

**ANDRESSA BOLSONI LOPES**

***EFEITOS DO ÁCIDO PALMITOLEICO NA CAPTAÇÃO E METABOLISMO  
DE GLICOSE E TRIACILGLICEROL EM ADIPÓCITOS BRANCOS***

Tese apresentada ao Programa de Pós-Graduação em Fisiologia Humana do Instituto de Ciências Biomédicas da Universidade de São Paulo, para obtenção do Título de Doutor em Ciências.

Área de Concentração: Fisiologia Humana

Orientador: Prof. Dr. Fábio Bessa Lima  
Coorientadora: Prof<sup>a</sup>. Dr<sup>a</sup>. Maria Isabel C. Alonso Vale

Versão original

São Paulo  
2014

## RESUMO

Bolsoni-Lopes A. Efeitos do ácido palmitoleico na captação e metabolismo de glicose e triacilglicerol em adipócitos brancos. [tese (Doutorado em Fisiologia Humana)]. São Paulo: Instituto de Ciências Biomédicas, Universidade de São Paulo; 2014.

Neste trabalho nós testamos a hipótese que o ácido palmitoleico, um ácido graxo que aumenta a captação de glicose e sensibilidade à insulina no músculo esquelético e fígado e suprime a esteatose hepática, modula o metabolismo de glicose e triacilglicerol (TAG) em adipócitos brancos. Para isso, células 3T3-L1 diferenciadas, tratadas com ácido palmitoleico (16:1n7, 200  $\mu$ M) ou ácido palmítico (16:0, 200  $\mu$ M) por 24h e adipócitos primários epididimais de camundongos C57BL6 selvagens ou deficientes para PPAR $\alpha$ , tratados com 16:1n7 (300 mg/kg/dia) ou ácido oleico (18:1n9, 300 mg/kg/dia), via gavagem durante 10 dias, foram avaliados quanto a captação de glicose e a sua oxidação, conversão em lactato e incorporação em ácidos graxos e glicerol de TAG, bem como, a incorporação de acetato em ácidos graxos, lipólise, síntese de TAG e expressão gênica e proteica de proteínas reguladoras das vias estudadas. Nossos resultados demonstram que o tratamento com 16:1n7, mas não com 18:1n9 (*in vivo*) ou 16:0 (*in vitro*), aumenta a captação de glicose basal e estimulada por insulina e o conteúdo de GLUT4 e phosphoThr172AMPK $\alpha$ , sem afetar o GLUT1 ou phosphoSer473AKT. Tal aumento de GLUT4 induzido por 16:1n7 foi abolido pela inibição farmacológica da AMPK. Além disso, o 16:1n7 aumenta a conversão de glicose em lactato e CO<sub>2</sub> (oxidação de glicose) e diminui a síntese *de novo* de ácidos graxos *in vitro*. Corroborando com esses achados, a atividade das enzimas glicose-6-fosfato desidrogenase e ATP citrato liase também foram reduzidas pelo tratamento de células 3T3-L1 com 16:1n7. Ainda, o tratamento de células 3T3-L1 com 16:1n7, mas não com 16:0, aumentou a lipólise basal e estimulada por isoproterenol, os níveis de mRNA da lipase de triglicerídeos do adipócito (ATGL) e lipase hormônio sensível (HSL), além do conteúdo proteico da ATGL e phosphoSer660HSL. O aumento na lipólise induzido por 16:1n7, que pode ser abolido através da inibição farmacológica de PPAR $\alpha$ , foi associado a maior atividade transcrional de PPAR $\alpha$  *in vitro*. Ademais, 16:1n7 e 16:0 aumentaram a incorporação de ácidos graxos em TAG e a síntese de glicerol 3-fosfato a partir da glicose, sem modificar a gliceroneogênese e a expressão da gliceroquinase. Corroborando com os resultados *in vitro*, o tratamento de camundongos selvagens, mas não os deficientes em PPAR $\alpha$ , com 16:1n7 aumentou a lipólise basal e estimulada e os níveis de mRNA da ATGL e HSL em adipócitos epididimais. Em contraste com a lipólise *in vivo*, o tratamento com 16:1n7 aumentou a incorporação de ácidos graxos em TAG e a síntese de glicerol 3-fosfato a partir da glicose, tanto em camundongos selvagens quanto em deficientes para PPAR $\alpha$ . Em resumo, o ácido palmitoleico aumenta a captação de glicose e sua utilização pelos adipócitos brancos, um efeito que está associado com o aumento da expressão de GLUT4 e ativação da AMPK. Além disso, este ácido aumenta a lipólise e lipases em adipócitos por um mecanismo dependente de PPAR $\alpha$ .

**Palavras-chave:** Metabolismo de glicose. Lipogênese. Lipólise. Ácidos graxos.

## ABSTRACT

Bolsoni-Lopes A. Effects of palmitoleic acid on the uptake and metabolism of glucose and triacylglycerol in white adipocytes. [Ph. D. thesis (Human Physiology)]. São Paulo: Instituto de Ciências Biomédicas, Universidade de São Paulo; 2014.

Herein we investigated whether palmitoleic acid, a fatty acid that enhances whole body glucose disposal and suppresses hepatic steatosis, modulates glucose and triacylglycerol (TAG) metabolism in white adipocytes. For this, both differentiated 3T3-L1 cells treated with either palmitoleic acid (16:1n7, 200  $\mu$ M) or palmitic acid (16:0, 200  $\mu$ M) for 24h and primary epididimal adipocytes from wild type or PPAR $\alpha$  knockout mice treated with 16:1n7 (300 mg/kg/day) or oleic acid (18:1n9, 300 mg/kg/day) by gavage for 10 days were evaluated for glucose uptake and its oxidation, conversion into lactate and incorporation into fatty acids and glycerol of TAG, as well as, incorporation of acetate into fatty acids, lipolysis, TAG synthesis and gene and protein expression profile. Our results demonstrate that treatment with 16:1n7, but not 18:1n9 (*in vivo*) or 16:0 (*in vitro*) acids, increases both basal and in sulin-stimulated glucose uptake and the content of GLUT4 and phosphoThr172AMPK $\alpha$ , without affecting GLUT1 or phosphoSer473AKT. Such increase in GLUT4 induced by 16:1n7 was prevented by pharmacological inhibition of AMPK. In addition, 16:1n7 increases glucose conversion into lactate and CO<sub>2</sub> (glucose oxidation), and decreases *de novo* fatty acids synthesis *in vitro*. Corroborating these findings, the activity of glucose-6-phosphate dehydrogenase and ATP-citrate liase enzymes were also decreased by treatment of 3T3-L1 cells with 16:1n7. Furthermore, treatment of 3T3-L1 cells with 16:1n7, but not 16:0, increased basal and isoproterenol-stimulated lipolysis, mRNA levels of adipose triglyceride lipase (ATGL) and hormone sensitive lipase (HSL) and protein content of ATGL and phosphoSer660HSL. Such increase in lipolysis induced by 16:1n7, which can be prevented by pharmacological inhibition of PPAR $\alpha$ , was associated with higher rates of PPAR $\alpha$  binding to DNA *in vitro*. In addition, both 16:1n7 and 16:0 increased fatty acid incorporation into TAG and glycerol 3-phosphate synthesis from glucose, without affecting glyceroneogenesis and glycerokinase expression. Corroborating *in vitro* findings, treatment of wild type, but not PPAR $\alpha$  deficient mice with 16:1n7 increased epididimal adipocytes basal and stimulated lipolysis and ATGL and HSL mRNA levels. In contrast to lipolysis *in vivo*, however, 16:1n7 treatment increased fatty acid incorporation into TAG and glycerol 3-phosphate synthesis from glucose in both wild type and PPAR $\alpha$  deficient mice. In conclusion, palmitoleic acid increases glucose uptake and utilization by white adipocytes, an effect that was associated with increase of GLUT4 expression and AMPK activation. Furthermore, palmitoleic acid increases adipocyte lipolysis and lipases by a mechanism that requires a functional PPAR $\alpha$ .

