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**Sistemas de liberação para aplicação tópica de peptídeos sob
estímulo elétrico para tratamento de feridas**

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Sistemas de liberação para aplicação tópica de peptídeos sob estímulo elétrico para tratamento de feridas

Tese de Doutorado apresentada ao Programa de Pós-Graduação em Ciências Farmacêuticas da Faculdade de Ciências Farmacêuticas de Ribeirão Preto/USP para obtenção do Título de Doutor em Ciências

Área de Concentração: Medicamentos e Cosméticos

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AUTORIZO A REPRODUÇÃO E DIVULGAÇÃO TOTAL OU PARCIAL DESTE TRABALHO, POR QUALQUER MEIO CONVENCIONAL OU ELETRÔNICO, PARA FINS DE ESTUDO E PESQUISA, DESDE QUE CITADA A FONTE.

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1.Cicatrização. 2.Iontoforese. 3.Fibroína da seda.

RESUMO

LEMOS, C. N. **Sistemas de liberação para aplicação tópica de peptídeos sob estímulo elétrico para tratamento de feridas**. 2019. 144f. Tese (Doutorado em Ciências). Faculdade de Ciências Farmacêuticas de Ribeirão Preto – Universidade de São Paulo, Ribeirão Preto, 2019.

O tratamento de feridas crônicas é um grande desafio para a saúde pública na atualidade. Devido à complexidade do processo de cicatrização, envolvendo respostas inflamatórias, de proliferação celular e reconstrução do tecido epitelial, o mercado carece de medicamentos que aliem proteção e modulação desse processo para que as feridas cicatrizem adequadamente. Dentre as diversas estratégias estudadas para modular a cicatrização estão a administração prolongada de peptídeos com ação anti-inflamatória e de proliferação celular, a estimulação elétrica e a aplicação de curativos que protejam a ferida contra infecções e perda de fluidos. O objetivo deste trabalho foi associar, de forma racional, essas estratégias em um único sistema de liberação e verificar sua influência na modulação do processo de cicatrização. Para tanto, filmes de fibroína da seda (FS) carreadores de neurotensina (NT) foram desenvolvidos de forma a sustentar a liberação da NT, proteger a ferida e ainda permitir a aplicação de uma corrente elétrica constante e de baixa intensidade, conhecida como iontoforese. Filmes de FS contendo NT (FS-NT) e glicerina como plastificante foram obtidos por evaporação de solvente. Filmes bilaminados contendo um eletrodo de prata também foram preparados para a aplicação da iontoforese. Os filmes apresentaram-se homogêneos, transparentes, permeáveis ao vapor de água, com baixa capacidade de intumescimento e elevada resistência mecânica. Análises de espectroscopia de infravermelho por transformada de Fourier e calorimetria diferencial sugeriram uma maior organização das fibras de FS do filme em conformação de folhas- β /Seda II. A ocorrência de interações não-covalentes entre NT e FS, capazes de estabilizar a estrutura do filme, foi sugerida por essas análises e confirmada por calorimetria de titulação isotérmica. Assim, o filme de FS-NT sustentou a liberação da NT por até 72 h. Quando aplicada, a iontoforese promoveu uma rápida e alta liberação do fármaco do filme, observada por MALDI *imaging*. Análises por microscopia de força atômica mostraram que na presença do fármaco os filmes apresentaram-se mais rugosos, sugerindo maior aderência ao tecido. Estudos *in vitro* em cultura de macrófagos, fibroblastos e queratinócitos mostraram que tanto os filmes quanto a iontoforese não apresentaram efeitos citotóxicos. Ainda em cultura de macrófagos, foi possível observar que os filmes de FS-NT reduziram significativamente a produção de interleucina 6 e fator de necrose tumoral alfa, o que não foi observado para a solução de NT. A aplicação da iontoforese modulou a secreção dessas citocinas em função do sistema de liberação. A iontoforese anódica, mas não a catódica, foi capaz de inibir o crescimento de bactérias Gram positivas e fungo, mas não teve influência no crescimento de bactérias Gram negativas. *In vivo*, em feridas crônicas induzidas em ratos diabéticos, os filmes de FS protegeram as feridas e suportaram o crescimento de células. Os filmes de FS-NT sustentaram a liberação da NT, modulando a expressão de mediadores pró-inflamatórios, a ativação de macrófagos, de miofibroblastos e de queratinócitos. Associados a iontoforese, os filmes reduziram significativamente a expressão de citocinas pró-inflamatórias, modularam a ativação de neutrófilos e macrófagos, estimularam a angiogênese e a proliferação de fibroblastos. No entanto, quando aplicada na etapa final do processo de cicatrização, principalmente associada a NT, a iontoforese parece ter estimulado excessivamente a proliferação de queratinócitos, sugerindo que seja aplicada apenas na primeira semana do tratamento. A aplicação do filme de FS-NT durante todo o período de cicatrização parece ser importante para a manutenção da modulação desencadeada pela aplicação inicial da iontoforese. Desta forma, os filmes de FS-NT elétrico-estimulados desenvolvidos apresentam-se como uma estratégia promissora para o tratamento tópico de feridas crônicas.

Palavras-chave: Cicatrização. Iontoforeses. Fibroína da seda.

ABSTRACT

LEMOS, C. N. **Delivery systems for peptides topical application under electrical stimulation for wounds treatment.** 2019. 144f. Thesis (Doctor of Science). Faculdade de Ciências Farmacêuticas de Ribeirão Preto – Universidade de São Paulo, Ribeirão Preto, 2019.

The treatment of chronic wounds is still a major public health challenge. Due to the complexity of the healing process, which involves inflammatory responses, cell proliferation and epithelial tissue reconstitution, the market still lacks dressings that combine wound protection and modulation of these processes for proper wounds healing. Strategies that have been studied to modulate wound healing includes the sustained administration of peptides that possess anti-inflammatory and cell proliferating properties, electrical stimulation and application of dressings that protect the wound against infection and fluid loss. The aim of this work was to associate, rationally, these strategies into a single delivery system and verify its influence on the modulation of the wound healing process. To achieve this purpose, neurotensin (NT)-loaded silk fibroin (FS) films were developed to support the release of NT, protect the wound and allow the application of a constant and low intensity of electric current known as iontophoresis. FS films containing NT (FS-NT) and glycerine as plasticizer were developed by film casting, while bi-laminated films containing a silver electrode were also prepared for iontophoresis application. The films were homogeneous, transparent, water vapor permeable, and has low swelling capacity and high mechanical resistance. Fourier transform infrared spectroscopy and differential scanning calorimetry analysis suggested a greater organization of the FS film's fibers into β -sheet/Silk II conformation. The occurrence of non-covalent interactions between NT and FS capable of stabilizing the film structure was suggested by these analyzes and confirmed by isothermal titration calorimetry. FS-NT film sustained the release of NT for up to 72 h, iontophoresis application promoted a fast and high release of NT from the film as observed by MALDI imaging. Atomic force microscopy showed that in the presence of NT, films were rougher, suggesting greater adherence to film structure. *In vitro* studies in macrophages, fibroblasts and keratinocytes showed no cytotoxic effects of the films and iontophoresis. However, FS-NT films significantly reduced interleukin-6 and α -TNF production in macrophages as compared to NT solution. Iontophoresis application modulated the secretion of these cytokines as a function of the delivery system. Anodic, but not cathodic iontophoresis inhibited the growth of Gram positive bacteria and fungus, but had no influence on the growth of Gram negative bacteria. *In vivo*, in diabetic rat-induced chronic wounds, FS films protected the wounds and supported cell growth. FS-NT films supported NT release by modulating the expression of proinflammatory mediators, macrophages, myofibroblasts and keratinocytes activation. When FS-NT was associated with iontophoresis, the films significantly reduced proinflammatory cytokine expression, modulated neutrophil and macrophage activation, stimulated angiogenesis and fibroblast proliferation. However, when applied in the final stage of the healing process, especially in association with NT, iontophoresis appears to have excessively stimulated keratinocyte proliferation suggesting that its application may be more beneficial in the early stage rather than the later stages of wound healing. The application of FS-NT film throughout the healing period seems to be important for maintaining the modulation triggered by the initial application of iontophoresis. Thus, the developed electric-stimulated FS-NT film is a promising strategy for the topical treatment of chronic wounds.

Keywords: Healing. Iontophoresis. Silk fibroin.

1. INTRODUÇÃO

Feridas crônicas, de difícil cicatrização, incluindo úlceras de pressão, venosas e aquelas decorrentes da Diabetes *mellitus* (DM) são um problema de saúde mundial (JAMES et al., 2008; YANG et al., 2016). Elas ocorrem devido a falhas na evolução de uma ou várias etapas do processo de cicatrização fisiológico e, quando finalmente fecham resultam em cicatrizes esteticamente desagradáveis e até em invalidez de partes do corpo, com impactos na economia, na saúde e na qualidade de vida do paciente.

