parasite infects mammalian species, like cattle, sheep, goats, horses, and dogs. Neosporosis causes abortion in cattle and neuromuscular disorders in dogs. The economic loss related to neosporosis in milk and livestock production is estimated at billions of dollars per annum. In addition, despite the efforts of industry and scientific community *Neosporosis* still does not have a specific treatment. For this reason, when animals

are diagnosed with Neosporosis they are sacrificed to avoid a disease outbreak (PEREIRA et al. 2017).

The investigation of natural products as anti-*Neospora* agents is very limited and has been focused on plant extracts (LEESOMBUN; BOONMASAWAI; NISHIKAWA, 2017; SEO et al., 2013). The first report of marine-derived compounds against *N. caninum* was published by our research group and revealed that the Antarctic seaweed *D. antarctica* presented anti-Neospora activity, highlighting the potential of marine-derived natural products in the prospection of new anti-Neospora treatments (DOS SANTOS et al., 2020).

1.7.5 Antibacterial and antibiofilm

Antibiotic resistance is one of the biggest threats to global health. Despite the fact that antibiotic resistance occurs naturally, this process has been accelerated by the misuse of antibiotics in human and animal health. The advent of multidrug resistant bacteria is rapidly rising, causing diseases such as severe pneumonia, tuberculosis, blood poisoning, gonorrhea, and food borne diseases. Due to the new resistance mechanisms evolved by bacteria, these diseases are becoming harder, and sometimes impossible to treat (ASLAM et al., 2018; WHO, 2022).

Biofilm forming bacteria are a major public health concern due to biofilm's ability to withstand external stresses and contribute to the persistence of chronic infections (DE LA FUENTE-NÚÑEZ et al., 2013). Biofilms are immobile microbial communities which can colonize and grow on different surfaces such as medical implants and catheters. In biofilms, bacterial populations are crammed by extra cellular matrix (ECM) which possesses bacterial secreted polymers such as exopolysaccharides (EPS), extracellular DNA, proteins amyloidogenic proteins (SHARMA; MISBA; KHAN, 2019; WHITCHURCH et al., 2002).

It is estimated that 80% of chronic and recurrent microbial infections are caused by bacterial biofilms. Microbial biofilms display different resistance mechanisms when not observed in planktonic cells, including lower cell permeability, efflux pumps, drug modifying enzymes and drug neutralizing proteins. These characteristics make the search for antibiofilm compounds an important initiative in order to aid in the fight against microbial resistance (SHARMA; MISBA; KHAN, 2019).

1. 7. 6 Aedes aegypti control

The *Aedes aegypti* mosquito is the vector responsible for the transmission of viral pathogens such as dengue, chikungunya, yellow fever and zika viruses. This species, native to Africa, is now distributed to all continents (KRAEMER et al., 2015; MCGREGOR; CONNELLY, 2021).

In Brazil, this mosquito has been responsible for the outbreaks of dengue, chikungunya, and zika viruses. The country has been facing an uninterrupted dengue epidemic since 1986 (VALLE; NACIF PIMENTA; AGUIAR, 2016). The WHO recommends that since there is no treatment for the viral diseases transmitted by *A. aegypti*, combating the vector is the most efficient strategy for controlling outbreaks (BENELLI, 2015).

Recently, the Brazilian National Dengue Control Program has promoted the replacement of highly toxic synthetic insecticides (organophosphates, pyrethroids and carbamates) for more environmentally friendly compounds, such as pyrethroids (cypermethrin and deltamethrin). However, they still display toxicity to the environment (PILON et al., 2022). In this context, we aimed to prospect new insecticidal compounds from Antarctic marine organisms in order to contribute to the development of safer and ecofriendly alternatives to those currently on the market.

1.8 Preface

This doctoral thesis is divided into four chapters. Chapters I and III were developed in the School of Pharmaceutical Sciences of Ribeirão Preto – University of São Paulo, Brazil. Chapters II and IV were developed in the Strathclyde Institute of Pharmacy and Biomedical Sciences – University of Strathclyde, United Kingdom.

Parts of the introduction of this thesis were published as a review article "Natural Products from the Poles: Structural Diversity and Biological Activities" in the Brazilian Journal of Pharmacognosy.

Chapter I describes the evaluation of the antibacterial, antiparasitic, and insecticidal potential of the seaweed *Phaeurus antarcticus* and the annotation of bioactive metabolites through the combination of multivariate analysis and GC-MS Molecular Networking.

Chapter II provides an insight into the spatial variability of secondary metabolite production and the antibiofilm activity of *P. antarcticus*, through the combination of proton nuclear magnetic resonance (¹H NMR) and high-resolution mass spectrometry (HRMS) data and multivariate analysis.

Chapter III presents the isolation of photoprotective, immunomodulatory, and antileishmanial compounds from the endophytic fungi *Penicillium purpurogenum*.

Chapter IV reports the metabolomic profiling of endophytic fungi isolated from *P. antarcticus* collected in different locations in the South Shetland Islands and the isolation of antibiofilm compounds from the fungus *Epicoccum nigrum*.

5.5 CONCLUSION

A total of 27 fungal strains were isolated from the inner tissues of the seaweed Phaeurus antarcticus collected in different locations across the Antarctic Peninsula. Sequencing of the ITS region led to the identification of fungi belonging to the Arthrinium, Cladosporium, Curvularia, Epicoccum, Nigrospora, Penidiella, Penicillium and Periconia genera. Additional sequencing experiments of the ACT region led to the identification of six Cladosporium species associated with this seaweed. A chemical profiling of 21 strains was achieved by proton NMR, HRMS and multivariate analysis. The strains identified as Penidiella aggregata and Cladosporium halotolerans presented unique chemical constituents and appeared as outliers in the PCA scatter plot. In the PCA loading scatter plot was possible to target the unique secondary metabolites responsible for the chemical distinction. In the C. halotolerans extracts, nitrogen containing compounds were annotated (rigidiusculamide A, huaspenone C, stachyline A, monascuskaochroman). In the Penidiella extracts, a polyketide, (sporminarin A), was annotated as discriminant metabolite. Despite the unique chemical profiles, in the antibiofilm evaluation Penidiella and C. halotolerans extracts were not among the most active samples against methicillin and oxacillin resistant S. aureus. The antibiofilm assays revealed that P. terrigenum, C. perangustum (LMC 1016) and E. nigrum extracts exhibited the highest antibacterial and antibiofilm effects.

A chemotaxonomic study was conducted with the *Cladosporium* isolates using proton NMR and HRMS data combined to the molecular identification results. A PLS-DA model was constructed to visualize the distribution of metabolites among the six different species of *Cladosporium*. Results revealed that the clustering observed in the phylogenetic tree was similar to the clustering observed in the PLS-DA model, indicating that the closest related strains also presented similar chemical profiles. These findings are an important contribution to the understanding of chemosystematics of Antarctic derived *Cladosporium* and could be used to guide future bioprospection and ecological investigations.

In the search for antibacterial and antibiofilm compounds, the fungus *E. nigrum* presented a more diverse chemical profile when compared to *P. terrigenum* and *C. perangustum* (LMC1016). For this reason, the strain was selected for large scale fermentation and isolation of metabolites. The purification of bioactive fractions led to the isolation of 4 metabolites, epicorazine A (9), epicorazine B (10),

epicorazine C (12), and a novel terpene (11). Epicorazine A – C exhibited MIC and MBEC values of 50 μ g/mL, 200 μ g/mL and 25 μ g/mL, respectively, but no effect on post-biofilm viability was observed. Compound 11 presented MIC and MBEC values of 25 μ g/mL and was able to inhibit 100% of post-biofilm viability at a concentration of 100 μ g/mL.

In conclusion, fungi endophytes associated with the seaweed *P. antarcticus* presented a diverse chemical profile, that could source novel antimicrobial and antibiofilm compounds. *E. nigrum* produced interesting sulfur containing metabolites with potential biotechnological applications in the pharmaceutical industry. Thus, despite the highly cytotoxic effects of the isolated compounds reported in the literature, *in vivo* models are encouraged to assess the toxic potential of epicorazines.

6. REFERENCES

AGRAWAL, Shivankar; ADHOLEYA, Alok; BARROW, Colin J.; DESHMUKH, Sunil Kumar. Marine fungi: An untapped bioresource for future cosmeceuticals. **Phytochemistry Letters**, v. 23, p. 15–20, 2018.

AKSENOV, Alexander A. et al. Auto-deconvolution and molecular networking of gas chromatography–mass spectrometry data. **Nature Biotechnology**, v. 39, n. 2, p. 169–173, 2021.

AL-FATIMI, M. A. A.; JÜLICH, W. D.; JANSEN, R.; LINDEQUIST, U. Bioactive components of the traditionally used mushroom *Podaxis pistillaris*. **Evidence-based Complementary and Alternative Medicine**, v. 3, n. 1, p. 87–92, 2006.

AMICO, Vincenzo. Marine brown algae of family Cystoseiraceae: Chemistry and chemotaxonomy. **Phytochemistry**, v. 39, n. 6, p. 1257–1279, 1995.

AMULIC, Borko; CAZALET, Christel; HAYES, Garret L.; METZLER, Kathleen D.; ZYCHLINSKY, Arturo. Neutrophil Function: From Mechanisms to Disease. **Annual Review of Immunology**, 2012.

ANDRADE, Micássio F.; KABEYA, Luciana M.; AZZOLINI, Ana Elisa C. S.; SANTOS, Everton O. L.; FIGUEIREDO-RINHEL, Andréa S. G.; PARIS, Márcio R. P.; EMERY, Flávio S.; PUPO, Mônica T.; LUCISANO-VALIM, Yara M. 3-Phenylcoumarin derivatives selectively modulate different steps of reactive oxygen species production by immune complex-stimulated human neutrophils. **International Immunopharmacology**, v. 15, n. 2, p. 387–394, 2013.

ASLAM, Bilal et al. Antibiotic resistance: a rundown of a global crisis. **Infection and Drug Resistance**, v. Volume 11, p. 1645–1658, 2018.

AVILA, Conxita; TABOADA, Sergi; NÚÑEZ-PONS, Laura. Antarctic marine chemical ecology: What is next? Marine Ecology, 2008.