**Keywords:** Glucose metabolism. Lipogenesis. Lipolysis. Fatty acids.

## 1 INTRODUÇÃO

O ácido palmitoleico ou ácido delta-9-cis-hexadecenóico é um ácido graxo monoinsaturado ômega 7 de dezesseis carbonos (16:1n7) sintetizado endogenamente em células como adipócitos e hepatócitos como resultado da dessaturação do ácido palmítico (16:0), processo catalisado pela enzima *estearoil-CoA-dessaturase 1* (SCD-1). Dessa forma, este ácido graxo é tido como um marcador da síntese *de novo* de ácidos graxos (1). Além da síntese endógena, o ácido palmitoleico também pode ser encontrado principalmente no óleo de semente de macadâmia (cerca de 30% do total de lipídeos) e na polpa de espinheiro-marítimo (*Hippophae rhamnoides*, entre 20 e 30%) (2).

Este ácido graxo é facilmente encontrado no plasma e tecido adiposo branco (TAB) de mamíferos e, apesar de praticamente ausente em pré-adipócitos, sua concentração aumenta acentuadamente durante a adipogênese, tornando-se o quarto ou quinto ácido graxo mais abundante que compõe os triacilgliceróis (TAG) de adipócitos humanos (variando conforme o depósito de TAB estudado) (3-5) e o mais abundante nos TAG de células 3T3-L1 (6, 7). Além disso, o ácido palmitoleico ocupa principalmente a posição *sn2* da molécula de TAG (8, 9).

Ácidos graxos são importantes moléculas sinalizadoras celular, capazes de modular o metabolismo das células e tecidos por mecanismos distintos, que favorecem ou previnem o desenvolvimento de doenças como o diabetes tipo II, aterosclerose e obesidade (10-13). Em sua estrutura, o tamanho da cadeia carbônica e o número/posição das duplas ligações determinam suas propriedades físicas e químicas, conferindo a cada um deles funções biológicas únicas (14). Neste contexto, o ácido palmitoleico foi recentemente descrito como um hormônio lipídico, denominado lipocina, que é sintetizado e secretado por adipócitos e atua em órgãos alvos distantes modulando importantes processos metabólicos e inflamatórios de tecidos corporais como o músculo, fígado e pâncreas (1, 15).

Estudos *in vitro* e *in vivo* (roedores e humanos) encontraram que o ácido palmitoleico possui ações importantes no músculo esquelético e fígado aumentando a captação de glicose e a sensibilidade à insulina, bem como reduzindo a esteatose hepática, respectivamente (1, 16-18). Mais especificamente, estudos verificaram que células musculares esqueléticas tratadas com ácido palmitoleico apresentam

aumento na captação de glicose, maior conteúdo proteico de transportadores de glicose 1 e 4 (GLUTs) na membrana celular e aumento na fosforilação da AKT, do receptor de insulina (IR) e dos substratos do receptor de insulina 1 - 2 (IRS), além de aumento na oxidação de glicose (1, 17, 19). Interessantemente, Dimopoulos et al. (2006) demonstraram que o tratamento com ácido palmitoleico bloqueou os efeitos deletérios do ácido palmítico no metabolismo da glicose e sensibilidade à insulina em células musculares L6 (17).

Já no tecido hepático, o tratamento com ácido palmitoleico também aumentou a fosforilação da AKT, IR e IRS 1, além de inibir a síntese *de novo* de ácidos graxos, esteatose hepática e a ativação do promotor e expressão gênica da SCD-1 (1). Esse último achado indica a existência de um sistema de retroalimentação negativa no qual o ácido palmitoleico inibe a expressão de SCD-1 e assim a sua formação a partir do ácido palmítico neste tecido.

Corroborando estes estudos, camundongos diabéticos tratados com ácido palmitoleico (300 mg/kg) por quatro semanas apresentaram uma atenuação da hiperglicemia, hipertrigliceridemia e melhora na sensibilidade à insulina. Além disso, houve redução na expressão gênica das enzimas lipogênicas *ácido graxo sintase* (FAS), SREBP-1 e SCD-1 no fígado e dos mediadores pró-inflamatórios - fator de necrose tumoral alfa (TNF- $\alpha$ ) e resistina no TAB (16). Efeitos similares foram também encontrados após tratamento de adipócitos bovinos com concentrações crescentes de ácido palmitoleico (50-300  $\mu$ M), como evidenciado pela redução na expressão gênica da FAS e SCD-1 e aumento na expressão da enzima *carnitina palmitoil-transferase 1A* (CPT1), que transporta ácidos graxos para a mitocôndria para serem oxidados (20). Além dos efeitos no metabolismo de glicose e lipídeos, o tratamento com 16:1n7 também produziu efeitos citoprotetores, protegendo células beta pancreáticas da morte induzida por ácido palmítico (15, 21).

Em estudos clínicos também se observam efeitos do ácido palmitoleico na homeostase metabólica de humanos. Hiraoka-Yamamoto et al. (2004) relataram que mulheres japonesas jovens com dieta rica em óleo de macadâmia têm o peso corporal, índice de massa corpórea e concentrações séricas de colesterol total e de LDL significativamente diminuídos (22). Interessantemente, o acúmulo de gordura glúteo-femural em humanos é inversamente associado a riscos cardiovasculares, efeito este, que é parcialmente justificado pelo grande acúmulo e liberação de ácido

palmitoleico por este tecido (5). Além disso, a concentração plasmática deste ácido graxo foi inversamente associada com o risco de câncer (23, 24). Contudo, no plasma de crianças e adultos obesos são detectadas concentrações elevadas de ácido palmitoleico e esses sujeitos também apresentam aumento da atividade da SCD-1 (25, 26).