As feridas crônicas ocorrem corriqueiramente em indivíduos com idade avançada, obesos, imunossuprimidos e diabéticos. O processo de cicatrização nessas situações apresenta velocidade similar de progressão e de reparo (STEED, 2003), além de ambiente excessivamente inflamado, com predomínio de neutrófilos que preservam essa inflamação (HU; LAN, 2016). Para reverter esse processo e permitir a cicatrização dessas feridas há a necessidade de modular o processo de cicatrização com estratégias inteligentes. Desta forma, várias abordagens terapêuticas têm sido estudadas para um tratamento mais rápido e efetivo das feridas crônicas.

A terapia tópica é a abordagem mais comum e menos invasiva. Ela possui algumas premissas básicas, como estimular o processo de cicatrização, que é complexo e dinâmico, promover a melhoria da qualidade de vida do paciente, limitar infecções, proteger a ferida da perda de fluidos e proteínas e dissipar o estresse mecânico (BOATENG et al., 2008).

Filmes são sistemas de liberação de fármacos considerados bons candidatos para desempenhar essas funções, mas a grande maioria dos curativos tradicionais presentes no mercado foram desenvolvidos apenas para cobrir a ferida, sem exercer influência significativa no processo de cicatrização. Por isso, o tratamento tópico envolve tradicionalmente, antes da aplicação do curativo, a administração, na ferida limpa, de uma formulação semissólida que contém substâncias destinadas a estimular o processo de cicatrização ou com ação antimicrobiana. Sobre essa formulação, que precisa ser aplicada várias vezes ao dia, é colocado um revestimento (curativo), o qual deve ser impermeável a bactérias e removível sem provocar trauma, que tem a função de manter alta a umidade na interface ferida/curativo, de permitir a troca gasosa e de

fornecer isolamento térmico (KAMOUN; KENAWY; CHEN, 2017; ZARRINTAJ et al., 2017).

O processo de cicatrização fisiológico envolve a ação simultânea de mediadores, células sanguíneas e parenquimais e matriz extracelular. Devido a sua complexidade, as substâncias usadas nas formulações destinadas a acelerar o processo de cicatrização devem idealmente ser capazes de modular esse processo e atuar nas suas 4 fases: homeostática, inflamatória, proliferativa e de remodelamento (MOURA et al., 2013a). Dificilmente, no entanto, uma única substância é capaz de atuar em todas essas etapas. Portanto, mais de uma substância ou estratégias associadas precisam ser empregadas para propiciar uma cicatrização adequada da ferida crônica.

Dentre as várias substâncias que têm mostrado resultados interessantes nessa modulação merecem destaque os neuropeptídeos, dentre eles a neurotensina (NT).

A NT é um neuropeptídeo que participa do controle de várias atividades biológicas, tanto do sistema nervoso central quanto do periférico (KLECZKOWSKA; LIPKOWSKI, 2013). Especificamente na pele, a NT estimula o crescimento celular (SCARPA; CARRAWAY; COCHRANE, 2005) e aumenta a permeabilidade vascular (PEREIRA et al., 2014), funções essas importantes na fase de proliferação do processo de cicatrização. Mas a NT também pode atuar na fase inflamatória, inibindo a cascata de sinalização da via fator nuclear kappa β (NF- $\kappa\beta$), envolvida em diversas respostas imunológicas, e reduzindo a expressão da interleucina 6 (IL-6), interleucina 10 (IL-10) e fator de necrose tumoral α (TNF- α) (PEREIRA et al., 2014). A NT tem mostrado assim, ser capaz de diminuir o estado pró-inflamatório e de atrair fibroblastos, além de aumentar a expressão do fator de crescimento epidérmico (EGF) (PEREIRA et al., 2014). O EGF desempenha papel importante na regeneração e proliferação das células da pele, aumentando a proliferação celular e o acúmulo de colágeno tipo 1 α (BALBINO; PEREIRA, LEONARDO MADEIRA; CURI, 2005).

Sendo assim, de acordo com as evidências, a NT influencia no processo de cicatrização de feridas crônicas, visto que pode reduzir a quimiotaxia de células inflamatórias e o próprio estado pró-inflamatório, possibilitando a transição para as fases de proliferação e de remodelamento (PEREIRA et al., 2014) e estimulando a produção de células e mediadores importantes para essas fases (RAMOT; PAUS, 2014).

Para além do uso de substâncias químicas ou biológicas, a estimulação elétrica é outra estratégia que pode contribuir na modulação do processo de cicatrização em feridas crônicas (ISSEROFF; DAHLE, 2012; CASSETTARI et al., 2014; FARBER et al., 2014; KLOTH, 2014; POLAK; FRANEK; TARADAJ, 2014). Já foi demonstrado, por exemplo, que a aplicação de uma corrente elétrica, seja ela pulsátil ou contínua, com densidade de 10-100 $\mu\text{A}/\text{cm}^2$ promove a migração de fibroblastos e queratinócitos (ZHAO, 2009; KLOTH, 2014), podendo, portanto, modular a fase inflamatória e afetar a fase proliferativa do processo de cicatrização.

Tanto a aplicação da NT quanto a da corrente elétrica requerem, no entanto, sistemas de liberação que permitam sua administração tópica adequada. Por ser um peptídeo, a NT tem meia-vida curta, susceptível a ação das peptidases presentes no ambiente inflamado da ferida e propícia a perder rapidamente sua atividade (SWEITZER, SARAH M.; FANN, STEPHEN A.; BORG, THOMAS K.; BAYNES, JOHN W.; YOST, 2006). Sistemas de liberação capazes de sustentar a liberação da NT são, dessa forma, convenientes para protegê-la da degradação e propiciar sua entrega por períodos prolongados, em várias etapas da cicatrização. A aplicação do estímulo elétrico também requer uma plataforma adequada, que permita a passagem e distribuição da corrente elétrica nas etapas em que possa ser necessária.

A hipótese desse trabalho baseia-se, portanto, na premissa de que o desenvolvimento de um filme que permita a aplicação da corrente elétrica e a liberação sustentada da NT possa servir como um curativo promissor para o tratamento de feridas crônicas.

Para tanto, o filme deve ter características que sirvam ao tratamento e a proteção da ferida. O material que o compõe, dessa forma, requer propriedades que permitam que esse propósito seja alcançado. Dentre os materiais capazes de formar filme e com características promissoras para servir de plataforma curativa está a fibroína da seda (FS).

A FS é uma proteína fibrosa presente no casulo do bicho da seda (*Bomby mori*) (ALTMAN, GREGORY H.; DIAZ, FRANK; JAKUBA, CAROLINE; CALABRO, TARA; HORAN, REBECA L.; CHEN, JINGSONG; LU, HELEN; RICHMOND, JOHN; KAPLAN, 2003). É um biopolímero natural que apresenta baixa reação inflamatória, permeabilidade ao oxigênio e vapor d'água, biocompatibilidade e biodegradabilidade, além de resistência mecânica (WENK; MERKLE; MEINEL, 2011). Estudos revelam ainda que a FS interage com algumas células presentes na pele, como fibroblastos

(SERVOLI et al., 2005), queratinócitos (GUPTA et al., 2007) e células endoteliais (FUCHS et al., 2006). Sendo assim, ela vem sendo utilizada como *scaffold* para a reposição de diversos tecidos (MELKE et al., 2016; ZHOU et al., 2017, 2019; BANDYOPADHYAY; MANDAL, 2019). Também serve como base para o preparo de diversas formas farmacêuticas, como filmes, géis, espumas, nano e/ ou micropartículas (WENK; MERKLE; MEINEL, 2011; KUNDU et al., 2013). Desta forma, pretende-se nesse trabalho desenvolver um curativo a base de FS que sirva como sistema de liberação sustentada da NT, plataforma para aplicação de um estímulo elétrico e *scaffold* para a regeneração da pele ferida.

7. CONCLUSÃO

A metodologia para obtenção de dispersão aquosa de FS a partir de casulos de *Bombyx mori* foi padronizada e permitiu a formação de filmes transparentes e homogêneos, com predominância de folhas- β , baixa hidratação, permeabilidade ao vapor de água e propriedades mecânicas adequadas para administração tópica e proteção de feridas. Os filmes de FS foram ainda capazes de incorporar a NT e de sustentar sua liberação, além de servirem como uma plataforma para aplicação de iontoforese. Essa, por sua vez, apresentou efeito bacteriostático contra microrganismos Gram positivos e modulou a expressão de citocinas pró-inflamatórias quando administrada associada ao filme de FS-NT *in vitro*. Aplicados em feridas crônicas induzidas em ratos, confirmou-se a capacidade dos filmes de FS-NT elétrico-estimulados de diminuir o processo inflamatório e permitirem a proliferação modulada de fibroblastos e queratinócitos. A investigação do efeito dos tratamentos em dias-chaves permitiu compreender a influência do filme de FS, da NT e da iontoforese em todas as etapas do processo de cicatrização. Desta forma, pode-se afirmar que a iontoforese deve ser associada aos filmes na primeira semana de tratamento, mas é prudente não a utilizar nas etapas finais para evitar a proliferação excessiva de queratinócitos e, por consequência, a formação de uma cicatriz patológica. Os filmes de FS-NT deram suporte ao crescimento de células, além da modulação do processo de cicatrização. Estimulados pela iontoforese nas etapas iniciais desse processo, são promissores curativos para o tratamento de feridas crônicas.