BALDI, Franco; MARCHETTO, Davide; PINI, Francesco; FANI, Renato; MICHAUD, Luigi; LO GIUDICE, Angelina; BERTO, Daniela; GIANI, Michele. Biochemical and microbial features of shallow marine sediments along the Terra Nova Bay (Ross Sea, Antarctica). **Continental Shelf Research**, v. 30, n. 15, p. 1614–1625, 2010.

BECERRA, Monica et al. Antileishmanial activity of fucosterol recovered from *Lessonia vadosa* Searles (Lessoniaceae) by SFE, PSE and CPC. **Phytochemistry Letters**, 2015.

BENELLI, Giovanni. Plant-borne ovicides in the fight against mosquito vectors of medical and veterinary importance: a systematic review. **Parasitology Research**, v. 114, n. 9, p. 3201–3212, 2015.

BENELLI, Giovanni; PAVELA, Roman; DRENAGGI, Ettore; DESNEUX, Nicolas; MAGGI, Filippo. Phytol, (E)-nerolidol and spathulenol from Stevia rebaudiana leaf essential oil as effective and eco-friendly botanical insecticides against *Metopolophium dirhodum*. Industrial Crops and Products, 2020.

BENSCH, K.; BRAUN, U.; GROENEWALD, J. Z.; CROUS, P. W. The genus *Cladosporium*. **Studies in Mycology**, v. 72, p. 1–401, 2012.

BENSON, Dennis A.; CAVANAUGH, Mark; CLARK, Karen; KARSCH-MIZRACHI, Ilene; OSTELL, James; PRUITT, Kim D.; SAYERS, Eric W. GenBank. **Nucleic Acids Research**, v. 46, n. D1, p. D41–D47, 2018.

BERNARDINI, Giulia; MINETTI, Mariagiulia; POLIZZOTTO, Giuseppe; BIAZZO, Manuele; SANTUCCI, Annalisa. Pro-Apoptotic activity of French polynesian *Padina pavonica* extract on human osteosarcoma cells. **Marine Drugs**, v. 16, n. 12, 2018.

BIBER, Knut; OWENS, Trevor; BODDEKE, Erik. What is microglia neurotoxicity (Not)? GLIA, 2014.

BLUNT, John W.; COPP, Brent R.; KEYZERS, Robert A.; MUNRO, Murray H. G.; PRINSEP, Michèle R. Marine natural products. **Natural Product Reports**, v. 34, n. 3, p. 235–294, 2017.

BOJARSKA, Joanna et al. Cyclic dipeptides: The biological and structural landscape with special focus on the anti-cancer proline-based scaffold. **Biomolecules**, 2021.

BRAUN, Glaucia H. et al. Evaluation of antileishmanial activity of harzialactone a isolated from the marine-derived fungus *Paecilomyces* sp. **Natural Product Research**, 2021.

BUCHALSKA, Marta; KRAS, Gabriela; OSZAJCA, Marcin; ŁASOCHA, Wiesław;

MACYK, Wojciech. Singlet oxygen generation in the presence of titanium dioxide materials used as sunscreens in suntan lotions. **Journal of Photochemistry and Photobiology A: Chemistry**, 2010.

BUGNI, Tim S.; IRELAND, Chris M. Marine-derived fungi: A chemically and biologically diverse group of microorganisms. **Natural Product Reports**, v. 21, n. 1, p. 143–163, 2004.

CANTRELL, Charles L.; JONES, A. Maxwell P.; ALI, Abbas. Isolation and Identification of Mosquito (Aedes aegypti) Biting-Deterrent Compounds from the Native American Ethnobotanical Remedy Plant *Hierochloë odorata* (Sweetgrass). **Journal of Agricultural and Food Chemistry**, 2016.

CARBONE, Ignazio; KOHN, Linda M. A method for designing primer sets for speciation studies in filamentous ascomycetes. **Mycologia**, 1999.

CARROLL, Anthony R.; COPP, Brent R.; DAVIS, Rohan A.; KEYZERS, Robert A.; PRINSEP, Michèle R. Marine natural products. **Natural Product Reports**, v. 38, n. 2, p. 362–413, 2021.

CASTENHOLZ, Richard W. Culturing Methods for Cyanobacteria. **Methods in Enzymology**, 1988.

CENTKO, Ryan M.; WILLIAMS, David E.; PATRICK, Brian O.; AKHTAR, Yasmin; GARCIA CHAVEZ, Miguel Angel; WANG, Yan Alexander; ISMAN, Murray B.; DE SILVA, E. Dilip; ANDERSEN, Raymond J. Dhilirolides E-N, meroterpenoids produced in culture by the fungus *Penicillium purpurogenum* collected in Sri Lanka: Structure elucidation, stable isotope feeding studies, and insecticidal activity. **Journal of Organic Chemistry**, 2014.

CHAMBERS, Henry F.; DELEO, Frank R. Waves of resistance: *Staphylococcus aureus* in the antibiotic era. **Nature Reviews Microbiology**, v. 7, n. 9, p. 629–641, 2009.

CHEN, Jiayun; LI, Hong; ZHAO, Zishuo; XIA, Xue; LI, Bo; ZHANG, Jinrong; YAN, Xiaojun. Diterpenes from the marine algae of the genus *Dictyota*. **Marine Drugs**, 2018.

CHEN, Zhihong; TRAPP, Bruce D. Microglia and neuroprotection. Journal of

Neurochemistry, 2016.

CHONG, Jasmine; WISHART, David S.; XIA, Jianguo. Using MetaboAnalyst 4.0 for Comprehensive and Integrative Metabolomics Data Analysis. **Current Protocols in Bioinformatics**, 2019.

CHOU, Ting Chao; TALALAY, Paul. Quantitative analysis of dose-effect relationships: the combined effects of multiple drugs or enzyme inhibitors. **Advances in Enzyme Regulation**, 1984.

CHRIST, Bastien; HÖRTENSTEINER, Stefan. Mechanism and Significance of Chlorophyll Breakdown. Journal of Plant Growth Regulation, 2014.

CLAYTON, M. N.; WIENCKE, C. The anatomy, life history and development of the Antarctic brown alga *Phaeurus antarcticus* (Desmarestiales, Phaeophyceae). **Phycologia**, v. 29, n. 3, p. 303–315, 1990.

CLEMENTINO, Leandro Costa; TORRES, Fabio Aurelio Esteves; VELASQUEZ, Angela Maria Arenas; VILLELA, Leonardo; MUTUE, Toyota Fujii; COLEPICOLO, Pio; GRAMINHA, Marcia A. S. Bioguided study of the Antarctic alga *Himantothallus grandifolius* (A. Geep & E.S. Geep) indicates 13E-Docosenamide as potential antileishmanial agent. **Journal of Applied Pharmaceutical Science**, v. 10, n. 12, p. 98–103, 2020. a.

CLEMENTINO, Leandro da Costa; ODA, Fernando Bombarda; TEIXEIRA, Thaiz Rodrigues; TAVARES, Renata Spagolla Napoleão; COLEPICOLO, Pio; SANTOS, André Gonzaga Dos; DEBONSI, Hosana Maria; GRAMINHA, Márcia A. S. The antileishmanial activity of the antarctic brown alga *Ascoseira mirabilis* Skottsberg. **Natural Product Research**, v. 6419, p. 1–5, 2020. b.

CLSI. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Approved Standard — Second Edition Serving the World 's Medical Science Community Through Voluntary Consensus. [s.l: s.n.]. v. 22, 2008.

CONTE, Mariarosaria; FONTANA, Elisabetta; NEBBIOSO, Angela; ALTUCCI, Lucia. Marine-derived secondary metabolites as promising epigenetic bio-compounds for anticancer therapy. **Marine Drugs**, 2021. DE FREITAS, Samantha C. et al. Bioactivity evaluation and composition of extracts from sub-Antarctic macroalgae *Mazzaella laminarioides* at distinct development phases. **Brazilian Journal of Botany**, v. 43, n. 4, p. 689–696, 2020.

DE LA FUENTE-NÚÑEZ, César; REFFUVEILLE, Fany; FERNÁNDEZ, Lucía; HANCOCK, Robert E. W. Bacterial biofilm development as a multicellular adaptation: Antibiotic resistance and new therapeutic strategies. **Current Opinion in Microbiology**, 2013.

DE MORAES, Josué; DE OLIVEIRA, Rosimeire N.; COSTA, Jéssica P.; JUNIOR, Antonio L. G.; DE SOUSA, Damião P.; FREITAS, Rivelilson M.; ALLEGRETTI, Silmara M.; PINTO, Pedro L. S. Phytol, a Diterpene Alcohol from Chlorophyll, as a Drug against Neglected Tropical Disease *Schistosomiasis Mansoni*. **PLoS Neglected Tropical Diseases**, 2014.

DE SOUSA, Jordana R. P.; GONÇALVES, Vívian N.; DE HOLANDA, Rodrigo A.; SANTOS, Daniel A.; BUELONI, Cinthia F. L. G.; COSTA, Adriana O.; PETRY, Maria V.; ROSA, Carlos A.; ROSA, Luiz H. Pathogenic potential of environmental resident fungi from ornithogenic soils of Antarctica. **Fungal Biology**, 2017.

DE VRIES, Henry J. C.; SCHALLIG, Henk D. Cutaneous Leishmaniasis: A 2022 Updated Narrative Review into Diagnosis and Management Developments. **American Journal of Clinical Dermatology**, v. 23, n. 6, p. 823–840, 2022.

DEFFIEUX, Gérard; FILLEAU, Marie José; BAUTE, Marie Antoinette; BAUTE, Robert. New antibiotics from the fungus *Epicoccum nigrum:* II. epicorazine A: Structure elucidation and absolute configuration. **The Journal of Antibiotics**, 1978.

DEFFIEUX, Gérard; FILLEAU, Marie José; BAUTE, Robert. New Antibiotics from the Fungus *Epicoccum Nigrum*. III. Epicorazine B: Structure Elucidation and Absolute Configuration. **The Journal of Antibiotics**, v. 31, n. 11, p. 1106–1109, 1978.

DEMARQUE, Daniel P.; DUSI, Renata G.; DE SOUSA, Francisco D. M.; GROSSI, Sophia M.; SILVÉRIO, Maira R. S.; LOPES, Norberto P.; ESPINDOLA, Laila S. Mass spectrometry-based metabolomics approach in the isolation of bioactive natural products. **Scientific Reports**, 2020.