Apesar de todos estes efeitos metabólicos importantes no fígado, músculo esquelético e células beta pancreáticas, ainda não foi demonstrado se o ácido palmitoleico também modula o metabolismo e funcionamento do TAB, um órgão fundamental para o controle da homeostase energética corporal. No presente estudo, portanto, nós testamos a hipótese de que o ácido palmitoleico é um importante modulador do metabolismo de glicose e triacilglicerol de adipócitos brancos. Para isto, utilizando experimentos *in vitro* e *in vivo* com células 3T3-L1 diferenciadas e adipócitos primários de murinos, nós investigamos, em dois estudos distintos, os efeitos e mecanismos de ação do ácido palmitoleico na captação e utilização de glicose (Estudo I) e no metabolismo do TAG (Estudo II) em adipócitos brancos.

## 7 CONCLUSÃO

Nossos dados sugerem fortemente que o ácido palmitoleico atua como uma importante molécula sinalizadora capaz de modular o metabolismo da glicose e do TAG em adipócitos brancos. Resumidamente, o tratamento de adipócitos com ácido palmitoleico, diferentemente do ácido palmítico ou oleico, induziu um aumento no conteúdo de GLUT4, na captação e utilização da glicose. Estes efeitos estão, ao menos em parte, associados com ativação da AMPK no tecido adiposo. O ácido palmitoleico também promoveu aumento da lipólise e do conteúdo das lipases ATGL e HSL através de um mecanismo dependente de PPAR $\alpha$ . O aumento na lipólise está associado a um aumento concomitante da reesterificação de ácidos graxos a TAG, sugerindo um aumento no ciclo TAG-ácido graxo. Assim, podemos concluir que este ácido parece acelerar o metabolismo celular de adipócitos promovendo a ativação das vias de produção de energia.

## REFERÊNCIAS\*

1. Cao H, Gerhold K, Mayers JR, Wiest MM, Watkins SM, Hotamisligil GS. Identification of a lipokine, a lipid hormone linking adipose tissue to systemic metabolism. *Cell*. 2008;134(6):933-44.
2. Kallio H, Yang B, Peippo P, Tahvonon R, Pan R. Triacylglycerols, glycerophospholipids, tocopherols, and tocotrienols in berries and seeds of two subspecies (ssp. *sinensis* and *mongolica*) of Sea Buckthorn (*Hippophaë rhamnoides*). *J Agric Food Chem*. 2002;50(10):3004-9.
3. Collins JM, Neville MJ, Pinnick KE, Hodson L, Ruyter B, van Dijk TH, et al. De novo lipogenesis in the differentiating human adipocyte can provide all fatty acids necessary for maturation. *J Lipid Res*. 2011;52(9):1683-92.
4. Caron-Jobin M, Mauvoisin D, Michaud A, Veilleux A, Noël S, Fortier MP, et al. Stearic acid content of abdominal adipose tissues in obese women. *Nutr Diabetes*. 2012;2:e23.
5. Pinnick KE, Neville MJ, Fielding BA, Frayn KN, Karpe F, Hodson L. Gluteofemoral adipose tissue plays a major role in production of the lipokine palmitoleate in humans. *Diabetes*. 2012;61(6):1399-403.
6. Roberts LD, Virtue S, Vidal-Puig A, Nicholls AW, Griffin JL. Metabolic phenotyping of a model of adipocyte differentiation. *Physiol Genomics*. 2009;39(2):109-19.
7. Bolsoni-Lopes A, Festuccia WT, Farias TS, Chimin P, Torres-Leal FL, Derogis PB, et al. Palmitoleic acid (n-7) increases white adipocyte lipolysis and lipase content in a PPAR $\alpha$ -dependent manner. *Am J Physiol Endocrinol Metab*. 2013;305(9):E1093-102.
8. Eichmann TO, Kumari M, Haas JT, Farese RV, Zimmermann R, Lass A, et al. Studies on the substrate and stereo/regioselectivity of adipose triglyceride lipase, hormone-sensitive lipase, and diacylglycerol-O-acyltransferases. *J Biol Chem*. 2012;287(49):41446-57.
9. Weber N, Bergenthal D, Kokate CK, Mangold HK. Biologically active ether lipids: incorporation of long-chain precursors into 1(3),2-diacylglycero-3(1)-O-4'-(N,N,N-trimethyl)homoserines and other lipids of *Chlorella fusca*. *J Lipid Mediat*. 1989;1(1):37-48.
10. Tvrzicka E, Kremmyda LS, Stankova B, Zak A. Fatty acids as biocompounds: their role in human metabolism, health and disease--a review. Part 1: classification, dietary sources and biological functions. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub*. 2011;155(2):117-30.

\*De acordo com:

International Committee of Medical Journal Editors. Uniform requirements for manuscripts submitted to Biomedical Journal: sample references. Available from: <http://www.icmje.org> [2011 Jul 15].