9. REFERÊNCIAS

ABDEL-MOTTALEB, M. M. A.; NEUMANN, D.; LAMPRECHT, A. Lipid nanocapsules for dermal application: A comparative study of lipid-based versus polymer-based nanocarriers. **European Journal of Pharmaceutics and Biopharmaceutics**, v. 79, n. 1, p. 36–42, 2011.

ABLA, N. et al. Contributions of electromigration and electroosmosis to peptide iontophoresis across intact and impaired skin. **Journal of Controlled Release**, v. 108, n. 2–3, p. 319–330, 2005.

ÅGREN, M. S. et al. Collagenase in wound healing: Effect of wound age and type. **Journal of Investigative Dermatology**, v. 99, n. 6, p. 709–714, 1992.

AHER, N. D.; NAIR, H. A. Bilayered films based on novel polymer derivative for improved ocular therapy of gatifloxacin. **The Scientific World Journal**, v. 2014, 2014.

ALBERTI, K. G. M. M.; ZIMMET, P. Z. Definition, Diagnosis and Classification of Diabetes Mellitus and its Complications Part 1: Diagnosis and Classification of Diabetes Mellitus Provisional Report of a WHO Consultation. **Diabetic Medicine**, v. 15, p. 539–553, 1998.

ALTMAN, GREGORY H.; DIAZ, FRANK; JAKUBA, CAROLINE; CALABRO, TARA; HORAN, REBECA L.; CHEN, JINGSONG; LU, HELEN; RICHMOND, JOHN; KAPLAN, D. L. Silk-based biomaterials. **Biomaterials**, v. 3, p. 401–416, 2003.

ANDREWS, J. P. et al. **Keloids: The paradigm of skin fibrosis - Pathomechanisms and treatment** *Matrix Biology* Elsevier B.V., 2016.

ANSELL, D. M.; HOLDEN, K. A.; HARDMAN, M. J. Animal models of wound repair: Are they cutting it? **Experimental Dermatology**, v. 21, n. 8, p. 581–585, 2012.

APIKOGLU-RABUS, S. et al. Effect of topical insulin on cutaneous wound healing in rats with or without acute diabetes. **Clinical and Experimental Dermatology**, v. 35, n. 2, p. 180–185, 2010.

AZNAR-CERVANTES, S. D. et al. Influence of the protocol used for fibroin extraction on the mechanical properties and fiber sizes of electrospun silk mats. **Materials Science and Engineering C**, v. 33, n. 4, p. 1945–1950, 2013.

BALAKRISHNAN, B. et al. Evaluation of an in situ forming hydrogel wound dressing based on oxidized alginate and gelatin. **Biomaterials**, v. 26, n. 32, p. 6335–6342, nov. 2005.

BALBINO, C. A.; PEREIRA, LEONARDO MADEIRA; CURI, R. Mecanismos envolvidos na cicatrização: uma revisão. **Revista Brasileira de Ciências Farmacêuticas**, v. 41, n. 1, p. 27–51, 2005.

BANDYOPADHYAY, A.; MANDAL, B. B. A three-dimensional printed silk-based biomimetic tri-layered meniscus for potential patient-specific implantation.

Biofabrication, v. 12, n. 1, p. 015003, 21 out. 2019.

BARRIENTOS, S. et al. Growth factors and cytokines in wound healing. **Wound Repair and Regeneration**, v. 16, n. 5, p. 585–601, 2008.

BAŠIĆ-KES, V.; ZAVOREO, I.; ROTIM, K.; BORNSTEIN, N.; RUNDEK, T.; DEMARIN, V. RECOMMENDATIONS FOR DIABETIC POLYNEUROPATHY TREATMENT. **Acta Clin Croat**, v. 50, p. 289–302, 2011.

BAUM, CHRISTIAN L.; ARPEY, C. J. Normal Cutaneous Wound Healing : Clinical Correlation with cellular and molecular events. **Dermatologic Surgery**, v. 31, n. 6, p. 674–686, 2005.

BAYRAKTAR, O. et al. Silk fibroin as a novel coating material for controlled release of theophylline. **European Journal of Pharmaceutics and Biopharmaceutics**, v. 60, n. 3, p. 373–381, 2005.

BERMAN, B.; MADERAL, A.; RAPHAEL, B. **Keloids and hypertrophic scars: Pathophysiology, classification, and treatment** *Dermatologic Surgery* Lippincott Williams and Wilkins, 2017.

BERMUDEZ, D. M. et al. Impaired biomechanical properties of diabetic skin: Implications in pathogenesis of diabetic wound complications. **American Journal of Pathology**, v. 178, n. 5, p. 2215–2223, 2011.

BERNARDI, D. S. et al. Effective transcutaneous immunization using a combination of iontophoresis and nanoparticles. **Nanomedicine: Nanotechnology, Biology, and Medicine**, v. 12, n. 8, p. 2439–2448, 2016.

BHATTACHARJEE, P.; FERNÁNDEZ-PÉREZ, J.; AHEARNE, M. Potential for combined delivery of riboflavin and all-trans retinoic acid, from silk fibroin for corneal bioengineering. **Materials Science and Engineering C**, v. 105, n. 110093, 2019.

BLAŽIĆ, T. M.; BRAJAC, I. Defective induction of senescence during wound healing is a possible mechanism of keloid formation. **Medical Hypotheses**, v. 66, p. 649–652, 2006.

BOATENG, J. S. et al. Wound Healing Dressings and Drug Delivery Systems : A Review. **Journal of Pharmaceutical Sciences**, v. 97, n. 8, p. 2892–2923, 2008.

BOMMER, C. et al. The global economic burden of diabetes in adults aged 20–79 years: a cost-of-illness study. **The Lancet Diabetes and Endocrinology**, v. 5, n. 6, p. 423–430, 2017.

BOUWSTRA, J. A. et al. Structure of the skin barrier and its modulation by vesicular formulations. **Progress in Lipid Research**, v. 42, n. 1, p. 1–36, 2003.

BOWEN, W. R. et al. A new technique for membrane characterisation: direct measurement of the force of adhesion of a single particle using an atomic force microscope. **Journal of Membrane Science**, v. 139, n. 2, p. 269–274, 1998.

BRAY, L. J. et al. Human corneal epithelial equivalents constructed on Bombyx mori

silk fibroin membranes. **Biomaterials**, v. 32, n. 22, p. 5086–5091, 2011.

BREM, H. et al. Primary cultured fibroblasts derived from patients with chronic wounds: a methodology to produce human cell lines and test putative growth factor therapy such as GMCSF. **Journal of Translational Medicine**, v. 6, n. 75, 2008.

BREM, H.; TOMIC-CANIC, M. Cellular and molecular basis of wound healing in diabetes Find the latest version : Cellular and molecular basis of wound healing in diabetes. **The Journal of Clinical Investigation**, v. 117, n. 5, p. 1219–1222, 2007.

BRINKMANN, V. et al. Neutrophil Extracellular Traps Kill Bacteria. **Science**, v. 303, n. 5663, p. 1532–1535, 2004.

BROUGHTON II, GEORGE; JANIS, JEFFREY E.; ATTINGER, C. E. Wound Healing : An Overview. **Plastic and Reconstructive Surgery**, v. 117, n. 7, p. 1–32, 2006.

BROWNLEE, M. Biochemistry and molecular cell biology of diabetic complications. **Nature**, v. 414, p. 813–820, 2001.

BRUN, P. et al. Neuropeptide neurotensin stimulates intestinal wound healing following chronic intestinal inflammation. **AJP: Gastrointestinal and Liver Physiology**, v. 288, n. 4, p. G621–G629, 2005.

BUTLER, WILLIAM T.; CUNNINGHAM, L. W. Evidence for the Linkage of a Disaccharide to Hydroxylysine in tropocollagen. v. 241, p. 3882–3888, 1966.

CAMPOS, A. C. L.; GROTH, A. K.; BRANCO, A. B. Assessment and nutritional aspects of wound healing. **Current Opinion in Clinical Nutrition and Metabolic Care**, v. 11, p. 281–288, 2008.

CARRAWAY, R. E. et al. Presence of Neurotensin and Neuromedin-N Within a Common Precursor from a Human Pancreatic Neuroendocrine Tumor. **The Journal of Clinical Endocrinology & Metabolism**, v. 66, n. 6, p. 1323–1328, 1988.

CASSETTARI, L. L. et al. Continuous electrical current and zinc sulphate administered by transdermal iontophoresis improves skin healing in diabetic rats induced by alloxan: morphological and ultrastructural analysis. **Journal of diabetes research**, 2014.

CÁZARES-DELGADILLO, J. et al. Controlled transdermal iontophoresis for poly-pharmacotherapy: Simultaneous delivery of granisetron, metoclopramide and dexamethasone sodium phosphate in vitro and in vivo. **European Journal of Pharmaceutical Sciences**, v. 85, p. 31–38, 2016.