DESBOIS, Andrew P.; SMITH, Valerie J. Antibacterial free fatty acids: Activities,

mechanisms of action and biotechnological potential. Applied Microbiology and Biotechnology, 2010.

DHINGRA, Sameer; RAHMAN, Nor Azlina A.; PEILE, Ed; RAHMAN, Motiur; SARTELLI, Massimo; HASSALI, Mohamed Azmi; ISLAM, Tariqul; ISLAM, Salequl; HAQUE, Mainul. Microbial Resistance Movements: An Overview of Global Public Health Threats Posed by Antimicrobial Resistance, and How Best to Counter. **Frontiers in Public Health**, v. 8, 2020.

DINASQUET, Julie; ORTEGA-RETUERTA, Eva; LOVEJOY, Connie; OBERNOSTERER, Ingrid. Editorial: Microbiology of the rapidly changing polar environments. **Frontiers in Marine Science**, v. 5, n. MAY, p. 4–6, 2018.

DODDS, Klaus. Governing Antarctica: Contemporary Challenges and the Enduring Legacy of the 1959 Antarctic Treaty. **Global Policy**, v. 1, n. 1, p. 108–115, 2010.

DOS SANTOS, Gustavo Souza et al. GC-MS Analysis, Bioactivity-based Molecular Networking and Antiparasitic Potential of the Antarctic Alga *Desmarestia antarctica*. **Planta Medica International Open**, v. 07, n. 03, p. e122–e132, 2020.

DOS SANTOS, Gustavo Souza; TEIXEIRA, Thaiz Rodrigues; COLEPICOLO, Pio; DEBONSI, Hosana Maria. Natural Products from the Poles: Structural Diversity and Biological Activities. **Revista Brasileira de Farmacognosia**, v. 31, n. 5, p. 531–560, 2021.

DOS SANTOS VARJÃO, Márcio Thomaz; DUARTE, Alysson Wagner Fernandes; ROSA, Luiz Henrique; ALEXANDRE-MOREIRA, Magna Suzana; DE QUEIROZ, Aline Cavalcanti. Leishmanicidal activity of fungal bioproducts: A systematic review. **Fungal Biology Reviews**, 2022.

DOWNS, C. A. et al. Toxicopathological Effects of the Sunscreen UV Filter, Oxybenzone (Benzophenone-3), on Coral Planulae and Cultured Primary Cells and Its Environmental Contamination in Hawaii and the U.S. Virgin Islands. **Archives of Environmental Contamination and Toxicology**, v. 70, n. 2, p. 265–288, 2016.

DOWNS, C. A.; DINARDO, Joseph C.; STIEN, Didier; RODRIGUES, Alice M. S.; LEBARON, Philippe. Benzophenone Accumulates over Time from the Degradation of Octocrylene in Commercial Sunscreen Products. **Chemical Research in Toxicology**,

2021..

DUCKLOW, Hugh W.; ERICKSON, Matthew; KELLY, Joann; MONTES-HUGO, Martin; RIBIC, Christine A.; SMITH, Raymond C.; STAMMERJOHN, Sharon E.; KARL, David M. Particle export from the upper ocean over the continental shelf of the west Antarctic Peninsula: A long-term record, 1992-2007. **Deep-Sea Research Part II: Topical Studies in Oceanography**, v. 55, n. 18–19, p. 2118–2131, 2008.

DUTRA, Luiz Antônio et al. Leishmanicidal activities of novel synthetic furoxan and benzofuroxan derivatives. **Antimicrobial Agents and Chemotherapy**, 2014.

FALKENBERG, Miriam et al. Bioactive compounds against neglected diseases isolated from macroalgae: a review. **Journal of Applied Phycology**, v. 31, n. 2, p. 797–823, 2019.

FAN, Yaqin; MA, Zhiheng; ZHANG, Yan; WANG, Yufei; DING, Yousong; WANG, Cong; CAO, Shugeng. Sulfur-Containing Compounds from Endophytic Fungi: Sources, Structures and Bioactivities. **Journal of Fungi**, v. 8, n. 6, p. 628, 2022.

FARIMAN, Gilan Attarn; SHASTAN, Salim Jangizehi; ZAHEDI, Mir Mehdi. Seasonal variation of total lipid, fatty acids, fucoxanthin content, and antioxidant properties of two tropical brown algae (*Nizamuddinia zanardinii* and *Cystoseira indica*) from Iran. **Journal of Applied Phycology**, v. 28, n. 2, p. 1323–1331, 2016.

FELSENSTEIN, Joseph. Confidence Limits on Phylogenies: An Approach Using the Bootstrap. **Evolution**, 1985.

FINDLAY, John A.; LI, Guoqiang; MILLER, J. David; WOMILOJU, Taiwo O. Insect toxins from spruce endophytes. **Canadian Journal of Chemistry**, 2003.

FONDA UMANI, S.; MONTI, M.; BERGAMASCO, A.; CABRINI, M.; DE VITTOR, C.; BURBA, N.; DEL NEGRO, P. Plankton community structure and dynamics versus physical structure from Terra Nova Bay to Ross Ice Shelf (Antarctica). **Journal of Marine Systems**, v. 55, n. 1–2, p. 31–46, 2005.

FRISVAD, Jens C.; ANDERSEN, Birgitte; THRANE, Ulf. The use of secondary metabolite profiling in chemotaxonomy of filamentous fungi. **Mycological Research**, v. 112, n. 2, p. 231–240, 2008.

FURBINO, Laura E. et al. Diversity Patterns, Ecology and Biological Activities of Fungal Communities Associated with the Endemic Macroalgae Across the Antarctic Peninsula. **Microbial Ecology**, v. 67, n. 4, p. 775–787, 2014.

FUSKA, J.; UHRÍN, D.; PROKSA, B.; VOTICKÝ, Z.; RUPPELDT, J. The structure of vermistatin, a new metabolite from *Penicillium vermiculatum*. **The Journal of Antibiotics**, v. 39, n. 11, p. 1605–1608, 1986.

GASPAR, Lorena R.; THARMANN, Julian; MAIA CAMPOS, Patricia M. B. G.; LIEBSCH, Manfred. Skin phototoxicity of cosmetic formulations containing photounstable and photostable UV-filters and vitamin A palmitate. **Toxicology in Vitro**, v. 27, n. 1, p. 418–425, 2013.

GODINHO, Valéria M. et al. Diversity and bioprospecting of fungal communities associated with endemic and cold-adapted macroalgae in Antarctica. **ISME Journal**, v. 7, n. 7, p. 1434–1451, 2013.

GRIFFITHS, William J.; KOAL, Therese; WANG, Yuqin; KOHL, Matthias; ENOT, David
P.; DEIGNER, Hans-Peter. Targeted Metabolomics for Biomarker Discovery.
Angewandte Chemie International Edition, v. 49, n. 32, p. 5426–5445, 2010..

GUO, Huijuan; SUN, Bingda; GAO, Hao; CHEN, Xulin; LIU, Shuchun; YAO, Xinsheng; LIU, Xingzhong; CHE, Yongsheng. Diketopiperazines from the Cordyceps -Colonizing Fungus *Epicoccum nigrum*. **Journal of Natural Products**, v. 72, n. 12, p. 2115–2119, 2009.

HA, Tran Minh; KIM, Dong Cheol; SOHN, Jae Hak; YIM, Joung Han; OH, Hyuncheol. Anti-inflammatory and protein tyrosine phosphatase 1b inhibitory metabolites from the antarctic marine-derived fungal strain *Penicillium glabrum* sf-7123. **Marine Drugs**, v. 18, n. 5, p. 1–15, 2020.

HARDING, Tommy; JUNGBLUT, Anne D.; LOVEJOY, Connie; VINCENT, Warwick F. Microbes in high arctic snow and implications for the cold biosphere. **Applied and Environmental Microbiology**, v. 77, n. 10, p. 3234–3243, 2011.

HARDT, Ingo H.; FENICAL, William; CRONIN, Greg; HAY, Mark E. Acutilols, potent herbivore feeding deterrents from the tropical brown alga, *Dictyota acutiloba*. **Phytochemistry**, 1996.

HARWOKO, Harwoko et al. Azacoccones F-H, new flavipin-derived alkaloids from an endophytic fungus *Epicoccum nigrum* MK214079. **Fitoterapia**, 2020.

HIRSCHFELD, Josefine; WHITE, Phillipa C.; MILWARD, Michael R.; COOPER, Paul R.; CHAPPLE, Iain L. C. Modulation of Neutrophil Extracellular Trap and Reactive Oxygen Species Release by Periodontal Bacteria. **Infection and Immunity**, v. 85, n. 12, 2017.

HUANG, Zhongjing; YANG, Ruiyun; GUO, Zhiyong; SHE, Zhigang; LIN, Yongcheng. A new naphtho-γ-pyrone from mangrove endophytic fungus ZSU-H26. **Chemistry of Natural Compounds**, v. 46, n. 1, p. 15–18, 2010.

HUNG, The Dang; HYE, Ja Lee; EUN, Sook Yoo; SHINDE, Pramod B.; YOON, Mi Lee; HONG, Jongki; DONG, Kyoo Kim; JUNG, Jee H. Anti-inflammatory constituents of the Red Alga *Gracilaria verrucosa* and their synthetic analogues. **Journal of Natural Products**, 2008.

IKEN, Katrin; AMSLER, Charles D.; AMSLER, Margaret O.; MCCLINTOCK, James B.; BAKER, Bill J. Field studies on deterrent properties of phlorotannins in Antarctic brown algae. **Botanica Marina**, v. 52, n. 6, p. 547–557, 2009. a.

IKEN, Katrin; AMSLER, Charles D.; AMSLER, Margaret O.; MCCLINTOCK, James B.; BAKER, Bill J. Field studies on deterrent properties of phlorotannins in Antarctic brown algae. **Botanica Marina**, 2009. b.

IOANNOU, Efstathia; QUESADA, Antonio; RAHMAN, M. Mukhlesur; GIBBONS, Simon; VAGIAS, Constantinos; ROUSSIS, Vassilios. Dolabellanes with antibacterial activity from the brown alga *Dilophus spiralis*. **Journal of Natural Products**, 2011.