11. Queiroz JC, Alonso-Vale MI, Curi R, Lima FB. [Control of adipogenesis by fatty acids]. *Arq Bras Endocrinol Metabol.* 2009;53(5):582-94.
12. Parillo M, Riccardi G. Diet composition and the risk of type 2 diabetes: epidemiological and clinical evidence. *Br J Nutr.* 2004;92(1):7-19.
13. Hirabara SM, Curi R, Maechler P. Saturated fatty acid-induced insulin resistance is associated with mitochondrial dysfunction in skeletal muscle cells. *J Cell Physiol.* 2010;222(1):187-94.
14. Leonard AE, Pereira SL, Sprecher H, Huang YS. Elongation of long-chain fatty acids. *Prog Lipid Res.* 2004;43(1):36-54.
15. Diakogiannaki E, Dhayal S, Childs CE, Calder PC, Welters HJ, Morgan NG. Mechanisms involved in the cytotoxic and cytoprotective actions of saturated versus monounsaturated long-chain fatty acids in pancreatic beta-cells. *J Endocrinol.* 2007;194(2):283-91.
16. Yang ZH, Miyahara H, Hatanaka A. Chronic administration of palmitoleic acid reduces insulin resistance and hepatic lipid accumulation in KK-Ay Mice with genetic type 2 diabetes. *Lipids Health Dis.* 2011;10:120.
17. Dimopoulos N, Watson M, Sakamoto K, Hundal HS. Differential effects of palmitate and palmitoleate on insulin action and glucose utilization in rat L6 skeletal muscle cells. *Biochem J.* 2006;399(3):473-81.
18. Stefan N, Kantartzis K, Celebi N, Staiger H, Machann J, Schick F, et al. Circulating palmitoleate strongly and independently predicts insulin sensitivity in humans. *Diabetes Care.* 2010;33(2):405-7.
19. Obanda DN, Cefalu WT. Modulation of cellular insulin signaling and PTP1B effects by lipid metabolites in skeletal muscle cells. *J Nutr Biochem.* 2013;24(8):1529-37.
20. Burns TA, Kadegowda AK, Duckett SK, Pratt SL, Jenkins TC. Palmitoleic (16:1 cis-9) and cis-vaccenic (18:1 cis-11) acid alter lipogenesis in bovine adipocyte cultures. *Lipids.* 2012;47(12):1143-53.
21. Morgan NG, Dhayal S. Unsaturated fatty acids as cytoprotective agents in the pancreatic beta-cell. *Prostaglandins Leukot Essent Fatty Acids.* 2010;82(4-6):231-6.
22. Hiraoka-Yamamoto J, Ikeda K, Negishi H, Mori M, Hirose A, Sawada S, et al. Serum lipid effects of a monounsaturated (palmitoleic) fatty acid-rich diet based on macadamia nuts in healthy, young Japanese women. *Clin Exp Pharmacol Physiol.* 2004;31 Suppl 2:S37-8.
23. Pouchieu C, Chajès V, Laporte F, Kesse-Guyot E, Galan P, Hercberg S, et al. Prospective associations between plasma saturated, monounsaturated and polyunsaturated fatty acids and overall and breast cancer risk - modulation by antioxidants: a nested case-control study. *PLoS One.* 2014;9(2):e90442.

24. Simonsen NR, Fernandez-Crehuet Navajas J, Martin-Moreno JM, Strain JJ, Huttunen JK, Martin BC, et al. Tissue stores of individual monounsaturated fatty acids and breast cancer: the EURAMIC study. *European Community Multicenter Study on Antioxidants, Myocardial Infarction, and Breast Cancer. Am J Clin Nutr.* 1998;68(1):134-41.
25. Okada T, Furuhashi N, Kuromori Y, Miyashita M, Iwata F, Harada K. Plasma palmitoleic acid content and obesity in children. *Am J Clin Nutr.* 2005;82(4):747-50.
26. Kunesová M, Hainer V, Tvrzicka E, Phinney SD, Stich V, Parízková J, et al. Assessment of dietary and genetic factors influencing serum and adipose fatty acid composition in obese female identical twins. *Lipids.* 2002;37(1):27-32.
27. Hausman GJ, Barb CR. Adipose tissue and the reproductive axis: biological aspects. *Endocr Dev.* 2010;19:31-44.
28. Otto TC, Lane MD. Adipose development: from stem cell to adipocyte. *Crit Rev Biochem Mol Biol.* 2005;40(4):229-42.
29. Langin D, Arner P. Importance of TNFalpha and neutral lipases in human adipose tissue lipolysis. *Trends Endocrinol Metab.* 2006;17(8):314-20.
30. Fonseca-Alaniz MH, Takada J, Alonso-Vale MI, Lima FB. [The adipose tissue as a regulatory center of the metabolism]. *Arq Bras Endocrinol Metabol.* 2006;50(2):216-29.
31. Grousse A, Langin D. Adipocyte lipases and lipid droplet-associated proteins: insight from transgenic mouse models. *Int J Obes (Lond).* 2012;36(4):581-94.
32. Brasaemle DL. Thematic review series: adipocyte biology. The perilipin family of structural lipid droplet proteins: stabilization of lipid droplets and control of lipolysis. *J Lipid Res.* 2007;48(12):2547-59.
33. Large V, Peroni O, Letexier D, Ray H, Beylot M. Metabolism of lipids in human white adipocyte. *Diabetes Metab.* 2004;30(4):294-309.
34. Wajchenberg BL. Subcutaneous and visceral adipose tissue: their relation to the metabolic syndrome. *Endocr Rev.* 2000;21(6):697-738.
35. Tchernof A, Després JP. Pathophysiology of human visceral obesity: an update. *Physiol Rev.* 2013;93(1):359-404.
36. Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional cloning of the mouse obese gene and its human homologue. *Nature.* 1994;372(6505):425-32.
37. Cannon B, Nedergaard J. Brown adipose tissue: function and physiological significance. *Physiol Rev.* 2004;84(1):277-359.

38. Festuccia WT, Blanchard PG, Deshaies Y. Control of Brown Adipose Tissue Glucose and Lipid Metabolism by PPAR $\gamma$ . *Front Endocrinol (Lausanne)*. 2011;2:84.
39. Nedergaard J, Bengtsson T, Cannon B. Unexpected evidence for active brown adipose tissue in adult humans. *Am J Physiol Endocrinol Metab*. 2007;293(2):E444-52.
40. Cypess AM, Kahn CR. Brown fat as a therapy for obesity and diabetes. *Curr Opin Endocrinol Diabetes Obes*. 2010;17(2):143-9.
41. Lafontan M, Langin D. Lipolysis and lipid mobilization in human adipose tissue. *Prog Lipid Res*. 2009;48(5):275-97.
42. Lafontan M. Advances in adipose tissue metabolism. *Int J Obes (Lond)*. 2008;32 Suppl 7:S39-51.
43. WEISS SB, KENNEDY EP, KIYASU JY. The enzymatic synthesis of triglycerides. *J Biol Chem*. 1960;235:40-4.
44. Dircks L, Sul HS. Acyltransferases of de novo glycerophospholipid biosynthesis. *Prog Lipid Res*. 1999;38(5-6):461-79.
45. Lehner R, Kuksis A. Biosynthesis of triacylglycerols. *Prog Lipid Res*. 1996;35(2):169-201.
46. Agarwal AK, Garg A. Congenital generalized lipodystrophy: significance of triglyceride biosynthetic pathways. *Trends Endocrinol Metab*. 2003;14(5):214-21.
47. West DB, York B. Dietary fat, genetic predisposition, and obesity: lessons from animal models. *Am J Clin Nutr*. 1998;67(3 Suppl):505S-12S.
48. Carman GM, Han GS. Roles of phosphatidate phosphatase enzymes in lipid metabolism. *Trends Biochem Sci*. 2006;31(12):694-9.
49. Ahmadian M, Duncan RE, Jaworski K, Sarkadi-Nagy E, Sul HS. Triacylglycerol metabolism in adipose tissue. *Future Lipidol*. 2007;2(2):229-37.
50. Reshef L, Hanson RW, Ballard FJ. A possible physiological role for glyceroneogenesis in rat adipose tissue. *J Biol Chem*. 1970;245(22):5979-84.
51. Mead JR, Irvine SA, Ramji DP. Lipoprotein lipase: structure, function, regulation, and role in disease. *J Mol Med (Berl)*. 2002;80(12):753-69.
52. Abumrad NA, el-Maghrabi MR, Amri EZ, Lopez E, Grimaldi PA. Cloning of a rat adipocyte membrane protein implicated in binding or transport of long-chain fatty acids that is induced during preadipocyte differentiation. Homology with human CD36. *J Biol Chem*. 1993;268(24):17665-8.
53. Schaffer JE, Lodish HF. Expression cloning and characterization of a novel adipocyte long chain fatty acid transport protein. *Cell*. 1994;79(3):427-36.