CEVC, G. Lipid vesicles and other colloids as drug carriers on the skin. **Advanced Drug Delivery Reviews**, v. 56, n. 5, p. 675–711, 2004.

CHARAN, J.; KANTHARIA, N. How to calculate sample size in animal studies? **Journal of Pharmacology and Pharmacotherapeutics**, v. 4, n. 4, p. 303–306, 2013.

CHENG, G. et al. Differences in regenerated silk fibroin prepared with different solvent systems: From structures to conformational changes. **Journal of Applied Polymer Science**, v. 132, n. 22, 2015.

CHO, H. J. et al. Molecular weight distribution and solution properties of silk fibroins with different dissolution conditions. **International Journal of Biological Macromolecules**, v. 51, n. 3, p. 336–341, 2012.

CHOI, J. U. et al. Preparation and in vivo evaluation of cationic elastic liposomes comprising highly skin-permeable growth factors combined with hyaluronic acid for enhanced diabetic wound-healing therapy. **Acta Biomaterialia**, v. 57, p. 197–215, 2017.

CIRILLO, B.; MORRA, M.; CATAPANO, G. Adhesion and Function of Rat Liver Cells Adherent to Silk Fibroin/Collagen Blend Films. **The International Journal of Artificial Organs**, v. 27, n. 1, p. 60–68, 2004.

CLARK, R. A. F.; GHOSH, K.; TONNESEN, M. G. Tissue engineering for cutaneous wounds. **Journal of Investigative Dermatology**, v. 127, n. 5, p. 1018–1029, 2007.

CORSETTI, G. et al. Topical application of dressing with amino acids improves cutaneous wound healing in aged rats. **Acta Histochemica**, v. 112, n. 5, p. 497–507, 2010.

CRUZ-CAZARIM, E. L. C. et al. Prospective insulin-based ophthalmic delivery systems for the treatment of dry eye syndrome and corneal injuries. **European Journal of Pharmaceutics and Biopharmaceutics**, v. 140, n. April, p. 1–10, 2019.

CUBAYACHI, C.; COUTO, R.O.; GAITANI, C.M.; PEDRAZZI, V.; FREITAS, O.; LOPEZ, R. F. V. Needle-free buccal anesthesia using iontophoresis and amino amide salts combined in a mucoadhesive formulation. **Colloids and Surfaces B: Biointerfaces**, v. 136, p. 1193–1201, 2015.

CUBAYACHI, C. et al. Silk fibroin films stabilizes and releases bioactive insulin for the treatment of corneal wounds. **European Polymer Journal**, v. 118, p. 502–513, 2019.

DA SILVA, L.; CARVALHO, E.; CRUZ, M. T. Role of neuropeptides in skin inflammation and its involvement in diabetic wound healing. **Expert Opinion on Biological Therapy**, v. 10, n. 10, p. 1427–1439, 2010.

DARBY, I. A. et al. Fibroblasts and myofibroblasts in wound healing. *Clinical, Cosmetic and Investigational Dermatology*, **Dove Medical Press Ltd.**, 2014.

DAVIS, C. P. et al. Quantification, qualification, and microbial killing efficiencies of antimicrobial chlorine-based substances produced by iontophoresis. **Antimicrobial Agents and Chemotherapy**, v. 38, n. 12, p. 2768–2774, 1994.

DE MORAES, M. A. et al. Silk fibroin and sodium alginate blend: Miscibility and physical characteristics. **Materials Science and Engineering C**, v. 40, p. 85–91, 2014.

DE PAIVA, C. S. et al. Corticosteroid and doxycycline suppress MMP-9 and inflammatory cytokine expression, MAPK activation in the corneal epithelium in experimental dry eye. **Experimental Eye Research**, v. 83, n. 3, p. 526–535, 2006.

DECKERS, J.; HAMMAD, H.; HOSTE, E. **Langerhans cells: Sensing the**

environment in health and disease *Frontiers in Immunology* Frontiers Media S.A., 2018. .

DEL POZO, J. L.; ROUSE, M. S.; PATEL, R. Bioelectric effect and bacterial biofilms. A systematic review. **The International journal of artificial organs**, v. 31, n. 9, p. 786–95, 2008.

DHIVYA, S.; PADMA, V. V.; SANTHINI, E. Wound dressings - A review. **BioMedicine**, v. 5, n. 4, p. 24–28, 2015.

DUTET, J.; DELGADO-CHARRO, M. B. In vivo transungual iontophoresis : Effect of DC current application on ionic transport and on transonychial water loss. **Journal of Controlled Release**, v. 140, n. 2, p. 117–125, 2009.

DYCK, P. J.; KRATZ, K.M.; KARNES, J. L.; LITCHY, W.J.; KLEIN, R.; PACH, J.M.; WILSON, D.M.; O'BRIEN, P.C.; MELTON III, L. J. The prevalence by staged severity of various types of diabetic neuropathy , retinopathy , and nephropathy in a population-based cohort : The Rochester Diabetic Neuropathy Study. **Neurology**, v. 43, n. April, p. 817–824, 1993.

EHRENREICH, M.; RUSZCZAK, Z. Update on Tissue-Engineered Biological Dressings. **Tissue Engineering**, v. 12, n. 9, p. 2407–2424, 2006.

ELLIOT, S. et al. A Modeling Conundrum: Murine Models for Cutaneous Wound Healing. **Journal of Investigative Dermatology**, v. 138, p. 736–740, 2018.

EMING, S. A.; MARTIN, P.; TOMIC-CANIC, M. Wound repair and regeneration: Mechanisms , signaling , and translation. **Science Translational Medicine**, v. 6, n. 265, 2014.

ENOCH, S.; LEAPER, D. J.; BELDON, P. Basic science of wound healing. **Surgery (Oxford)**, v. 28, n. 9, p. 409–412, 2010.

FANG, J. et al. Transdermal iontophoresis of 5-fluorouracil combined with electroporation and laser treatment. **International Journal of Pharmaceutics**, v. 270, p. 241–249, 2004.

FARBER, P. L. et al. Electricity and colloidal stability: How charge distribution in the tissue can affects wound healing. **Medical Hypotheses**, v. 82, n. 2, p. 199–204, 2014.

FENG, Q. L. et al. A mechanistic study of the antibacterial effect of silver ions on *Escherichia coli* and *Staphylococcus aureus*. **Journal of Biomedical Materials Research**, v. 52, n. 4, p. 662–668, 2000.

FERRANTE, C. J.; LEIBOVICH, S. J. Regulation of Macrophage Polarization and Wound Healing. **Advances in Wound Care**, v. 1, n. 1, p. 10–16, 2012.

FIERRO, I. M. et al. Induction of NOS in rat blood PMN in vivo and in vitro: Modulation by tyrosine kinase and involvement in bactericidal activity. **Journal of Leukocyte Biology**, v. 65, n. 4, p. 508–514, 1999.

FUCHS, S. et al. Outgrowth endothelial cells isolated and expanded from human

- peripheral blood progenitor cells as a potential source of autologous cells for endothelialization of silk fibroin biomaterials. **Biomaterials**, v. 27, n. 31, p. 5399–5408, 2006.
- FUKAI, T.; TAKEDA, A.; UCHINUMA, E. Wound healing in denervated rat skin. **Wound Repair and Regeneration**, v. 13, n. 2, p. 175–180, 2005.
- GETHIN, G. The significance of surface pH in chronic wounds. **Wounds UK**, v. 3, n. 3, p. 52–56, 2007.
- GHAZAWI, F. M. et al. Insights into the Pathophysiology of Hypertrophic Scars and Keloids. **Advances in Skin & Wound Care**, v. 31, n. 1, p. 582–595, 2018.
- GHOLOUPOURMALEKABADI, M. et al. Silk fibroin for skin injury repair: Where do things stand? **Advanced Drug Delivery Reviews**, 2019.
- GILABERTE, Y. et al. Anatomy and Function of the Skin. In: **Nanoscience in Dermatology**. [s.l.] Elsevier Inc., 2016. p. 1–14.
- GINHOUX, F.; GUILLIAMS, M. **Tissue-Resident Macrophage Ontogeny and HomeostasisImmunity** Cell Press, 2016. .
- GOSAIN, A.; DIPIETRO, L. A. Aging and Wound Healing. **World Journal of Surgery**, v. 28, n. 3, p. 321–326, 2004.
- GRATIERI, T.; GELFUSO, G. M.; THOMAZINI, J. A.; LOPEZ, R. F. V. Excised Porcine Cornea Integrity Evaluation in an in vitro Model of Iontophoretic Ocular Research. **Ophthalmic Research**, v. 43, n. 4, p. 208–216, 2010.
- GREENHALGH, D. G. Wound healing and diabetes mellitus. **Clinics in Plastic Surgery**, v. 30, p. 37–45, 2003.
- GRINNELL, F. Fibroblast biology in three-dimensional collagen matrices. **Trends in Cell Biology**, v. 13, n. 5, p. 264–269, 2003.
- GUO, S.; DIPIETRO, L. A. Critical review in oral biology & medicine: Factors affecting wound healing. **Journal of Dental Research**, v. 89, n. 3, p. 219–229, 2010.
- GUPTA, M. K. et al. Patterned silk films cast from ionic liquid solubilized fibroin as scaffolds for cell growth. **Langmuir**, v. 23, n. 3, p. 1315–1319, 2007.
- GURTNER, G. C. et al. Wound repair and regeneration. **Nature**, v. 453, n. 7193, p. 314–321, 2008.
- HARPER, D.; YOUNG, A.; MCNAUGHT, C.-E. The physiology of wound healing. **Surgery**, v. 32, n. 9, p. 445–450, 2014.
- HART, J. Inflammation 1: its role in the healing of acute wounds. **Wound Care**, v. 11, n. 6, p. 205–209, 2002.
- HE, J. et al. Stabilization of RNA Encapsulated in Silk. **ACS Biomaterials Science & Engineering**, p. acsbiomaterials.8b00207, 2018.