IOANNOU, Efstathia; ROUSSIS, Vassilios. Natural products from seaweeds. *In*: **Plant-derived Natural Products: Synthesis, Function, and Application**. [s.l: s.n.]. p. 51–81.

JABER, Saif Aldeen Mohammad Fayiz. **Metabolomic profiling of antibiofilm compounds from fungal endophytes derived from scottish seaweeds**. 2022. University of Strathclyde, 2022.

JÉGOU, Camille; CULIOLI, Gérald; STIGER-POUVREAU, Valérie. Meroditerpene

from *Cystoseira nodicaulis* and its taxonomic significance. **Biochemical Systematics and Ecology**, v. 44, p. 202–204, 2012.

JESUS, Ana; SOUSA, Emília; CRUZ, Maria; CIDADE, Honorina; LOBO, José; ALMEIDA, Isabel. UV Filters: Challenges and Prospects. **Pharmaceuticals**, v. 15, n. 3, p. 263, 2022.

JIANG, Zhi-Dong; GERWICK, William H. Novel oxylipins from the temperate red alga *Polyneura latissima*: Evidence for an arachidonate 9(S)-lipoxygenase. **Lipids**, v. 32, n. 3, p. 231–235, 1997.

JONES, E. B. Gareth; KOHLMEYER, Erika; KOHLMEYER, Jan. Marine Mycology: The Higher Fungi. **Mycologia**, v. 72, n. 3, p. 650, 1980..

JUNG, Misong; KYOUNG, Hwa Jang; KIM, Bora; BONG, Ho Lee; BYOUNG, Wook Choi; OH, Ki Bong; SHIN, Jongheon. Meroditerpenoids from the brown alga *Sargassum siliquastrum.* **Journal of Natural Products**, 2008.

KALYANARAMAN, Balaraman et al. Measuring reactive oxygen and nitrogen species with fluorescent probes: challenges and limitations. **Free Radical Biology and Medicine**, v. 52, n. 1, p. 1–6, 2012.

KASPERKIEWICZ, Kinga; ERKIERT-POLGUJ, Anna; BUDZISZ, Elzbieta. Sunscreening and Photosensitizing Properties of Coumarins and their Derivatives. Letters in Drug Design & Discovery, 2016.

KAZLAUSKAS, Rymantas; MULDER, John; MURPHY, PT; WELLS, RJ. New metabolites from the brown alga *Caulocystis cephalornithos*. **Australian Journal of Chemistry**, v. 33, n. 9, p. 2097, 1980.

KEJLOVÁ, K.; JÍROVÁ, D.; BENDOVÁ, H.; KANDÁROVÁ, H.; WEIDENHOFFER, Z.; KOLÁŘOVÁ, H.; LIEBSCH, M. Phototoxicity of bergamot oil assessed by in vitro techniques in combination with human patch tests. **Toxicology in Vitro**, 2007.

KHALIFA, Shaden A. M. et al. Marine Natural Products: A Source of Novel Anticancer Drugs. **Marine Drugs**, v. 17, n. 9, p. 491, 2019.

KIM, Dong-Cheol et al. Dihydroisocoumarin Derivatives from Marine-Derived Fungal Isolates and Their Anti-inflammatory Effects in Lipopolysaccharide-Induced BV2 Microglia. Journal of Natural Products, v. 78, n. 12, p. 2948–2955, 2015.

KIM, Dong-Cheol; LEE, Hee-Suk; KO, Wonmin; LEE, Dong-Sung; SOHN, Jae; YIM, Joung; KIM, Youn-Chul; OH, Hyuncheol. Anti-Inflammatory Effect of Methylpenicinoline from a Marine Isolate of *Penicillium* sp. (SF-5995): Inhibition of NFκB and MAPK Pathways in Lipopolysaccharide-Induced RAW264.7 Macrophages and BV2 Microglia. **Molecules**, v. 19, n. 11, p. 18073–18089, 2014.

KIM, Kyoung-Su; CUI, Xiang; LEE, Dong-Sung; SOHN, Jae; YIM, Joung; KIM, Youn-Chul; OH, Hyuncheol. Anti-Inflammatory Effect of Neoechinulin A from the Marine Fungus *Eurotium* sp. SF-5989 through the Suppression of NF-κB and p38 MAPK Pathways in Lipopolysaccharide-Stimulated RAW264.7 Macrophages. **Molecules**, v. 18, n. 11, p. 13245–13259, 2013.

KIMURA, Junji; MAKI, Noritsugu. New loliolide derivatives from the brown alga Undaria pinnatifida. Journal of Natural Products, 2002.

KIMURA, Yasuo; NAKAJIMA, Hiromitsu; HAMASAKI, Takashi. Sescandelin, a New Root Promoting Substance Produced by the Fungus, *Sesquicillium candelabrum*. **Agricultural and Biological Chemistry**, 1990.

KLEINWÄCHTER, PETER; DAHSE, HANS-MARTIN; LUHMANN, UDO; SCHLEGEL, BRIGITTE; DORNBERGER, KLAUSJÜRGEN. Epicorazine C, an Antimicrobial Metabolite from *Stereum hirsutum* HKI 0195. **The Journal of Antibiotics**, v. 54, n. 6, p. 521–525, 2001.

KOMIYA, Takashi et al. Phytol induces programmed cell death in human lymphoid leukemia Molt 4B cells. International Journal of Molecular Medicine, 1999.

KONG, Fandong; WANG, Yi; LIU, Peipei; DONG, Tianhan; ZHU, Weiming. Thiodiketopiperazines from the Marine-Derived Fungus *Phoma* sp. OUCMDZ-1847. **Journal of Natural Products**, v. 77, n. 1, p. 132–137, 2014.

KRAEMER, Moritz U. G. et al. The global distribution of the arbovirus vectors *Aedes aegypti* and *Ae. albopictus*. **eLife**, v. 4, 2015.

KUAI, Benke; CHEN, Junyi; HÖRTENSTEINER, Stefan. The biochemistry and molecular biology of chlorophyll breakdown. Journal of Experimental Botany, v. 69,

n. 4, p. 751–767, 2018.

KUMAR, S. Sadish; KUMAR, Y.; KHAN, M. S. Y.; GUPTA, V. New antifungal steroids from *Turbinaria conoides* (J. Agardh) Kutzing. **Natural Product Research**, v. 24, n. 15, p. 1481–1487, 2010.

LE LANN, Klervi; RUMIN, Judith; CÉRANTOLA, Stéphane; CULIOLI, Gérald; STIGER-POUVREAU, Valérie. Spatiotemporal variations of diterpene production in the brown macroalga *Bifurcaria bifurcata* from the western coasts of Brittany (France). **Journal of Applied Phycology**, v. 26, n. 2, p. 1207–1214, 2014.

LEBAR, Matthew D.; HEIMBEGNER, Jaime L.; BAKER, Bill J. Cold-water marine natural products. Natural Product Reports, 2007.

LEESOMBUN, Arpron; BOONMASAWAI, Sookruetai; NISHIKAWA, Yoshifumi. Effects of *Thai piperaceae* plant extracts on *Neospora caninum* infection. **Parasitology International**, v. 66, n. 3, p. 219–226, 2017.

LI, Jian; LIU, Shuchun; NIU, Shubin; ZHUANG, Wenying; CHE, Yongsheng. Pyrrolidinones from the ascomycete fungus *Albonectria rigidiuscula*. **Journal of Natural Products**, v. 72, n. 12, p. 2184–2187, 2009.

LI, Shan shan; LI, Jun; SUN, Jing; GUO, Ran; YU, Lan zhi; ZHAO, Yun fang; ZHU, Zhi xiang; TU, Peng fei. Berkeleyacetal C, a meroterpenoid isolated from the fungus *Penicillium purpurogenum* MHZ 111, exerts anti-inflammatory effects via inhibiting NFκB, ERK1/2 and IRF3 signaling pathways. **European Journal of Pharmacology**, v. 814, n. August, p. 283–293, 2017.

LI, Shangde; WEI, Meiyan; CHEN, Guangying; LIN, Yongcheng. Two new dihydroisocoumarins from the endophytic fungus *Aspergillus* sp. Collected from the South China Sea. **Chemistry of Natural Compounds**, 2012.

LI, Wen ting; LUO, Dan; HUANG, Jia ning; WANG, Lin lin; ZHANG, Feng guo; XI, Tao; LIAO, Jian min; LU, Yuan yuan. Antibacterial constituents from Antarctic fungus, *Aspergillus sydowi* SP-1. **Natural Product Research**, v. 32, n. 6, p. 662–667, 2018.

LIU, Cong Cong; ZHANG, Zhen Zhen; FENG, Yan Yan; GU, Qian Qun; LI, De Hai; ZHU, Tian Jiao. Secondary metabolites from Antarctic marine-derived fungus

Penicillium crustosum HDN153086. Natural Product Research, v. 33, n. 3, p. 414–419, 2019.

LIU, Jing-Tang; LU, Xiao-Ling; LIU, Xiao-Yu; GAO, Yun; HU, Bo; JIAO, Bing-Hua; ZHENG, Heng. Bioactive natural products from the Antarctic and Arctic organisms. **Mini-Reviews in Medicinal Chemistry**, v. 13, n. 4, p. 617–626, 2013.

LIU, Ling Li; WU, Chuan Hai; QIAN, Pei Yuan. Marine natural products as antifouling molecules–a mini-review (2014–2020). **Biofouling**, 2020.

LIU, Sen; WANG, Haibo; SU, Mingzhi; HWANG, Gwi Ja; HONG, Jongki; JUNG, Jee H. New metabolites from the sponge-derived fungus *Aspergillus sydowii* J05B-7F-4. **Natural Product Research**, v. 31, n. 14, p. 1682–1686, 2017.

LIU, Yayue; XIA, Guoping; LI, Hanxiang; MA, Lin; DING, Bo; LU, Yongjun; HE, Lei; XIA, Xuekui; SHE, Zhigang. Vermistatin derivatives with α -glucosidase inhibitory activity from the mangrove endophytic fungus *Penicillium* sp. HN29-3B1. **Planta Medica**, 2014.

LOQUE, Carolina P.; MEDEIROS, Adriana O.; PELLIZZARI, Franciane M.; OLIVEIRA, Eurico C.; ROSA, Carlos A.; ROSA, Luiz H. Fungal community associated with marine macroalgae from Antarctica. **Polar Biology**, v. 33, n. 5, p. 641–648, 2010.