54. Coe NR, Simpson MA, Bernlohr DA. Targeted disruption of the adipocyte lipid-binding protein (aP2 protein) gene impairs fat cell lipolysis and increases cellular fatty acid levels. *J Lipid Res.* 1999;40(5):967-72.
55. Strawford A, Antelo F, Christiansen M, Hellerstein MK. Adipose tissue triglyceride turnover, de novo lipogenesis, and cell proliferation in humans measured with 2H<sub>2</sub>O. *Am J Physiol Endocrinol Metab.* 2004;286(4):E577-88.
56. Kersten S. Mechanisms of nutritional and hormonal regulation of lipogenesis. *EMBO Rep.* 2001;2(4):282-6.
57. Govers R. Molecular mechanisms of GLUT4 regulation in adipocytes. *Diabetes Metab.* 2014.
58. DiGirolamo M, Newby FD, Lovejoy J. Lactate production in adipose tissue: a regulated function with extra-adipose implications. *FASEB J.* 1992;6(7):2405-12.
59. Festuccia WT, Blanchard PG, Turcotte V, Laplante M, Sariahmetoglu M, Brindley DN, et al. Depot-specific effects of the PPARgamma agonist rosiglitazone on adipose tissue glucose uptake and metabolism. *J Lipid Res.* 2009;50(6):1185-94.
60. Wood IS, Trayhurn P. Glucose transporters (GLUT and SGLT): expanded families of sugar transport proteins. *Br J Nutr.* 2003;89(1):3-9.
61. Shepherd PR, Kahn BB. Glucose transporters and insulin action--implications for insulin resistance and diabetes mellitus. *N Engl J Med.* 1999;341(4):248-57.
62. Bryant NJ, Govers R, James DE. Regulated transport of the glucose transporter GLUT4. *Nat Rev Mol Cell Biol.* 2002;3(4):267-77.
63. Capurso C, Capurso A. From excess adiposity to insulin resistance: the role of free fatty acids. *Vascul Pharmacol.* 2012;57(2-4):91-7.
64. Sano H, Kane S, Sano E, Miinea CP, Asara JM, Lane WS, et al. Insulin-stimulated phosphorylation of a Rab GTPase-activating protein regulates GLUT4 translocation. *J Biol Chem.* 2003;278(17):14599-602.
65. Wang Q, Somwar R, Bilan PJ, Liu Z, Jin J, Woodgett JR, et al. Protein kinase B/Akt participates in GLUT4 translocation by insulin in L6 myoblasts. *Mol Cell Biol.* 1999;19(6):4008-18.
66. Ceddia RB. The role of AMP-activated protein kinase in regulating white adipose tissue metabolism. *Mol Cell Endocrinol.* 2013;366(2):194-203.
67. Gaidhu MP, Perry RL, Noor F, Ceddia RB. Disruption of AMPKalpha1 signaling prevents AICAR-induced inhibition of AS160/TBC1D4 phosphorylation and glucose uptake in primary rat adipocytes. *Mol Endocrinol.* 2010;24(7):1434-40.
68. Richter EA, Hargreaves M. Exercise, GLUT4, and skeletal muscle glucose uptake. *Physiol Rev.* 2013;93(3):993-1017.

69. Huang YC, Chang WL, Huang SF, Lin CY, Lin HC, Chang TC. Pachymic acid stimulates glucose uptake through enhanced GLUT4 expression and translocation. *Eur J Pharmacol.* 2010;648(1-3):39-49.
70. Duncan RE, Ahmadian M, Jaworski K, Sarkadi-Nagy E, Sul HS. Regulation of lipolysis in adipocytes. *Annu Rev Nutr.* 2007;27:79-101.
71. Bézaire V, Langin D. Regulation of adipose tissue lipolysis revisited. *Proc Nutr Soc.* 2009;68(4):350-60.
72. Schweiger M, Schreiber R, Haemmerle G, Lass A, Fledelius C, Jacobsen P, et al. Adipose triglyceride lipase and hormone-sensitive lipase are the major enzymes in adipose tissue triacylglycerol catabolism. *J Biol Chem.* 2006;281(52):40236-41.
73. Ahmadian M, Wang Y, Sul HS. Lipolysis in adipocytes. *Int J Biochem Cell Biol.* 2010;42(5):555-9.
74. Yamaguchi T, Omatsu N, Matsushita S, Osumi T. CGI-58 interacts with perilipin and is localized to lipid droplets. Possible involvement of CGI-58 mislocalization in Chanarin-Dorfman syndrome. *J Biol Chem.* 2004;279(29):30490-7.
75. Miyoshi H, Souza SC, Endo M, Sawada T, Perfield JW, Shimizu C, et al. Perilipin overexpression in mice protects against diet-induced obesity. *J Lipid Res.* 2010;51(5):975-82.
76. Ahmadian M, Duncan RE, Sul HS. The skinny on fat: lipolysis and fatty acid utilization in adipocytes. *Trends Endocrinol Metab.* 2009;20(9):424-8.
77. Zechner R, Zimmermann R, Eichmann TO, Kohlwein SD, Haemmerle G, Lass A, et al. FAT SIGNALS--lipases and lipolysis in lipid metabolism and signaling. *Cell Metab.* 2012;15(3):279-91.
78. Jaworski K, Sarkadi-Nagy E, Duncan RE, Ahmadian M, Sul HS. Regulation of triglyceride metabolism. IV. Hormonal regulation of lipolysis in adipose tissue. *Am J Physiol Gastrointest Liver Physiol.* 2007;293(1):G1-4.
79. Wang S, Soni KG, Semache M, Casavant S, Fortier M, Pan L, et al. Lipolysis and the integrated physiology of lipid energy metabolism. *Mol Genet Metab.* 2008;95(3):117-26.
80. Jaworski K, Ahmadian M, Duncan RE, Sarkadi-Nagy E, Varady KA, Hellerstein MK, et al. AdPLA ablation increases lipolysis and prevents obesity induced by high-fat feeding or leptin deficiency. *Nat Med.* 2009;15(2):159-68.
81. Duncan RE, Sarkadi-Nagy E, Jaworski K, Ahmadian M, Sul HS. Identification and functional characterization of adipose-specific phospholipase A2 (AdPLA). *J Biol Chem.* 2008;283(37):25428-36.