- HENDRIX, S.; PETERS, E. M. J. Neuronal plasticity and neuroregeneration in the skin - The role of inflammation. **Journal of Neuroimmunology**, v. 184, n. 1–2, p. 113–126, 2007.
- HENG, M. C. Y. Wound healing in adult skin: Aiming for perfect regeneration. **International Journal of Dermatology**, v. 50, n. 9, p. 1058–1066, 2011.
- HERMANS, K. et al. Development and characterization of mucoadhesive chitosan films for ophthalmic delivery of cyclosporine A. **International Journal of Pharmaceutics**, v. 472, n. 1–2, p. 10–19, 2014.
- HOARE, J. I. et al. Electric fields are novel determinants of human macrophage functions. **Journal of Leukocyte Biology**, v. 99, n. 6, p. 1141–1151, 2016.
- HOFER, M.; WINTER, G.; MYSCHIK, J. Recombinant spider silk particles for controlled delivery of protein drugs. **Biomaterials**, v. 33, n. 5, p. 1554–1562, 2012.
- HOFMANN, S. et al. Silk fibroin as an organic polymer for controlled drug delivery. **Journal of Controlled Release**, v. 111, n. 1–2, p. 219–227, 2006.
- HU, S. C. S.; LAN, C. C. E. High-glucose environment disturbs the physiologic functions of keratinocytes: Focusing on diabetic wound healing. **Journal of Dermatological Science**, v. 84, n. 2, p. 121–127, 2016.
- HU, X.; KAPLAN, D.; CEBE, P. Determining Beta-Sheet Crystallinity in Fibrous Proteins by Thermal Analysis and Infrared Spectroscopy. **Macromolecules**, v. 39, n. 18, p. 6161–6170, 2006.
- HUBER, L. A. et al. Topical skin cancer therapy using doxorubicin-loaded cationic lipid nanoparticles and iontophoresis. **Journal of Biomedical Nanotechnology**, v. 11, n. 11, p. 1975–1988, 2015.
- INTERNATIONAL STANDARD, I. ISO 10993-5:2009 - Biological evaluation of medical devices- Part 5: Tests for in vitro cytotoxicity, 2009.
- ISSEROFF, R. R.; DAHLE, S. E. Electrical Stimulation Therapy and Wound Healing: Where Are We Now? **Advances in Wound Care**, v. 1, n. 6, p. 238–243, 2012.
- JAMES, G. A. et al. Biofilms in chronic wounds. **Wound Repair and Regeneration**, v. 16, n. 1, p. 37–44, 2008.
- JAO, D.; MOU, X.; HU, X. Tissue Regeneration: A Silk Road. **Journal of Functional Biomaterials**, v. 7, n. 3, p. 22, 2016.
- JIN, H. J.; KAPLAN, D. L. Mechanism of silk processing in insects and spiders. **Nature**, v. 424, n. 6952, p. 1057–1061, 2003.
- JORGE, A. G. et al. Aspirin prevents diabetic oxidative changes in rat lacrimal gland structure and function. **Endocrine**, v. 35, n. 2, p. 189–197, 2009.
- JUN, J. II; LAU, L. F. The matricellular protein CCN1 induces fibroblast senescence and restricts fibrosis in cutaneous wound healing. **Nature Cell Biology**, v. 12, n. 7, p.

676–685, 2010.

JÜNGER, M. et al. Local therapy and treatment costs of chronic, venous leg ulcers with electrical stimulation (Dermapulse®): A prospective, placebo controlled, double blind trial. **Wound Repair and Regeneration**, v. 16, n. 4, p. 480–487, 2008.

JUNQUEIRA, L. C.; CARNEIRO, J. **Histologia básica**, 9 edição; 2008.

KALIA, Y. N. et al. Iontophoretic drug delivery. **Advanced Drug Delivery Reviews**, v. 56, n. 5, p. 619–658, 2004.

KALNIN, N. N.; BAIKALOV, I. A.; VENYAMINOV, S. Y. Quantitative IR spectrophotometry of peptide compounds in water (H₂O) solutions. III. Estimation of the protein secondary structure. **Biopolymers**, v. 30, n. 13–14, p. 1273–1280, 1990.

KAMOUN, E. A.; KENAWY, E. S.; CHEN, X. REVIEW A review on polymeric hydrogel membranes for wound dressing applications: PVA-based hydrogel dressings. **Journal of Advanced Research**, v. 8, n. 3, p. 217–233, 2017.

KANITAKIS, J. Anatomy, histology and immunohistochemistry of normal human skin. **European journal of dermatology**, v. 12, n. 4, p. 390–401, 2002.

KANT, V. et al. ScienceDirect Combined effect of substance P and curcumin on cutaneous wound healing in diabetic rats. **Journal of Surgical Research**, v. 212, p. 130–145, 2017.

KARAGEORGIU, V. et al. Porous silk fibroin 3-D scaffolds for delivery of bone morphogenetic protein-2 in vitro and in vivo. **Journal of Biomedical Materials Research - Part A**, v. 78, n. 2, p. 324–334, 2006.

KAWASAKI, L. et al. The mechanisms and evidence of efficacy of electrical stimulation for healing of pressure ulcer: A systematic review. **Wound Repair and Regeneration**, v. 22, n. 2, p. 161–173, 2014.

KIM, D. W. et al. Effect of silk fibroin peptide derived from silkworm *Bombyx mori* on the anti-inflammatory effect of Tat-SOD in a mice edema model. **BMB Reports**, v. 44, n. 12, p. 787–792, 2011.

KIM, U. J. et al. Structure and properties of silk hydrogels. **Biomacromolecules**, v. 5, n. 3, p. 786–792, 2004.

KLECZKOWSKA, P.; LIPKOWSKI, A. W. Neurotensin and neurotensin receptors: Characteristic, structure-activity relationship and pain modulation - A review. **European Journal of Pharmacology**, v. 716, n. 1–3, p. 54–60, 2013.

KLOTH, L. C. Electrical Stimulation Technologies for Wound Healing. **Advances in Wound Care**, v. 3, n. 2, p. 81–90, 2014.

KO, M. S.; MARINKOVICH, M. P. Role of Dermal- Epidermal Basement Membrane Zone in Skin, Cancer, and Developmental Disorders. **Dermatologic Clinics**, v. 28, n. 1, p. 1–16, 2010.

KOH, L. et al. Progress in Polymer Science Structures , mechanical properties and applications of silk fibroin materials. **Progress in Polymer Science**, v. 46, p. 86–110, 2015. D

KOLACZKOWSKA, E.; KUBES, P. **Neutrophil recruitment and function in health and inflammation** *Nature Reviews Immunology*, 2013.

KUBO, A.; KAJIMURA, M.; SUEMATSU, M. Matrix-Assisted Laser Desorption/Ionization (MALDI) Imaging Mass Spectrometry (IMS): A Challenge for Reliable Quantitative Analyses. **Mass Spectrometry**, v. 1, n. 1, p. A0004–A0004, 2012.

KUNDU, B. et al. Silk fibroin biomaterials for tissue regenerations ☆. **Advanced Drug Delivery Reviews**, v. 65, n. 4, p. 457–470, 2013.

LAMMEL, A. S. et al. Controlling silk fibroin particle features for drug delivery. **Biomaterials**, v. 31, n. 16, p. 4583–4591, 2010.

LEBERT, D. C.; HUTTENLOCHER, A. Inflammation and wound repair. **Seminars in Immunology**, v. 26, n. 4, p. 315–320, 2014.

LEMOS, C. N. et al. Iontophoresis Improved Growth Reduction of Invasive Squamous Cell Carcinoma in Topical Photodynamic Therapy. **PLOS ONE**, v. 11, n. 1, p. e0145922, 2016.

LEMOS, C. N. et al. European Journal of Pharmaceutics and Biopharmaceutics Iontophoresis-stimulated silk fibroin films as a peptide delivery system for wound healing. **European Journal of Pharmaceutics and Biopharmaceutics**, v. 128, n. December 2017, p. 147–155, 2018.