LOTZ, Christian; SCHMID, Freia F.; ROSSI, Angela; KURDYN, Szymon; KAMPIK, Daniel; DE WEVER, Bart; WALLES, Heike; GROEBER, Florian K. Alternative methods for the replacement of eye irritation testing. **Altex**, 2016.

LOUDA, J. William; LI, Jie; LIU, Lei; WINFREE, M. Nancy; BAKER, Earl W. Chlorophyll-a degradation during cellular senescence and death. *In*: ORGANIC GEOCHEMISTRY 1998.

LUEPKE, N. P.; KEMPER, F. H. The HET-CAM test: An alternative to the draize eye test. **Food and Chemical Toxicology**, v. 24, n. 6–7, p. 495–496, 1986.

LUESCH, H.; MOORE, R. E.; PAUL, V. J.; MOOBERRY, S. L.; CORBETT, T. H. Isolation of dolastatin 10 from the marine cyanobacterium *Symploca species* VP642 and total stereochemistry and biological evaluation of its analogue symplostatin 1. **Journal of Natural Products**, 2001.

MACIEL, Olívia Maria Campanini; TAVARES, Renata Spagolla Napoleão; CALUZ, Daniela Ricardo Engracia; GASPAR, Lorena Rigo; DEBONSI, Hosana Maria. Photoprotective potential of metabolites isolated from algae-associated fungi *Annulohypoxylon stygium*. Journal of Photochemistry and Photobiology B: Biology, 2018.

MARINHA DO BRASIL. **Tratado da Antártica e Protocolo de Madri**. 2ª ediçã ed. Brasilia: Comissão Interministerial para os Recursos do Mar. Secretaria da Comissão, 2016.

MARTINS, Rosiane M.; NEDEL, Fernanda; GUIMARÃES, Victoria B. S.; DA SILVA, Adriana F.; COLEPICOLO, Pio; DE PEREIRA, Claudio M. P.; LUND, Rafael G. Macroalgae Extracts From Antarctica Have Antimicrobial and Anticancer Potential. **Frontiers in Microbiology**, v. 9, p. 1–10, 2018.

MAYSLICH, Constance; GRANGE, Philippe Alain; DUPIN, Nicolas. *Cutibacterium acnes* as an opportunistic pathogen: An update of its virulence-associated factors. **Microorganisms**, 2021.

MCGREGOR, Bethany L.; CONNELLY, C. Roxanne. A review of the control of *Aedes aegypti* (Diptera: Culicidae) in the continental United States. **Journal of Medical Entomology**, 2021.

MEAD, M. Nathaniel. Benefits of sunlight: a bright spot for human health. **Environmental health perspectives**, v. 116, n. 4, p. A160-7, 2008.

MEINITA, Maria Dyah Nur; HARWANTO, Dicky; TIRTAWIJAYA, Gabriel; NEGARA, Bertoka Fajar Surya Perwira; SOHN, Jae-Hak; KIM, Jin-Soo; CHOI, Jae-Suk. Fucosterol of Marine Macroalgae: Bioactivity, Safety and Toxicity on Organism. **Marine Drugs**, v. 19, n. 10, p. 545, 2021.

MELTOFTE, Hans. Biodiversity in the Polar Regions in a warming world. **The Routledge Handbook of the Polar Regions**, n. March, p. 137–148, 2019.

MILLEDGE, John J.; NIELSEN, Birthe V.; BAILEY, David. High-value products from macroalgae: the potential uses of the invasive brown seaweed, *Sargassum muticum*. **Reviews in Environmental Science and Biotechnology**, v. 15, n. 1, p. 67–88, 2016.

MOHAMED, Gamal A.; IBRAHIM, Sabrin R. M. Untapped potential of marineassociated *Cladosporium* species: An overview on secondary metabolites, biotechnological relevance, and biological activities. **Marine Drugs**, 2021.

MOLINSKI, Tadeusz F.; FAULKNER, D. John. Metabolites of the antarctic sponge *Dendrilla membranosa.* **The Journal of Organic Chemistry**, v. 52, n. 2, p. 296–298, 1987.

MONTUORI, Eleonora; DE PASCALE, Donatella; LAURITANO, Chiara. Recent Discoveries on Marine Organism Immunomodulatory Activities. **Marine Drugs**, v. 20, n. 7, p. 422, 2022.

MORTAZ, Esmaeil; ALIPOOR, Shamila D.; ADCOCK, Ian M.; MUMBY, Sharon; KOENDERMAN, Leo. Update on Neutrophil Function in Severe Inflammation. **Frontiers in Immunology**, v. 9, 2018.

MOSMANN, Tim. Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays. **Journal of Immunological Methods**, 1983.

MOUGET, Jean-Luc; ROSA, Philippe; TREMBLIN, Gérard. Acclimation of Haslea ostrearia to light of different spectral qualities – confirmation of chromatic adaptation' in diatoms. **Journal of Photochemistry and Photobiology B: Biology**, v. 75, n. 1–2, p. 1–11, 2004.

MU, Jing; MA, Huisheng; CHEN, Hong; ZHANG, Xiaoxia; YE, Mengyi. Luteolin Prevents UVB-Induced Skin Photoaging Damage by Modulating SIRT3/ROS/MAPK Signaling: An in vitro and in vivo Studies. **Frontiers in Pharmacology**, 2021.

MUDUR, Sanjay V.; GLOER, James B.; WICKLOW, Donald T. Sporminarins A and B: Antifungal Metabolites from a Fungicolous Isolate of *Sporormiella minimoides*. **The Journal of Antibiotics**, v. 59, n. 8, p. 500–506, 2006.

MUÑOZ, Julie; CULIOLI, Gérald; KÖCK, Matthias. Linear diterpenes from the marine brown alga *Bifurcaria bifurcata*: A chemical perspective. **Phytochemistry Reviews**, 2013.

MURBACH TELES ANDRADE, Bruna Fernanda; NUNES BARBOSA, Lidiane; DA SILVA PROBST, Isabella; FERNANDES JÚNIOR, Ary. Antimicrobial activity of
essential oils. Journal of Essential Oil Research, 2014.

NAMAN, C. Benjamin et al. Integrating Molecular Networking and Biological Assays To Target the Isolation of a Cytotoxic Cyclic Octapeptide, Samoamide A, from an American Samoan Marine Cyanobacterium. **Journal of Natural Products**, v. 80, n. 3, p. 625–633, 2017.

NĘDZAREK, A., RAKUSA-SUSZCZEWSKI, S. Decomposition of macroalgae and the release of nutrient. **Polar Bioscience**, v. 17, n. January 2004, p. 26–35, 2004.

NOMURA, Masatoshi; KAMOGAWA, Hiroyuki; SUSANTO, Eko; KAWAGOE, Chikara; YASUI, Hajime; SAGA, Naotsune; HOSOKAWA, Masashi; MIYASHITA, Kazuo. Seasonal variations of total lipids, fatty acid composition, and fucoxanthin contents of *Sargassum horneri* (Turner) and *Cystoseira hakodatensis* (Yendo) from the northern seashore of Japan. Journal of Applied Phycology, 2013.

NOOR, Ahmad Omar; ALMASRI, Diena Mohammedallam; BAGALAGEL, Alaa Abdullah; ABDALLAH, Hossam Mohamed; MOHAMED, Shaimaa Gamal Abdallah; MOHAMED, Gamal Abdallah; IBRAHIM, Sabrin Ragab Mohamed. Naturally occurring isocoumarins derivatives from endophytic fungi: Sources, isolation, structural characterization, biosynthesis, and biological activities. **Molecules**, 2020.

NÚÑEZ-PONS, Laura; AVILA, Conxita; ROMANO, Giovanna; VERDE, Cinzia; GIORDANO, Daniela. UV-protective compounds in marine organisms from the southern ocean. **Marine Drugs**, v. 16, n. 9, 2018.

NÚÑEZ-PONS, Laura; SHILLING, Andrew; VERDE, Cinzia; BAKER, Bill J.; GIORDANO, Daniela. Marine terpenoids from polar latitudes and their potential applications in biotechnology. **Marine Drugs**, v. 18, n. 8, 2020.

OECD/OCDE. Guidelines for Testing of Chemicals Test no. 432: In Vitro 3T3 NRU Phototoxicity Test. , 2019. a. Disponível em: https://www.oecdilibrary.org/environment/test-no-432-in-vitro-3t3-nru-phototoxicitytest_9789264071162-en.

OECD/OCDE. **Test No. 432: In Vitro 3T3 NRU Phototoxicity Test**. : OECD, 2019. b. Disponível em: https://www.oecd-ilibrary.org/environment/test-no-432-in-vitro-3t3-nru-phototoxicity-test_9789264071162-en.

OGAKI, Mayara B. et al. Cultivable fungi present in deep-sea sediments of Antarctica: taxonomy, diversity, and bioprospecting of bioactive compounds. **Extremophiles**, v. 24, n. 2, p. 227–238, 2020. a.

OGAKI, Mayara B. et al. Diversity and bioprospecting of cultivable fungal assemblages in sediments of lakes in the Antarctic Peninsula. **Fungal Biology**, v. 124, n. 6, p. 601– 611, 2020. b.

OKADA, Naomasa; SHIRATA, Katsutoshi; NIWANO, Mitsuru; KOSHINO, Hiroyuki; URAMOTO, Masakazu. Immunosuppressive activity of a monoterpene from *Eucommia ulmoides*. **Phytochemistry**, 1994.

OKUNADE, Adewole L.; WIEMER, David F. (-)-Loliolide, an ant-repellent compound from *Xanthoxyllum setulosum*. Journal of Natural Products, 1985.

OLIVEIRA, Mariana C.; PELLIZZARI, Franciane; MEDEIROS, Amanda S.; YOKOYA, Nair S. Diversity of Antarctic Seaweeds. *In*: GÓMEZ, I.; HUOVINEN, P. (org.). **Antarctic Seaweeds**. Cham: Springer International Publishing, 2020. a. p. 23–42. DOLIVEIRA, Mariana C.; PELLIZZARI, Franciane; MEDEIROS, Amanda S.; YOKOYA, Nair S. Diversity of Antarctic Seaweeds. *In*: **Antarctic Seaweeds**. Cham: Springer International Publishing, 2020. b. p. 23–42.