82. Ong KT, Mashek MT, Bu SY, Greenberg AS, Mashek DG. Adipose triglyceride lipase is a major hepatic lipase that regulates triacylglycerol turnover and fatty acid signaling and partitioning. *Hepatology*. 2011;53(1):116-26.
83. Festuccia WT, Laplante M, Berthiaume M, G elinas Y, Deshaies Y. PPARgamma agonism increases rat adipose tissue lipolysis, expression of glyceride lipases, and the response of lipolysis to hormonal control. *Diabetologia*. 2006;49(10):2427-36.
84. Zimmermann R, Strauss JG, Haemmerle G, Schoiswohl G, Birner-Gruenberger R, Riederer M, et al. Fat mobilization in adipose tissue is promoted by adipose triglyceride lipase. *Science*. 2004;306(5700):1383-6.
85. Jenkins CM, Mancuso DJ, Yan W, Sims HF, Gibson B, Gross RW. Identification, cloning, expression, and purification of three novel human calcium-independent phospholipase A2 family members possessing triacylglycerol lipase and acylglycerol transacylase activities. *J Biol Chem*. 2004;279(47):48968-75.
86. Villena JA, Roy S, Sarkadi-Nagy E, Kim KH, Sul HS. Desnutrin, an adipocyte gene encoding a novel patatin domain-containing protein, is induced by fasting and glucocorticoids: ectopic expression of desnutrin increases triglyceride hydrolysis. *J Biol Chem*. 2004;279(45):47066-75.
87. Haemmerle G, Lass A, Zimmermann R, Gorkiewicz G, Meyer C, Rozman J, et al. Defective lipolysis and altered energy metabolism in mice lacking adipose triglyceride lipase. *Science*. 2006;312(5774):734-7.
88. Schoiswohl G, Schweiger M, Schreiber R, Gorkiewicz G, Preiss-Landl K, Taschler U, et al. Adipose triglyceride lipase plays a key role in the supply of the working muscle with fatty acids. *J Lipid Res*. 2010;51(3):490-9.
89. Peyot ML, Guay C, Latour MG, Lamontagne J, Lussier R, Pineda M, et al. Adipose triglyceride lipase is implicated in fuel- and non-fuel-stimulated insulin secretion. *J Biol Chem*. 2009;284(25):16848-59.
90. Tang T, Abbott MJ, Ahmadian M, Lopes AB, Wang Y, Sul HS. Desnutrin/ATGL activates PPAR  to promote mitochondrial function for insulin secretion in islet   cells. *Cell Metab*. 2013;18(6):883-95.
91. Kershaw EE, Hamm JK, Verhagen LA, Peroni O, Katic M, Flier JS. Adipose triglyceride lipase: function, regulation by insulin, and comparison with adiponutrin. *Diabetes*. 2006;55(1):148-57.
92. Smirnova E, Goldberg EB, Makarova KS, Lin L, Brown WJ, Jackson CL. ATGL has a key role in lipid droplet/adiposome degradation in mammalian cells. *EMBO Rep*. 2006;7(1):106-13.
93. Lass A, Zimmermann R, Oberer M, Zechner R. Lipolysis - a highly regulated multi-enzyme complex mediates the catabolism of cellular fat stores. *Prog Lipid Res*. 2011;50(1):14-27.

94. Chakrabarti P, English T, Shi J, Smas CM, Kandror KV. Mammalian target of rapamycin complex 1 suppresses lipolysis, stimulates lipogenesis, and promotes fat storage. *Diabetes*. 2010;59(4):775-81.
95. Lu X, Yang X, Liu J. Differential control of ATGL-mediated lipid droplet degradation by CGI-58 and G0S2. *Cell Cycle*. 2010;9(14):2719-25.
96. Granneman JG, Moore HP, Mottillo EP, Zhu Z, Zhou L. Interactions of perilipin-5 (Plin5) with adipose triglyceride lipase. *J Biol Chem*. 2011;286(7):5126-35.
97. Grahn TH, Kaur R, Yin J, Schweiger M, Sharma VM, Lee MJ, et al. Fat-specific Protein 27 (FSP27) Interacts with Adipose Triglyceride Lipase (ATGL) to Regulate Lipolysis and Insulin Sensitivity in Human Adipocytes. *J Biol Chem*. 2014;289(17):12029-39.
98. Ellong EN, Soni KG, Bui QT, Sougrat R, Golinelli-Cohen MP, Jackson CL. Interaction between the triglyceride lipase ATGL and the Arf1 activator GBF1. *PLoS One*. 2011;6(7):e21889.
99. VAUGHAN M. The production and release of glycerol by adipose tissue incubated in vitro. *J Biol Chem*. 1962;237:3354-8.
100. Osuga J, Ishibashi S, Oka T, Yagyu H, Tozawa R, Fujimoto A, et al. Targeted disruption of hormone-sensitive lipase results in male sterility and adipocyte hypertrophy, but not in obesity. *Proc Natl Acad Sci U S A*. 2000;97(2):787-92.
101. Haemmerle G, Zimmermann R, Hayn M, Theussl C, Waeg G, Wagner E, et al. Hormone-sensitive lipase deficiency in mice causes diglyceride accumulation in adipose tissue, muscle, and testis. *J Biol Chem*. 2002;277(7):4806-15.
102. Wang SP, Laurin N, Himms-Hagen J, Rudnicki MA, Levy E, Robert MF, et al. The adipose tissue phenotype of hormone-sensitive lipase deficiency in mice. *Obes Res*. 2001;9(2):119-28.
103. Lucas S, Tavernier G, Tiraby C, Mairal A, Langin D. Expression of human hormone-sensitive lipase in white adipose tissue of transgenic mice increases lipase activity but does not enhance in vitro lipolysis. *J Lipid Res*. 2003;44(1):154-63.
104. Anthonsen MW, Rönstrand L, Wernstedt C, Degerman E, Holm C. Identification of novel phosphorylation sites in hormone-sensitive lipase that are phosphorylated in response to isoproterenol and govern activation properties in vitro. *J Biol Chem*. 1998;273(1):215-21.
105. Wang H, Hu L, Dalen K, Dorward H, Marcinkiewicz A, Russell D, et al. Activation of hormone-sensitive lipase requires two steps, protein phosphorylation and binding to the PAT-1 domain of lipid droplet coat proteins. *J Biol Chem*. 2009;284(46):32116-25.