LI, A. B. et al. Silk-based stabilization of biomacromolecules. **Journal of Controlled Release**, v. 219, p. 416–430, 2015.

LI, B.; WANG, J. H. C. Fibroblasts and myofibroblasts in wound healing: Force generation and measurement. **Journal of Tissue Viability**, v. 20, n. 4, p. 108–120, 2011.

LI, M. et al. Structure and properties of silk fibroin–poly(vinyl alcohol) gel. **International Journal of Biological Macromolecules**, v. 30, p. 89–94, 2002.

LI, X. et al. Functionalized silk fibroin dressing with topical bioactive insulin release for accelerated chronic wound healing. **Materials Science and Engineering C**, v. 72, p. 394–404, 2017.

LIAU, S. Y. et al. Interaction of silver nitrate with readily identifiable groups: relationship to the antibacterial action of silver ions. **Letters in Applied Microbiology**, v. 25, n. 4, p. 279–283, set. 1997.

LINDHOLM, C.; SEARLE, R. Wound management for the 21st century: combining effectiveness and efficiency. **International Wound Journal**, v. 13, p. 5–15, 2016.

LIU, J. et al. A bifunctional biosensor for subcutaneous glucose monitoring by reverse

- iontophoresis. **Journal of Electroanalytical Chemistry**, v. 660, n. 1, p. 8–13, 2011.
- LIU, J. et al. Controlled-release neurotensin-loaded silk fibroin dressings improve wound healing in diabetic rat model. **Bioactive Materials**, v. 4, p. 151–159, 2019.
- LJUBIMOV, A. V.; SAGHIZADEH, M. **Progress in corneal wound healing Progress in Retinal and Eye Research**, Elsevier Ltd, , 2015.
- LOPEZ, R. F. V. et al. Photodynamic therapy of skin cancer: Controlled drug delivery of 5-ALA and its esters. **Advanced Drug Delivery Reviews**, v. 56, n. 1, p. 77–94, 2004.
- LOSQUADRO, W. D. Anatomy of the Skin and the Pathogenesis of Nonmelanoma Skin Cancer. **Facial Plastic Surgery Clinics of North America**, v. 25, n. 3, p. 283–289, 2017.
- LU, Q. et al. Water-insoluble silk films with silk I structure. **Acta Biomaterialia**, v. 6, n. 4, p. 1380–1387, 2010a.
- LU, S. et al. Insoluble and flexible silk films containing glycerol. **Biomacromolecules**, v. 11, n. 1, p. 143–150, 2010b.
- MAGOSHI, J. et al. Physical properties and structure of silk. VI. Conformational changes in silk fibroin induced by immersion in water at 2 to 130°C. **J. Polym. Sci. Pol. Phys.**, v. 17, n. 3, p. 515–520, 1979.
- MAKRANTONAKI, E. et al. Diabetes mellitus and the skin. **Reviews in Endocrine and Metabolic Disorders**, 2016.
- MANTOVANI, A. et al. **Neutrophils in the activation and regulation of innate and adaptive immunity Nature Reviews Immunology**, 2011. .
- MANTOVANI, A. et al. Macrophage plasticity and polarization in tissue repair and remodelling. **The Journal of Pathology**, v. 229, n. 2, p. 176–185, 2013.
- MARRO, D. et al. Contributions of electromigration and electroosmosis to iontophoretic drug delivery. **Pharmaceutical Research**, v. 18, n. 12, p. 1701–1708, 2001.
- MARTIN, P. Wound healing - Aiming for perfect skin regeneration. **Science**, v. 276, n. 5309, p. 75–81, 1997.
- MARTIN, S.; VINCENT, J.-P.; MAZELLA, J. Involvement of the Neurotensin Receptor-3 in the Neurotensin-Induced Migration of Human Microglia. **The Journal of Neuroscience**, v. 23, n. 4, p. 1198–1205, 2003.
- MATHIEU, D.; LINKE, J.-C.; WATTEL, F. Non-Healing Wounds. In: **Handbook on Hyperbaric Medicine**. Berlin/Heidelberg: Springer-Verlag, 2006.
- MAYADAS, T. N.; CULLERE, X.; LOWELL, C. A. The Multifaceted Functions of Neutrophils. **Annual Review of Pathology: Mechanisms of Disease**, v. 9, n. 1, p. 181–218, 2014.

- MCCLEMENTS, D. J. Non-covalent interactions between proteins and polysaccharides. **Biotechnology Advances**, v. 24, n. 6, p. 621–625, 2006.
- MEINEL, L. et al. Silk implants for the healing of critical size bone defects. **Bone**, v. 37, n. 5, p. 688–698, 2005.
- MELKE, J. et al. Silk fibroin as biomaterial for bone tissue engineering. **Acta Biomaterialia**, v. 31, p. 1–16, 1 fev. 2016.
- MENKE, N. B. et al. Impaired wound healing. **Clinics in Dermatology**, v. 25, n. 1, p. 19–25, 2007.
- MENON, G. K. New insights into skin structure: Scratching the surface. **Advanced Drug Delivery Reviews**, v. 54, p. S3–S17, 2002.
- MERINO, V.; KALIA, Y. N.; GUY, R. H. Transdermal therapy and diagnosis by iontophoresis. **Trends in Biotechnology**, v. 15, n. 8, p. 288–290, 1997.
- MOORE, K. et al. Impact of the Controlled Release of a Connexin 43 Peptide on Corneal Wound Closure in an STZ Model of Type I Diabetes. **PLoS ONE**, v. 9, n. 1, p. e86570, 23 jan. 2014.
- MORALES, J. O.; MCCONVILLE, J. T. Manufacture and characterization of mucoadhesive buccal films. **European Journal of Pharmaceutics and Biopharmaceutics**, v. 77, n. 2, p. 187–199, 2011.
- MOREAU, J. E. et al. Sequential growth factor application in bone marrow stromal cell ligament engineering. **Tissue Engineering**, v. 11, n. 11–12, p. 1887–1897, 2005.
- MORONES, J. R. et al. The bactericidal effect of silver nanoparticles. **Nanotechnology**, v. 16, n. 10, p. 2346–2353, 2005.
- MORTON, LAUREL M.; PHILLIPS, T. J. Differential diagnosis and evaluation of chronic wounds. **Journal of American Dermatology**, v. 74, n. 4, p. 589–605, 2016.
- MOSSER, D. M.; EDWARDS, J. P. Exploring the full spectrum of macrophage activation. **Nature Publishing Group**, v. 8, n. 12, p. 958–969, 2008.
- MOURA, L. I. F. et al. Recent advances on the development of wound dressings for diabetic foot ulcer treatment - A review. **Acta Biomaterialia**, v. 9, n. 7, p. 7093–7114, 2013a.
- MOURA, L. I. F. et al. Neurotensin modulates the migratory and inflammatory response of macrophages under hyperglycemic conditions. **BioMed Research International**, v. 2013, 2013b.
- MOURA, L. I. F. et al. Neurotensin-loaded collagen dressings reduce inflammation and improve wound healing in diabetic mice. **Biochimica et Biophysica Acta - Molecular Basis of Disease**, v. 1842, n. 1, p. 32–43, 2014.
- MOURITZEN, M. V. et al. Neurotensin, substance P, and insulin enhance cell migration. **Journal of Peptide Science**, v. 24, n. 7, 2018.

NAKAZAWA, Y. et al. Development of small-diameter vascular grafts based on silk fibroin fibers from bombyx mori for vascular regeneration. **Journal of Biomaterials Science, Polymer Edition**, v. 22, n. 1–3, p. 195–206, 2011.

NAVARRE, W. W.; SCHNEEWIND, O. Surface proteins of gram-positive bacteria and mechanisms of their targeting to the cell wall envelope. **Microbiology and molecular biology reviews : MMBR**, v. 63, n. 1, p. 174–229, 1999.

NICKOLOFF, B. J. et al. Tumor Suppressor Maspin Is Up-Regulated during Keratinocyte Senescence, Exerting a Paracrine Antiangiogenic Activity. **Cancer Research**, v. 64, n. 9, p. 2956–2961, 2004.

NOMIYA, K. et al. Synthesis and structural characterization of silver(I), aluminium(III) and cobalt(II) complexes with 4-isopropyltropolone (hinokitiol) showing noteworthy biological activities. Action of silver(I)-oxygen bonding complexes on the antimicrobial activities. **Journal of Inorganic Biochemistry**, v. 98, n. 1, p. 46–60, 2004.

NOSENKO, M. A.; AMBARYAN, S. G.; DRUTSKAYA, M. S. Proinflammatory Cytokines and Skin Wound Healing in Mice. **Molecular Biology**, v. 53, n. 5, p. 741–754, 2019.

NUNAN, R.; HARDING, K. G.; MARTIN, P. Clinical challenges of chronic wounds: searching for an optimal animal model to recapitulate their complexity. **Disease Models & Mechanisms**, v. 7, n. 11, p. 1205–1213, 2014.