ONOUE, Satomi et al. Establishment and intra-/inter-laboratory validation of a standard protocol of reactive oxygen species assay for chemical photosafety evaluation. **Journal of Applied Toxicology**, p. n/a-n/a, 2012.

PALLELA, Ramjee; NA-YOUNG, Yoon; KIM, Se-Kwon. Anti-photoaging and photoprotective compounds derived from marine organisms. **Marine drugs**, v. 8, n. 4, p. 1189–202, 2010.

PALOMINO, Juan-Carlos; MARTIN, Anandi; CAMACHO, Mirtha; GUERRA, Humberto; SWINGS, Jean; PORTAELS, Françoise. Resazurin Microtiter Assay Plate: Simple and Inexpensive Method for Detection of Drug Resistance in *Mycobacterium tuberculosis*. **Antimicrobial Agents and Chemotherapy**, v. 46, n. 8, p. 2720–2722, 2002.

PANG, Ka-Lai et al. 'Marine fungi' and 'marine-derived fungi' in natural product chemistry research: Toward a new consensual definition. **Fungal Biology Reviews**,

v. 30, n. 4, p. 163–175, 2016.

PEREIRA, Florbela; AIRES-DE-SOUSA, Joao. Computational Methodologies in the Exploration of Marine Natural Product Leads. **Marine Drugs**, v. 16, n. 7, p. 236, 2018.

PEREIRA, Luiz Miguel; VIGATO-FERREIRA, Isabel Cristina; DE LUCA, Gabriela; BRONZON DA COSTA, Cássia Mariana; YATSUDA, Ana Patrícia. Evaluation of methylene blue, pyrimethamine and its combination on an in vitro *Neospora caninum* model. **Parasitology**, v. 144, n. 6, p. 827–833, 2017.

PERSSON, Frank; SVENSSON, Robin; NYLUND, Göran M.; FREDRIKSSON, N. Johan; PAVIA, Henrik; HERMANSSON, Malte. Ecological role of a seaweed secondary metabolite for a colonizing bacterial community. **Biofouling**, v. 27, n. 6, p. 579–588, 2011.

PETTIT, George R. et al. Isolation of dolastatins 10–15 from the marine mollusc. **Tetrahedron**, v. 49, n. 41, p. 9151–9170, 1993.

PHILIPPUS, Ana Cláudia; ZATELLI, Gabriele A.; WANKE, Tauana; GABRIELA DE BARROS, Maria A.; KAMI, Satomy A.; LHULLIER, Cintia; ARMSTRONG, Lorene; SANDJO, Louis P.; FALKENBERG, Miriam. Molecular networking prospection and characterization of terpenoids and C15-acetogenins in Brazilian seaweed extracts. **RSC Advances**, v. 8, n. 52, p. 29654–29661, 2018.

PILON, Alan Cesar; DEL GRANDE, Marcelo; SILVÉRIO, Maíra R. S.; SILVA, Ricardo R.; ALBERNAZ, Lorena C.; VIEIRA, Paulo Cézar; LOPES, João Luis Callegari; ESPINDOLA, Laila S.; LOPES, Norberto Peporine. Combination of GC-MS Molecular Networking and Larvicidal Effect against *Aedes aegypti* for the Discovery of Bioactive Substances in Commercial Essential Oils. **Molecules**, v. 27, n. 5, p. 1588, 2022.

PRASAD, Malini A.; ZOLNIK, Christine P.; MOLINA, Jeanmaire. Leveraging phytochemicals: The plant phylogeny predicts sources of novel antibacterial compounds. **Future Science OA**, 2019.

PRAUD, Annie; VALLS, Robert; PIOVETTI, Louis; BANAIGS, Bernard; BENAIM, Jean Yves. Meroditerpenes from the brown alga *Cystoseira crinita* off the french mediterranean coast. **Phytochemistry**, 1995.

QADER, M. Mallique; HAMED, Ahmed A.; SOLDATOU, Sylvia; ABDELRAOF, Mohamed; ELAWADY, Mohamed E.; HASSANE, Ahmed S. I.; BELBAHRI, Lassaad; EBEL, Rainer; RATEB, Mostafa E. Antimicrobial and antibiofilm activities of the fungal metabolites isolated from the marine endophytes *Epicoccum nigrum* m13 and *Alternaria alternata* 13a. **Marine Drugs**, 2021.

RAFFA, Robert B.; PERGOLIZZI, Joseph V.; TAYLOR, Robert; KITZEN, Jan M. Sunscreen bans: Coral reefs and skin cancer. **Journal of Clinical Pharmacy and Therapeutics**, v. 44, n. 1, p. 134–139, 2019.

RANGEL, Karen C.; DEBONSI, Hosana M.; CLEMENTINO, Leandro C.; GRAMINHA, Márcia A. S.; VILELA, Leonardo Z.; COLEPICOLO, Pio; GASPAR, Lorena R. Antileishmanial activity of the Antarctic red algae *Iridaea cordata* (Gigartinaceae; Rhodophyta). **Journal of Applied Phycology**, v. 31, n. 2, p. 825–834, 2019.

RANGEL, Karen Cristina; VILLELA, Leonardo Zambotti; PEREIRA, Karina de Castro; COLEPICOLO, Pio; DEBONSI, Hosana Maria; GASPAR, Lorena Rigo. Assessment of the photoprotective potential and toxicity of Antarctic red macroalgae extracts from *Curdiea racovitzae* and *Iridaea cordata* for cosmetic use. **Algal Research**, v. 50, p. 101984, 2020.

REN, Yi et al. Two new polyketides from the fungus *Penicillium oxalicum* MHZ153. **Natural Product Research**, 2019.

REPETTO, Guillermo; DEL PESO, Ana; ZURITA, Jorge L. Neutral red uptake assay for the estimation of cell viability/ cytotoxicity. **Nature Protocols**, 2008.

ROBOTTI, Elisa; MARENGO, Emilio. Chemometric Multivariate Tools for Candidate Biomarker Identification: LDA, PLS-DA, SIMCA, Ranking-PCA. *In*: **Methods in Molecular Biology**. p. 237–267.

ROCA, I. et al. The global threat of antimicrobial resistance: Science for intervention. **New Microbes and New Infections**, 2015.

ROCHA, Otávio P.; DE FELÍCIO, Rafael; RODRIGUES, Ana Helena B.; AMBRÓSIO, Daniela L.; CICARELLI, Regina Maria B.; DE ALBUQUERQUE, Sérgio; YOUNG, Maria Claudia M.; YOKOYA, Nair S.; DEBONSI, Hosana M. Chemical profile and biological potential of non-polar fractions from *Centroceras clavulatum* (C. Agardh) montagne (ceramiales, rhodophyta). Molecules, v. 16, n. 8, p. 7105–7114, 2011.

ROSA, Luiz H. et al. **Fungi of Antarctica**. Cham: Springer International Publishing, 2019. a..

ROSA, Luiz Henrique; ZANI, Carlos Leomar; CANTRELL, Charles Lowell; DUKE, Stephen Oscar; VAN DIJCK, Patrick; DESIDERI, Alessandro; ROSA, Carlos Augusto. Fungi in Antarctica: Diversity, Ecology, Effects of Climate Change, and Bioprospection for Bioactive Compounds. *In*: **Fungi of Antarctica**. Cham: Springer International Publishing, 2019. b. p. 1–17.

ROSALES, Carlos. Neutrophils at the crossroads of innate and adaptive immunity. **Journal of Leukocyte Biology**, v. 108, n. 1, p. 377–396, 2020.

SA, Larissa de Lima; RODRIGUES, Rodrigo Vieira; ALVES, Vani M.; GASPAR, Lorena Rigo. Strategies for the evaluation of the eye irritation potential of different types of surfactants and silicones used in cosmetic products. **Toxicology in Vitro**, v. 81, p. 105351, 2022.

SAIDE, Assunta; LAURITANO, Chiara; IANORA, Adrianna. Pheophorbide a: State of the Art. **Marine Drugs**, v. 18, n. 5, p. 257, 2020.

SAKAMOTO, Shuichi et al. Decalpenic acid, a novel small molecule from *Penicillium verruculosum* CR37010, induces early osteoblastic markers in pluripotent mesenchymal cells. **The Journal of Antibiotics**, v. 63, n. 12, p. 703–708, 2010.

SALVATORE, Maria Michela; ANDOLFI, Anna; NICOLETTI, Rosario. The genus *Cladosporium*: A rich source of diverse and bioactive natural compounds. **Molecules**, v. 26, 2021.

SALVATORE, Maria Michela; DELLAGRECA, Marina; ANDOLFI, Anna; NICOLETTI, Rosario. New Insights into Chemical and Biological Properties of Funicone-like Compounds. **Toxins**, v. 14, n. 7, p. 466, 2022.

SARASAN, Manomi; PUTHUMANA, Jayesh; JOB, Neema; HAN, Jeonghoon; LEE, Jae Seong; PHILIP, Rosamma. Marine algicolous endophytic fungi-a promising drug resource of the era. **Journal of Microbiology and Biotechnology**, v. 27, n. 6, p. 1039–1052, 2017.

SEO, Hun-Su; KIM, Kyoung Hee; KIM, Dae-Yong; PARK, Bong-Kyun; SHIN, Nam-Shik; KIM, Jae-Hoon; YOUN, Heejeong. GC/MS analysis of high-performance liquid chromatography fractions from *Sophora flavescens* and *Torilis japonica* extracts and their in vitro anti-neosporal effects on *Neospora caninum*. Journal of Veterinary Science, v. 14, n. 3, p. 241, 2013.

SHANNON, Emer; ABU-GHANNAM, Nissreen. Optimisation of fucoxanthin extraction from Irish seaweeds by response surface methodology. **Journal of Applied Phycology**, 2017.

SHANNON, Emer; ABU-GHANNAM, Nissreen. Seaweeds as nutraceuticals for health and nutrition. **Phycologia**, v. 58, n. 5, p. 563–577, 2019.

SHARMA, Divakar; MISBA, Lama; KHAN, Asad U. Antibiotics versus biofilm: An emerging battleground in microbial communities. **Antimicrobial Resistance and Infection Control**, 2019.