106. Shen WJ, Sridhar K, Bernlohr DA, Kraemer FB. Interaction of rat hormone-sensitive lipase with adipocyte lipid-binding protein. *Proc Natl Acad Sci U S A*. 1999;96(10):5528-32.
107. Smith AJ, Thompson BR, Sanders MA, Bernlohr DA. Interaction of the adipocyte fatty acid-binding protein with the hormone-sensitive lipase: regulation by fatty acids and phosphorylation. *J Biol Chem*. 2007;282(44):32424-32.
108. Karlsson M, Contreras JA, Hellman U, Tornqvist H, Holm C. cDNA cloning, tissue distribution, and identification of the catalytic triad of monoglyceride lipase. Evolutionary relationship to esterases, lysophospholipases, and haloperoxidases. *J Biol Chem*. 1997;272(43):27218-23.
109. Taschler U, Radner FP, Heier C, Schreiber R, Schweiger M, Schoiswohl G, et al. Monoglyceride lipase deficiency in mice impairs lipolysis and attenuates diet-induced insulin resistance. *J Biol Chem*. 2011;286(20):17467-77.
110. Bijland S, Mancini SJ, Salt IP. Role of AMP-activated protein kinase in adipose tissue metabolism and inflammation. *Clin Sci (Lond)*. 2013;124(8):491-507.
111. Xiao B, Heath R, Saiu P, Leiper FC, Leone P, Jing C, et al. Structural basis for AMP binding to mammalian AMP-activated protein kinase. *Nature*. 2007;449(7161):496-500.
112. Minokoshi Y, Kim YB, Peroni OD, Fryer LG, Müller C, Carling D, et al. Leptin stimulates fatty-acid oxidation by activating AMP-activated protein kinase. *Nature*. 2002;415(6869):339-43.
113. Orci L, Cook WS, Ravazzola M, Wang MY, Park BH, Montesano R, et al. Rapid transformation of white adipocytes into fat-oxidizing machines. *Proc Natl Acad Sci U S A*. 2004;101(7):2058-63.
114. Ahmadian M, Abbott MJ, Tang T, Hudak CS, Kim Y, Bruss M, et al. Desnutrin/ATGL is regulated by AMPK and is required for a brown adipose phenotype. *Cell Metab*. 2011;13(6):739-48.
115. Fryer LG, Parbu-Patel A, Carling D. The Anti-diabetic drugs rosiglitazone and metformin stimulate AMP-activated protein kinase through distinct signaling pathways. *J Biol Chem*. 2002;277(28):25226-32.
116. Wu X, Motoshima H, Mahadev K, Stalker TJ, Scalia R, Goldstein BJ. Involvement of AMP-activated protein kinase in glucose uptake stimulated by the globular domain of adiponectin in primary rat adipocytes. *Diabetes*. 2003;52(6):1355-63.
117. Gaidhu MP, Fediuc S, Anthony NM, So M, Mirpourian M, Perry RL, et al. Prolonged AICAR-induced AMP-kinase activation promotes energy dissipation in white adipocytes: novel mechanisms integrating HSL and ATGL. *J Lipid Res*. 2009;50(4):704-15.



118. Kramer HF, Witczak CA, Fujii N, Jessen N, Taylor EB, Arnolds DE, et al. Distinct signals regulate AS160 phosphorylation in response to insulin, AICAR, and contraction in mouse skeletal muscle. *Diabetes*. 2006;55(7):2067-76.
119. Poulsen L, Siersbæk M, Mandrup S. PPARs: fatty acid sensors controlling metabolism. *Semin Cell Dev Biol*. 2012;23(6):631-9.
120. Schupp M, Lazar MA. Endogenous ligands for nuclear receptors: digging deeper. *J Biol Chem*. 2010;285(52):40409-15.
121. Issemann I, Green S. Activation of a member of the steroid hormone receptor superfamily by peroxisome proliferators. *Nature*. 1990;347(6294):645-50.
122. Ijpenberg A, Jeannin E, Wahli W, Desvergne B. Polarity and specific sequence requirements of peroxisome proliferator-activated receptor (PPAR)/retinoid X receptor heterodimer binding to DNA. A functional analysis of the malic enzyme gene PPAR response element. *J Biol Chem*. 1997;272(32):20108-17.
123. Chandra V, Huang P, Hamuro Y, Raghuram S, Wang Y, Burris TP, et al. Structure of the intact PPAR-gamma-RXR- nuclear receptor complex on DNA. *Nature*. 2008;456(7220):350-6.
124. Escher P, Braissant O, Basu-Modak S, Michalik L, Wahli W, Desvergne B. Rat PPARs: quantitative analysis in adult rat tissues and regulation in fasting and refeeding. *Endocrinology*. 2001;142(10):4195-202.
125. Mueller E, Drori S, Aiyer A, Yie J, Sarraf P, Chen H, et al. Genetic analysis of adipogenesis through peroxisome proliferator-activated receptor gamma isoforms. *J Biol Chem*. 2002;277(44):41925-30.
126. Tontonoz P, Hu E, Spiegelman BM. Stimulation of adipogenesis in fibroblasts by PPAR gamma 2, a lipid-activated transcription factor. *Cell*. 1994;79(7):1147-56.
127. Ribon V, Johnson JH, Camp HS, Saltiel AR. Thiazolidinediones and insulin resistance: peroxisome proliferator-activated receptor gamma activation stimulates expression of the CAP gene. *Proc Natl Acad Sci U S A*. 1998;95(25):14751-6.
128. Ribet C, Montastier E, Valle C, Bezaire V, Mazzucotelli A, Mairal A, et al. Peroxisome proliferator-activated receptor-alpha control of lipid and glucose metabolism in human white adipocytes. *Endocrinology*. 2010;151(1):123-33.
129. Guzmán M, Lo Verme J, Fu J, Oveisi F, Blázquez C, Piomelli D. Oleoylethanolamide stimulates lipolysis by activating the nuclear receptor peroxisome proliferator-activated receptor alpha (PPAR-alpha). *J Biol Chem*. 2004;279(27):27849-54.
130. Li P, Zhu Z, Lu Y, Granneman JG. Metabolic and cellular plasticity in white adipose tissue II: role of peroxisome proliferator-activated receptor-alpha. *Am J Physiol Endocrinol Metab*. 2005;289(4):E617-26.