NUNES, K. M. et al. The Monoglyceride Content Affects the Self-Assembly Behavior, Rheological Properties, Syringeability, and Mucoadhesion of In Situ–Gelling Liquid Crystalline Phase. **Journal of Pharmaceutical Sciences**, v. 105, n. 8, p. 2355–2364, 2016.

PARK, H.-Y. et al. A long-standing hyperglycaemic condition impairs skin barrier by accelerating skin ageing process. **Experimental Dermatology**, v. 20, n. 12, p. 969–974, dez. 2011.

PAULO, S. M. da S. de S. **Atenção à Saúde - Protocolo de prevenção e tratamento de feridas** Secretaria Municipal de Saúde de São Paulo, 2016.

PAWAR, H. V; TETTEH, J.; BOATENG, J. S. Preparation, optimisation and characterisation of novel wound healing film dressings loaded with streptomycin and diclofenac. **Colloids and Surfaces B: Biointerfaces**, v. 102, p. 102–110, 2013.

PEREIRA, L. et al. Neurotensin Decreases the Proinflammatory Status of Human Skin Fibroblasts and Increases Epidermal Growth Factor Expression. **International Journal of Inflammation**, v. 2014, p. 1–9, 2014.

PETROFSKY, J. et al. Effects of a 2-, 3- and 4-electrode stimulator design on current dispersion on the surface and into the limb during electrical stimulation in controls and patients with wounds. **Journal of Medical Engineering and Technology**, v. 32, n. 6, p. 485–497, 2008.

PODSTAWKA-PRONIEWICZ, E. et al. Structure of Monolayers Formed from Neurotensin and Its Single-Site Mutants: Vibrational Spectroscopic Studies. **The**

Journal of Physical Chemistry B, v. 115, n. 20, p. 6709–6721, 2011.

POLAK, A.; FRANEK, A.; TARADAJ, J. High-Voltage Pulsed Current Electrical Stimulation in Wound Treatment. **Advances in Wound Care**, v. 3, n. 2, p. 104–117, 2014.

PRADHAN, L. et al. Gene expression of pro-inflammatory cytokines and neuropeptides in diabetic wound healing. **Journal of Surgical Research**, v. 167, n. 2, p. 336–342, 2011.

PRITCHARD, E. M. et al. Encapsulation of oil in silk fibroin biomaterials. **Journal of Applied Polymer Science**, v. 131, n. 6, 2014.

PROST-SQUARCIONI, C. **Histology of skin and hair follicle; Medecine Sciences**, fev. 2006.

PU, J.; ZHAO, M. Golgi polarization in a strong electric field. **Journal of Cell Science**, v. 118, n. 6, p. 1117–1128, 2005.

QING, C. The molecular biology in wound healing & non-healing wound. **Chinese Journal of Traumatology - English Edition**, v. 20, n. 4, p. 189–193, 2017.

RAMASASTRY, S. S. Acute wounds. **Clinics in Plastic Surgery**, v. 32, n. 2, p. 195–208, 2005.

RAMOT, Y.; PAUS, R. Harnessing neuroendocrine controls of keratin expression: A new therapeutic strategy for skin diseases? **BioEssays**, v. 36, n. 7, p. 672–686, 2014.

RAWDANOWICZ, T. J. et al. Matrix metalloproteinase production by cultured human endometrial stromal cells: identification of interstitial collagenase, gelatinase-A, gelatinase-B, and stromelysin-1 and their differential regulation by interleukin-1 alpha and tumor necrosis factor-alpha. **The Journal of Clinical Endocrinology & Metabolism**, v. 79, n. 2, p. 530–536, 1994.

REIS, M. B. et al. Lipoxin A4 encapsulated in PLGA microparticles accelerates wound healing of skin ulcers. **PLoS ONE**, v. 12, n. 7, p. 1–15, 2017.

RICHARD BOWEN, W.; DONEVA, T. A. Atomic Force Microscopy Studies of Membranes: Effect of Surface Roughness on Double-Layer Interactions and Particle Adhesion. **Journal of Colloid and Interface Science**, v. 229, n. 2, p. 544–549, 2000.

RIDIANDRIES, A.; TAN, J.; BURSILL, C. The Role of Chemokines in Wound Healing. **International Journal of Molecular Sciences**, v. 19, n. 10, p. 3217, 2018.

ROBSON, M. C.; STEED, D. L.; FRANZ, M. G. Wound Healing: Biologic Features and Approaches to Maximize Healing Trajectories. **Current Problems in Surgery**, v. 38, n. 2, p. 1–140, 2001.

ROCKWOOD, D. N. et al. Materials fabrication from Bombyx mori silk fibroin. **Nature Protocols**, v. 6, n. 10, p. 1612–1631, 2011.

RODRIGUEZ-NOGALES, A. et al. Effect of aqueous and particulate silk fibroin in a rat

- model of experimental colitis. **International Journal of Pharmaceutics**, v. 511, n. 1, p. 1–9, 2016.
- ROH, D. H. et al. Wound healing effect of silk fibroin/alginate-blended sponge in full thickness skin defect of rat. **Journal of Materials Science: Materials in Medicine**, v. 17, n. 6, p. 547–552, 2006.
- ROLDÁN-MARÍN, R. et al. Fixed sporotrichosis as a cause of a chronic ulcer on the knee. **International Wound Journal**, v. 6, n. 1, p. 63–66, 2009.
- ROTTY, J. D.; COULOMBE, P. A. A wound-induced keratin inhibits Src activity during keratinocyte migration and tissue repair. **Journal of Cell Biology**, v. 197, n. 3, p. 381–389, 2012.
- SANTIN, M. et al. In vitro evaluation of the inflammatory potential of the silk fibroin. **Journal of Biomedical Materials Research**, v. 46, n. 3, p. 382–389, 1999.
- SATO, M. et al. In vivo introduction of the interleukin 6 gene into human keratinocytes: Induction of epidermal proliferation by the fully spliced form of interleukin 6, but not by the alternatively spliced form. **Archives of Dermatological Research**, v. 291, n. 7–8, p. 400–404, 1999.
- SCARPA, R. C.; CARRAWAY, R. E.; COCHRANE, D. E. Insulin-like growth factor (IGF) induced proliferation of human lung fibroblasts is enhanced by neurotensin. **Peptides**, v. 26, n. 11, p. 2201–2210, 2005.
- SERVOLI, E. et al. Surface properties of silk fibroin films and their interaction with fibroblasts. **Macromolecular Bioscience**, v. 5, n. 12, p. 1175–1183, 2005.
- SHANG, S.; ZHU, L.; FAN, J. Intermolecular interactions between natural polysaccharides and silk fibroin protein. **Carbohydrate Polymers**, v. 93, n. 2, p. 561–573, 2013.
- SIDDIQUI, A. R.; BERNSTEIN, J. M. Chronic wound infection: Facts and controversies. **Clinics in Dermatology**, v. 28, n. 5, p. 519–526, 2010.
- SINGH, A. et al. Meta-analysis of randomized controlled trials on hydrocolloid occlusive dressing versus conventional gauze dressing in the healing of chronic wounds. **Asian Journal of Surgery**, v. 27, n. 4, p. 326–332, 2004.
- SO, T. et al. Tumor Necrosis Factor- α Stimulates the Biosynthesis of Matrix Metalloproteinases and Plasminogen Activator in Cultured Human Chorionic Cells. **Biology of Reproduction**, v. 46, n. 5, p. 772–778, 1992.
- SONG, B. et al. Application of direct current electric fields to cells and tissues in vitro and modulation of wound electric field in vivo. **Nature Protocols**, v. 2, n. 6, p. 1479–1489, 2007.
- SOUZA, J. G. et al. Topical delivery of ocular therapeutics: carrier systems and physical methods. **Journal of Pharmacy and Pharmacology**, v. 66, n. 4, p. 507–530, 2014.

STEED, D. L. Wound-healing trajectories. **Surgical Clinics of North America**, v. 83, n. 3, p. 547–555, 2003.

STELNICKI, E. J. et al. Nerve dependency in scarless fetal wound healing. **Plastic and Reconstructive Surgery**, v. 105, n. 1, p. 140–147, 2000.

SU, Y.; RICHMOND, A. Chemokine Regulation of Neutrophil Infiltration of Skin Wounds. **Advances in Wound Care**, v. 4, n. 11, p. 631–640, 2015.

SUH, H. et al. A New Electrode Design to Improve Outcomes in the Treatment of Chronic Non-Healing Wounds in Diabetes. **Diabetes Technology & Therapeutics**, v. 11, n. 5, p. 315–322, 2009.

SWAIN, S. D.; ROHN, T. T.; QUINN, M. T. Neutrophil priming in host defense: Role of oxidants as priming agents. **Antioxidants and Redox Signaling**, v. 4, n. 1, p. 69–83, 2002.

SWEITZER, SARAH M.; FANN, STEPHEN A.; BORG, THOMAS K.; BAYNES, JOHN W.; YOST, M. J. What Is the Future of Diabetic Wound Care? today's educator. **The Diabetes Educator**, v. 32, n. 2, p. 197–210, 2006.