SHILLING, Andrew J.; VON SALM, Jacqueline L.; SANCHEZ, Anthony R.; KEE, Younghoon; AMSLER, Charles D.; MCCLINTOCK, James B.; BAKER, Bill J. Anverenes B-E, new polyhalogenated monoterpenes from the antarctic red alga *Plocamium cartilagineum*. **Marine Drugs**, v. 17, n. 4, 2019. a.

SHILLING, Andrew J.; VON SALM, Jacqueline L.; SANCHEZ, Anthony R.; KEE, Younghoon; AMSLER, Charles D.; MCCLINTOCK, James B.; BAKER, Bill J. Anverenes B–E, New Polyhalogenated Monoterpenes from the Antarctic Red Alga *Plocamium cartilagineum*. **Marine Drugs**, v. 17, n. 4, p. 230, 2019. b.

SIEGERT, Martin et al. The Antarctic Peninsula under a 1.5°C global warming scenario. Frontiers in Environmental Science, 2019.

SILVA, Aurora; SILVA, Sofia A.; CARPENA, M.; GARCIA-OLIVEIRA, P.; GULLÓN, P.; BARROSO, M. Fátima; PRIETO, M. A.; SIMAL-GANDARA, J. Macroalgae as a Source of Valuable Antimicrobial Compounds: Extraction and Applications. **Antibiotics**, v. 9, n. 10, p. 642, 2020. a.

SILVA, Raquel L.; DEMARQUE, Daniel P.; DUSI, Renata G.; SOUSA, João Paulo B.; ALBERNAZ, Lorena C.; ESPINDOLA, Laila S. Residual Larvicidal Activity of Quinones against *Aedes aegypti*. **Molecules**, 2020. b.

SINGH, Rachana; PARIHAR, Parul; SINGH, Madhulika; BAJGUZ, Andrzej; KUMAR, Jitendra; SINGH, Samiksha; SINGH, Vijay P.; PRASAD, Sheo M. Uncovering potential applications of cyanobacteria and algal metabolites in biology, agriculture and medicine: Current status and future prospects. **Frontiers in Microbiology**, v. 8, n. APR, p. 1–37, 2017.

SLOMINSKI, Andrzej; PAWELEK, John. Animals under the sun: effects of ultraviolet radiation on mammalian skin. **Clinics in Dermatology**, v. 16, n. 4, p. 503–515, 1998.

SNEATH, P. H. A. Thirty Years of Numerical Taxonomy. **Systematic Biology**, v. 44, n. 3, p. 281–298, 1995.

SOLDATOU, Sylvia; BAKER, Bill J. Cold-water marine natural products, 2006 to 2016. **Natural Product Reports**, v. 34, n. 6, p. 585–626, 2017.

STERMITZ, Frank R.; TAWARA-MATSUDA, Jeanne; LORENZ, Peter; MUELLER, Paul; ZENEWICZ, Lauren; LEWIS, Kim. 5'-Methoxyhydnocarpin-D and Pheophorbide A: Berberis Species Components that Potentiate Berberine Growth Inhibition of Resistant *Staphylococcus aureus*. **Journal of Natural Products**, v. 63, n. 8, p. 1146–1149, 2000.

STIERLE, Andrea A.; STIERLE, Donald B.; GIRTSMAN, Teri. Caspase-1 inhibitors from an extremophilic fungus that target specific leukemia cell lines. **Journal of Natural Products**, 2012.

STIERLE, Donald B.; STIERLE, Andrea A.; PATACINI, Brianna. The Berkeleyacetals, Three Meroterpenes from a Deep Water Acid Mine Waste *Penicillium*. **Journal of Natural Products**, v. 70, n. 11, p. 1820–1823, 2007..

SUDATTI, Daniela Bueno; FUJII, Mutue Toyota; RODRIGUES, Silvana Vianna; TURRA, Alexander; PEREIRA, Renato Crespo. Effects of abiotic factors on growth and chemical defenses in cultivated clones of *Laurencia dendroidea* J. Agardh (Ceramiales, Rhodophyta). **Marine Biology**, 2011.

SUDATTI, Daniela Bueno; OLIVEIRA, Aline Santos; DA GAMA, Bernardo Antonio Perez; FUJII, Mutue Toyota; RODRIGUES, Silvana Vianna; PEREIRA, Renato Crespo. Variability in Seaweed Chemical Defense and Growth Under Common Garden Conditions. **Frontiers in Marine Science**, v. 8, 2021. SUN, Chunxiao et al. Antibacterial Cyclic Tripeptides from Antarctica-Sponge-Derived Fungus *Aspergillus insulicola* HDN151418. **Marine Drugs**, v. 18, n. 11, p. 532, 2020.

SUN, Jing et al. Nitric Oxide Inhibitory Meroterpenoids from the Fungus *Penicillium purpurogenum* MHZ 111. **Journal of Natural Products**, v. 79, n. 5, p. 1415–1422, 2016.

SURYANARAYANAN, T. S. Fungal endosymbionts of seaweeds. **Progress in** molecular and subcellular biology, 2012.

SUZUKI, Minoru; YAMADA, Hiroko; KURATA, Kazuya. Dictyterpenoids A and B, two novel diterpenoids with feeding-deterrent activity from the brown alga *Dilophus okamurae*. **Journal of Natural Products**, 2002.

TAMURA, Koichiro; NEI, Masatoshi; KUMAR, Sudhir. Prospects for inferring very large phylogenies by using the neighbor-joining method. **Proceedings of the National Academy of Sciences**, v. 101, n. 30, p. 11030–11035, 2004.

TAMURA, Koichiro; STECHER, Glen; KUMAR, Sudhir. MEGA11: Molecular Evolutionary Genetics Analysis Version 11. **Molecular Biology and Evolution**, v. 38, n. 7, p. 3022–3027, 2021.

TAN, Ren Xiang; JENSEN, Paul R.; WILLIAMS, Philip G.; FENICAL, William. Isolation and structure assignments of rostratins A-D, cytotoxic disulfides produced by the marine-derived fungus *Exserohilum rostratum*. **Journal of Natural Products**, 2004.

TAVARES, Renata Spagolla Napoleão; KAWAKAMI, Camila Martins; PEREIRA, Karina de Castro; DO AMARAL, Gabriela Timotheo; BENEVENUTO, Carolina Gomes; MARIA-ENGLER, Silvya Stuchi; COLEPICOLO, Pio; DEBONSI, Hosana Maria; GASPAR, Lorena Rigo. Fucoxanthin for topical administration, a phototoxic vs. Photoprotective potential in a tiered strategy assessed by in vitro methods. **Antioxidants**, v. 9, n. 4, 2020. a.

TAVARES, Renata Spagolla Napoleão; MARIA-ENGLER, Silvya Stuchi; COLEPICOLO, Pio; DEBONSI, Hosana Maria; SCHÄFER-KORTING, Monika; MARX, Uwe; GASPAR, Lorena Rigo; ZOSCHKE, Christian. Skin irritation testing beyond tissue viability: Fucoxanthin effects on inflammation, homeostasis, and metabolism. **Pharmaceutics**, v. 12, n. 2, p. 1–12, 2020. c.

TEIXEIRA, Thaiz Rodrigues; DOS SANTOS, Gustavo Souza; ARMSTRONG, Lorene; COLEPICOLO, Pio; DEBONSI, Hosana Maria. Antitumor potential of seaweed derived-endophytic fungi. **Antibiotics**, v. 8, n. 4, 2019. a.

TEIXEIRA, Thaiz Rodrigues; RANGEL, Karen Cristina; TAVARES, Renata Spagolla Napoleão; KAWAKAMI, Camila Martins; DOS SANTOS, Gustavo Souza; MARIA-ENGLER, Silvya Stuchi; COLEPICOLO, Pio; GASPAR, Lorena Rigo; DEBONSI, Hosana Maria. In Vitro Evaluation of the Photoprotective Potential of Quinolinic Alkaloids Isolated from the Antarctic Marine Fungus *Penicillium echinulatum* for Topical Use. **Marine Biotechnology**, p. 357–372, 2021.

TEIXEIRA, Thaiz Rodrigues; SANTOS, Gustavo Souza; TURATTI, Izabel Cristina Casanova; PAZIANI, Mário Henrique; VON ZESKA KRESS, Márcia Regina; COLEPICOLO, Pio; DEBONSI, Hosana Maria. Characterization of the lipid profile of Antarctic brown seaweeds and their endophytic fungi by gas chromatography–mass spectrometry (GC–MS). **Polar Biology**, v. 42, n. 8, p. 1431–1444, 2019. b.

TEZUKA, Yasuhiro; HUANG, Qing; KIKUCHI, Tohru; NISHI, Arasuke; TUBAKI, Keisuke. Studies on the Metabolites of Mycoparasitic Fungi. I. Metabolites of Cladobotryum varium. **Chemical and Pharmaceutical Bulletin**, v. 42, n. 12, p. 2612–2617, 1994.

THIESEN, Liliani Carolini; BACCARIN, Thaisa; FISCHER-MULLER, Ana Flávia; MEYRE-SILVA, Christiane; COUTO, Angelica Garcia; BRESOLIN, Tania Mari Bellé; SANTIN, José Roberto. Photochemoprotective effects against UVA and UVB irradiation and photosafety assessment of Litchi chinensis leaves extract. **Journal of Photochemistry and Photobiology B: Biology**, 2017.

TOLLER-KAWAHISA, Juliana Escher; CANICOBA, Nathália Cristina; VENANCIO, Vinicius Paula; KAWAHISA, Rogério; ANTUNES, Lusânia Maria Greggi; CUNHA, Thiago Mattar; MARZOCCHI-MACHADO, Cleni Mara. Systemic lupus erythematosus onset in lupus-prone B6.MRL/lpr mice Is influenced by weight gain and Is preceded by an increase in neutrophil oxidative burst activity. **Free Radical Biology and Medicine**, 2015.

TONG, Steven Y. C.; DAVIS, Joshua S.; EICHENBERGER, Emily; HOLLAND, Thomas L.; FOWLER, Vance G. Staphylococcus aureus Infections: Epidemiology,

Pathophysiology, Clinical Manifestations, and Management. Clinical Microbiology **Reviews**, v. 28, n. 3, p. 603–661, 2015.

TORRES-GUERRERO, Edoardo; QUINTANILLA-CEDILLO, Marco Romano; RUIZ-ESMENJAUD, Julieta; ARENAS, Roberto. Leishmaniasis: a review. **F1000Research**, v. 6, p. 750, 2017.