TANAKA, K. et al. Determination of the site of disulfide linkage between heavy and light chains of silk fibroin produced by *Bombyx mori*. **Biochimica et Biophysica Acta - Protein Structure and Molecular Enzymology**, v. 1432, n. 1, p. 92–103, 1999.

TAVEIRA, S. F. et al. Development of cationic solid lipid nanoparticles with factorial design-based studies for topical administration of doxorubicin. **Journal of Biomedical Nanotechnology**, v. 8, n. 2, p. 219–228, 2012.

TOYODA, M. et al. Anti-cancer vaccination by transdermal delivery of antigen peptide-loaded nanogels via iontophoresis. **International Journal of Pharmaceutics**, v. 483, n. 1–2, p. 110–114, 2015.

TURABELIDZE, A.; DIPIETRO, L. A. Inflammation and wound healing. **Endodontic Topics**, v. 24, n. 1, p. 26–38, 2011.

UEBERSAX, L. et al. Silk fibroin matrices for the controlled release of nerve growth factor (NGF). **Biomaterials**, v. 28, n. 30, p. 4449–4460, 2007.

UEBERSAX, L.; MERKLE, H. P.; MEINEL, L. Insulin-like growth factor I releasing silk fibroin scaffolds induce chondrogenic differentiation of human mesenchymal stem cells. **Journal of Controlled Release**, v. 127, n. 1, p. 12–21, 2008.

UM, I. C. et al. Structural characteristics and properties of the regenerated silk fibroin prepared from formic acid. **International Journal of Biological Macromolecules**, v. 29, n. 2, p. 91–97, 2001.

UNIVERSIDADE ABERTA DO SUS - UNA SUS. Fundamentação Teórica: Feridas. v. 3, p. 1–11, 2011.

VALENTINO, L. A. Matrix-assisted laser desorption/ionization (MALDI) imaging mass spectrometry (IMS): Peering into the cup of Jamshid. **Haemophilia**, v. 20, n. 3, p. 304–

305, 2014.

VELNAR, T.; BAILEY, T.; SMRKOLJ, V. The Wound Healing Process: an Overview of the Cellular and Molecular Mechanisms. **The Journal of International Medical Research**, v. 37, 2009.

VENTURA, S. A. et al. Bioelectrochemistry Cortisol extraction through human skin by reverse iontophoresis. **Bioelectrochemistry**, v. 114, p. 54–60, 2017.

VENUS, M.; WATERMAN, J.; MCNAB, I. Basic physiology of the skin. **Surgery**, v. 29, n. 10, p. 471–474, 2011.

VEPARI, C.; KAPLAN, D. L. Silk as a biomaterial. **Progress in Polymer Science**, v. 32, n. 8–9, p. 991–1007, 2007.

VEPARI, C. P.; KAPLAN, D. L. Covalently immobilized enzyme gradients within three-dimensional porous scaffolds. **Biotechnology and Bioengineering**, v. 93, n. 6, p. 1130–1137, 2006.

VIEIRA, CHRYSTIANY PLACIDO DE BRITO; ARAÚJO, T. M. E. Prevalence and factors associated with chronic wounds in older adults in primary care*. **Journal of School of Nursing**, v. 52, n. 03415, 2018.

WANG, Y. et al. Cartilage tissue engineering with silk scaffolds and human articular chondrocytes. **Biomaterials**, v. 27, n. 25, p. 4434–4442, 2006.

WANG, Z. et al. Improved MALDI imaging MS analysis of phospholipids using graphene oxide as new matrix. **Scientific Reports**, v. 7, n. March, p. 1–9, 2017.

WENK, E.; MERKLE, H. P.; MEINEL, L. Silk fibroin as a vehicle for drug delivery applications. **Journal of Controlled Release**, v. 150, n. 2, p. 128–141, 2011.

WERNER, S.; GROSE, R. Regulation of wound healing by growth factors and cytokines *Physiological Reviews*, **American Physiological Society**, 2003.

WEST, H. C.; BENNETT, C. L. Redefining the role of langerhans cells as immune regulators within the skin *Frontiers in Immunology*, **Frontiers Media S.A.**, 2018.

WITTE, M. B.; BARBUL, A. General principles of wound healing. **Surgical Clinics of North America**, v. 77, n. 3, p. 509–528, 1997.

YADAV, A.; GUPTA, A. Noninvasive red and near-infrared wavelength-induced photobiomodulation: promoting impaired cutaneous wound healing. **Photodermatology Photoimmunology and Photomedicine**, v. 33, n. 1, p. 4–13, 2017.

YAGI, T. et al. Preparation of double-raschel knitted silk vascular grafts and evaluation of short-term function in a rat abdominal aorta. **Journal of Artificial Organs**, v. 14, n. 2, p. 89–99, 2011.

YAN, C. et al. Targeting Imbalance between IL-1 β and IL-1 Receptor Antagonist Ameliorates Delayed Epithelium Wound Healing in Diabetic Mouse Corneas.

American Journal of Pathology, v. 186, n. 6, p. 1466–1480, 2016.

YAN, H. B. et al. Biosynthesis of insulin-silk fibroin nanoparticles conjugates and in vitro evaluation of a drug delivery system. **Journal of Nanoparticle Research**, v. 11, n. 8, p. 1937–1946, 2009.

YANG, M. C. et al. The cardiomyogenic differentiation of rat mesenchymal stem cells on silk fibroin-polysaccharide cardiac patches in vitro. **Biomaterials**, v. 30, n. 22, p. 3757–3765, 2009.

YANG, S. wei et al. Comparison of the histological morphology between normal skin and scar tissue. **Journal of Huazhong University of Science and Technology - Medical Science**, v. 36, n. 2, p. 265–269, 2016.

YANG, Y. et al. Biocompatibility evaluation of silk fibroin with peripheral nerve tissues and cells in vitro. **Biomaterials**, v. 28, n. 9, p. 1643–1652, mar. 2007a.

YANG, Y. et al. Development and evaluation of silk fibroin-based nerve grafts used for peripheral nerve regeneration. **Biomaterials**, v. 28, n. 36, p. 5526–5535, 2007b.

YUCEL, T. et al. Silk fibroin rods for sustained delivery of breast cancer therapeutics. **Biomaterials**, v. 35, n. 30, p. 8613–8620, 2014.

ZARRINTAJ, P. et al. Can regenerative medicine and nanotechnology combine to heal wounds? the search for the ideal wound dressing. **Nanomedicine**, v. 12, n. 19, p. 2403–2422, 2017.

ZHANG, F. et al. Facile fabrication of robust silk nanofibril films via direct dissolution of silk in CaCl₂-formic acid solution. **ACS Applied Materials and Interfaces**, v. 7, n. 5, p. 3352–3361, 2015.

ZHANG, X.; REAGAN, M. R.; KAPLAN, D. L. **Electrospun silk biomaterial scaffolds for regenerative medicine** *Advanced Drug Delivery Reviews*, 2009.

ZHAO, M. et al. Electrical stimulation directly induces pre-angiogenic responses in vascular endothelial cells by signaling through VEGF receptors. **Journal of Cell Science**, v. 117, n. 3, p. 397–405, 2004.

ZHAO, M. et al. Electrical signals control wound healing through phosphatidylinositol-3-OH kinase- γ and PTEN. **Nature**, v. 442, n. 7101, p. 457–460, 2006.

ZHAO, M. Electrical fields in wound healing-An overriding signal that directs cell migration. **Seminars in Cell and Developmental Biology**, v. 20, n. 6, p. 674–682, 2009.

ZHONG, S. P.; ZHANG, Y. Z.; LIM, C. T. Tissue scaffolds for skin wound healing and dermal reconstruction. **Wiley Interdisciplinary Reviews: Nanomedicine and Nanobiotechnology**, v. 2, n. 5, p. 510–525, 2010.

ZHOU, C. Z. et al. Silk fibroin: Structural implications of a remarkable amino acid sequence. **Proteins: Structure, Function and Genetics**, v. 44, n. 2, p. 119–122, 2001.

ZHOU, F. et al. Silk fibroin-chondroitin sulfate scaffold with immuno-inhibition property for articular cartilage repair. **Acta Biomaterialia**, v. 63, p. 64–75, 2017.

ZHOU, H. et al. Genipin-crosslinked polyvinyl alcohol/silk fibroin/nano-hydroxyapatite hydrogel for fabrication of artificial cornea scaffolds—a novel approach to corneal tissue engineering. **Journal of Biomaterials Science, Polymer Edition**, v. 30, n. 17, p. 1604–1619, 2019.

ZHOU, J. et al. Regenerated silk fibroin films with controllable nanostructure size and secondary structure for drug delivery. **ACS Applied Materials and Interfaces**, v. 6, n. 24, p. 21813–21821, 2014.

ZHU, Y.; MEHTA, K. A.; MCGINITY, J. W. Influence of plasticizer level on the drug release from sustained release film coated and hot-melt extruded dosage forms. **Pharmaceutical Development and Technology**, v. 11, n. 3, p. 285–294, 2006.