URBAN, Laurent; CHARLES, Florence; DE MIRANDA, Maria Raquel Alcântara; AARROUF, Jawad. Understanding the physiological effects of UV-C light and exploiting its agronomic potential before and after harvest. **Plant Physiology and Biochemistry**, 2016.

VALLE, Denise; NACIF PIMENTA, Denise; AGUIAR, Raquel. Zika, dengue e chikungunya: desafios e questões. **Epidemiologia e Serviços de Saúde**, v. 25, n. 2, p. 1–2, 2016.

VELÁSQUEZ, Angela M. A. et al. Antiprotozoal activity of the cyclopalladated complexes against *Leishmania amazonensis* and *Trypanosoma cruzi*. Journal of the Brazilian Chemical Society, 2016.

VOM DORP, Katharina; HÖLZL, Georg; PLOHMANN, Christian; EISENHUT, Marion; ABRAHAM, Marion; WEBER, Andreas P. M.; HANSON, Andrew D.; DÖRMANN, Peter. Remobilization of Phytol from Chlorophyll Degradation Is Essential for Tocopherol Synthesis and Growth of Arabidopsis. **The Plant Cell**, p. tpc.15.00395, 2015.

VON SALM, Jacqueline L.; SCHOENROCK, Kathryn M.; MCCLINTOCK, James B.; AMSLER, Charles D.; BAKER, Bill J. The Status of Marine Chemical Ecology in Antarctica. *In*: **Chemical Ecology**. [s.l.] : CRC Press, 2018. p. 27–69.

WAHL, Martin; GOECKE, Franz; LABES, Antje; DOBRETSOV, Sergey; WEINBERGER, Florian. The Second Skin: Ecological Role of Epibiotic Biofilms on Marine Organisms. **Frontiers in Microbiology**, v. 3, 2012.

WANG, Hui; WANG, Yi; WANG, Wei; FU, Peng; LIU, Peipei; ZHU, Weiming. Antiinfluenza virus polyketides from the acid-tolerant fungus *Penicillium purpurogenum* JS03-21. Journal of Natural Products, 2011. WANG, Jie; NONG, Xu Hua; ZHANG, Xiao Yong; XU, Xin Ya; AMIN, Muhammad; QI, Shu Hua. Screening of anti-biofilm compounds from marine-derived fungi and the effects of secalonic acid D on *Staphylococcus aureus* biofilm. **Journal of Microbiology and Biotechnology**, 2017.

WANG, Mingxun et al. Sharing and community curation of mass spectrometry data with Global Natural Products Social Molecular Networking. **Nature Biotechnology**, v. 34, n. 8, p. 828–837, 2016.

WAUCHOPE, Hannah S.; SHAW, Justine D.; TERAUDS, Aleks. A snapshot of biodiversity protection in Antarctica. **Nature Communications**, v. 10, n. 1, p. 1–6, 2019.

WEI, Hong; ITOH, Takuya; KINOSHITA, Masahiro; NAKAI, Yasuhide; KUROTAKI, Mineko; KOBAYASHI, Motomasa. Cytotoxic sesterterpenes, 6-epi-ophiobolin G and 6-epi-ophiobolin N, from marine derived fungus *Emericella variecolor* GF10. **Tetrahedron**, v. 60, n. 28, p. 6015–6019, 2004.

WENZEL, Lauren; GILBERT, Neil; GOLDSWORTHY, Lyn; TESAR, Clive; MCCONNELL, Martha; OKTER, Melis. Polar opposites? Marine conservation tools and experiences in the changing Arctic and Antarctic. **Aquatic Conservation: Marine and Freshwater Ecosystems**, v. 26, p. 61–84, 2016.

WERTHEIM, Heiman F. L.; MELLES, Damian C.; VOS, Margreet C.; VAN LEEUWEN, Willem; VAN BELKUM, Alex; VERBRUGH, Henri A.; NOUWEN, Jan L. The role of nasal carriage in *Staphylococcus aureus* infections. **The Lancet Infectious Diseases**, v. 5, n. 12, p. 751–762, 2005.

WHITCHURCH, Cynthia B.; TOLKER-NIELSEN, Tim; RAGAS, Paula C.; MATTICK, John S. Extracellular DNA required for bacterial biofilm formation. **Science**, 2002.

WHITE, T. J.; BRUNS, T.; LEE, S.; TAYLOR, J. AMPLIFICATION AND DIRECT SEQUENCING OF FUNGAL RIBOSOMAL RNA GENES FOR PHYLOGENETICS. *In*: **PCR Protocols**, 1990.

WHO. Antibiotic resistance. **World Health Organization**, 2022. Disponível em: https://www.who.int/news-room/fact-sheets/detail/antibiotic-resistance.

WIENCKE, C. et al. Life strategy, ecophysiology and ecology of seaweeds in polar waters. **Reviews in Environmental Science and Biotechnology**, v. 6, n. 1–3, p. 95–126, 2007.

WOLD, Svante; ESBENSEN, Kim; GELADI, Paul. Principal component analysis. **Chemometrics and Intelligent Laboratory Systems**, v. 2, n. 1–3, p. 37–52, 1987.

WOLFENDER, Jean Luc; NUZILLARD, Jean Marc; VAN DER HOOFT, Justin J. J.; RENAULT, Jean Hugues; BERTRAND, Samuel. Accelerating Metabolite Identification in Natural Product Research: Toward an Ideal Combination of Liquid Chromatography-High-Resolution Tandem Mass Spectrometry and NMR Profiling, in Silico Databases, and Chemometrics. **Analytical Chemistry**, 2019.

WU, Chang Jing; YI, Le; CUI, Cheng Bin; LI, Chang Wei; WANG, Nan; HAN, Xiao. Activation of the silent secondary metabolite production by introducing neomycin-resistance in a marine-derived *Penicillium purpurogenum* G59. **Marine Drugs**, v. 13, n. 4, p. 2465–2487, 2015.

WU, Yu Zhuo; XIA, Gui Yang; XIA, Huan; WANG, Ling Yan; WANG, Ya Nan; LI, Li; SHANG, Hong Cai; LIN, Sheng. Seco and Nor-seco Isodhilarane-Type Meroterpenoids from *Penicillium purpurogenum* and the Configuration Revisions of Related Compounds. **Journal of Natural Products**, 2022.

XIA, Menglu; LIU, Chunping; GAO, Lei; LU, Yanbin. One-Step Preparative Separation of Phytosterols from Edible Brown Seaweed *Sargassum horneri* by High-Speed Countercurrent Chromatography. **Marine Drugs**, v. 17, n. 12, p. 691, 2019.

XU, Jianzhou; YI, Mengqi; DING, Lijian; HE, Shan. A review of anti-inflammatory compounds from marine fungi, 2000-2018. **Marine Drugs**, 2019.

YANG, Jane Y. et al. Molecular networking as a dereplication strategy. **Journal of Natural Products**, 2013.

YANG, Xiliang; LIU, Jinping; MEI, Jiahui; JIANG, Rui; TU, Shizheng; DENG, Huafeng; LIU, Jing; YANG, Sumei; LI, Juan. Origins, Structures, and Bioactivities of Secondary Metabolites from Marine-derived *Penicillium* Fungi. **Mini-Reviews in Medicinal Chemistry**, 2021.

YANG, Xiudong; KANG, Min-Cheol; LEE, Ki-Wan; KANG, Sung-Myung; LEE, Won-Woo; JEON, You-Jin. Antioxidant activity and cell protective effect of loliolide isolated from *Sargassum ringgoldianum* subsp. coreanum. **Algae**, v. 26, n. 2, p. 201–208, 2011.

YAO, Guangmin; SEBISUBI, Fred Musoke; VOO, Lok Yung Christopher; HO, Coy Choke; TAN, Ghee Teng; CHANG, Leng Chee. Citrinin derivatives from the soil filamentous fungus *Penicillium* sp. H9318. **Journal of the Brazilian Chemical Society**, v. 22, n. 6, p. 1125–1129, 2011.

YOUNG, Ryan; VON SALM, Jacqueline; AMSLER, Margaret; LOPEZ-BAUTISTA, Juan; AMSLER, Charles; MCCLINTOCK, James; BAKER, Bill. Site-Specific Variability in the Chemical Diversity of the Antarctic Red Alga *Plocamium cartilagineum*. **Marine Drugs**, v. 11, n. 6, p. 2126–2139, 2013.

YUYAMA, Kamila Tomoko; ROHDE, Manfred; MOLINARI, Gabriella; STADLER, Marc; ABRAHAM, Wolf-Rainer. Unsaturated Fatty Acids Control Biofilm Formation of *Staphylococcus aureus* and Other Gram-Positive Bacteria. **Antibiotics**, v. 9, n. 11, p. 788, 2020.

ZHANG, Dahai; YANG, Xiudong; KANG, Jung Sook; CHOI, Hong Dae; SON, Byeng Wha. Circumdatin I, a new ultraviolet-A protecting benzodiazepine alkaloid from a marine isolate of the fungus *Exophiala*. **Journal of Antibiotics**, 2008.

ZHANG, Yonggang; LIU, Shuchun; CHE, Yongsheng; LIU, Xingzhong. Epicoccins A– D, Epipolythiodioxopiperazines from a Cordyceps-Colonizing Isolate of *Epicoccum nigrum*. **Journal of Natural Products**, v. 70, n. 9, p. 1522–1525, 2007.

ZHAO, Lu; KIM, Jin-Cheol; PAIK, Man-Jeong; LEE, Wonjae; HUR, Jae-Seoun. A Multifunctional and Possible Skin UV Protectant, (3R)-5-Hydroxymellein, Produced by an Endolichenic Fungus Isolated from *Parmotrema austrosinense*. **Molecules**, v. 22, n. 1, p. 26, 2016.

ZHAO, Qi; ZHANG, Jia-Le; LI, Fei. Application of Metabolomics in the Study of Natural Products. **Natural Products and Bioprospecting**, v. 8, n. 4, p. 321–334, 2018.

ZHENG, Cai Juan et al. Bioactive hydroanthraquinones and anthraquinone dimers from a soft coral-derived *Alternaria* sp. fungus. **Journal of Natural Products**, 2012.