131. Hiuge A, Tenenbaum A, Maeda N, Benderly M, Kumada M, Fisman EZ, et al. Effects of peroxisome proliferator-activated receptor ligands, bezafibrate and fenofibrate, on adiponectin level. *Arterioscler Thromb Vasc Biol.* 2007;27(3):635-41.
132. Haemmerle G, Moustafa T, Woelkart G, Büttner S, Schmidt A, van de Weijer T, et al. ATGL-mediated fat catabolism regulates cardiac mitochondrial function via PPAR- $\alpha$  and PGC-1. *Nat Med.* 2011;17(9):1076-85.
133. Chandak PG, Radovic B, Aflaki E, Kolb D, Buchebner M, Fröhlich E, et al. Efficient phagocytosis requires triacylglycerol hydrolysis by adipose triglyceride lipase. *J Biol Chem.* 2010;285(26):20192-201.
134. Cnop M, Hannaert JC, Hoorens A, Eizirik DL, Pipeleers DG. Inverse relationship between cytotoxicity of free fatty acids in pancreatic islet cells and cellular triglyceride accumulation. *Diabetes.* 2001;50(8):1771-7.
135. Anderson SG, Sanders TA, Cruickshank JK. Plasma fatty acid composition as a predictor of arterial stiffness and mortality. *Hypertension.* 2009;53(5):839-45.
136. Kennedy AD, DeLeo FR. Neutrophil apoptosis and the resolution of infection. *Immunol Res.* 2009;43(1-3):25-61.
137. Reynoso R, Salgado LM, Calderón V. High levels of palmitic acid lead to insulin resistance due to changes in the level of phosphorylation of the insulin receptor and insulin receptor substrate-1. *Mol Cell Biochem.* 2003;246(1-2):155-62.
138. Chavez JA, Summers SA. Characterizing the effects of saturated fatty acids on insulin signaling and ceramide and diacylglycerol accumulation in 3T3-L1 adipocytes and C2C12 myotubes. *Arch Biochem Biophys.* 2003;419(2):101-9.
139. Siriwardhana N, Kalupahana NS, Cekanova M, LeMieux M, Greer B, Moustaid-Moussa N. Modulation of adipose tissue inflammation by bioactive food compounds. *J Nutr Biochem.* 2013;24(4):613-23.
140. Murumalla RK, Gunasekaran MK, Padhan JK, Bencharif K, Gence L, Festy F, et al. Fatty acids do not pay the toll: effect of SFA and PUFA on human adipose tissue and mature adipocytes inflammation. *Lipids Health Dis.* 2012;11:175.
141. Regassa A, Kim WK. Effects of oleic acid and chicken serum on the expression of adipogenic transcription factors and adipogenic differentiation in hen preadipocytes. *Cell Biol Int.* 2013;37(9):961-71.
142. Ricchi M, Odoardi MR, Carulli L, Anzivino C, Ballestri S, Pinetti A, et al. Differential effect of oleic and palmitic acid on lipid accumulation and apoptosis in cultured hepatocytes. *J Gastroenterol Hepatol.* 2009;24(5):830-40.
143. Moldes M, Zuo Y, Morrison RF, Silva D, Park BH, Liu J, et al. Peroxisome-proliferator-activated receptor gamma suppresses Wnt/beta-catenin signalling during adipogenesis. *Biochem J.* 2003;376(Pt 3):607-13.

144. Nicoletti I, Migliorati G, Pagliacci MC, Grignani F, Riccardi C. A rapid and simple method for measuring thymocyte apoptosis by propidium iodide staining and flow cytometry. *J Immunol Methods*. 1991;139(2):271-9.
145. Masood A, Stark KD, Salem N. A simplified and efficient method for the analysis of fatty acid methyl esters suitable for large clinical studies. *J Lipid Res*. 2005;46(10):2299-305.
146. RODBELL M. METABOLISM OF ISOLATED FAT CELLS. I. EFFECTS OF HORMONES ON GLUCOSE METABOLISM AND LIPOLYSIS. *J Biol Chem*. 1964;239:375-80.
147. Fine JB, DiGirolamo M. A simple method to predict cellular density in adipocyte metabolic incubations. *Int J Obes Relat Metab Disord*. 1997;21(9):764-8.
148. Amengual J, Petrov P, Bonet ML, Ribot J, Palou A. Induction of carnitine palmitoyl transferase 1 and fatty acid oxidation by retinoic acid in HepG2 cells. *Int J Biochem Cell Biol*. 2012;44(11):2019-27.
149. Mercader J, Madsen L, Felipe F, Palou A, Kristiansen K, Bonet ML. All-trans retinoic acid increases oxidative metabolism in mature adipocytes. *Cell Physiol Biochem*. 2007;20(6):1061-72.
150. Alonso-Vale MI, Andreotti S, Mukai PY, Borges-Silva C, Peres SB, Cipolla-Neto J, et al. Melatonin and the circadian entrainment of metabolic and hormonal activities in primary isolated adipocytes. *J Pineal Res*. 2008;45(4):422-9.
151. Bergmeyer H, Bernt E, Schmidt F, Stork H. In *Methods of Enzymatic Analysis*. Orlando, FL:: Academic Press; 1974.
152. Bazin R, Ferré P. Assays of lipogenic enzymes. *Methods Mol Biol*. 2001;155:121-7.
153. Papa PC, Seraphim PM, Machado UF. Loss of weight restores GLUT 4 content in insulin-sensitive tissues of monosodium glutamate-treated obese mice. *Int J Obes Relat Metab Disord*. 1997;21(11):1065-70.
154. Catalá A. Five Decades with Polyunsaturated Fatty Acids: Chemical Synthesis, Enzymatic Formation, Lipid Peroxidation and Its Biological Effects. *J Lipids*. 2013;2013:710290.
155. Harris WS, Dayspring TD, Moran TJ. Omega-3 fatty acids and cardiovascular disease: new developments and applications. *Postgrad Med*. 2013;125(6):100-13.
156. Thorens B, Charron MJ, Lodish HF. Molecular physiology of glucose transporters. *Diabetes Care*. 1990;13(3):209-18.
157. Hernandez R, Teruel T, Lorenzo M. Akt mediates insulin induction of glucose uptake and up-regulation of GLUT4 gene expression in brown adipocytes. *FEBS Lett*. 2001;494(3):225-31.

158. Jäger S, Handschin C, St-Pierre J, Spiegelman BM. AMP-activated protein kinase (AMPK) action in skeletal muscle via direct phosphorylation of PGC-1 $\alpha$ . *Proc Natl Acad Sci U S A*. 2007;104(29):12017-22.
159. McGee SL, van Denderen BJ, Howlett KF, Mollica J, Schertzer JD, Kemp BE, et al. AMP-activated protein kinase regulates GLUT4 transcription by phosphorylating histone deacetylase 5. *Diabetes*. 2008;57(4):860-7.
160. Kubota N, Terauchi Y, Yamauchi T, Kubota T, Moroi M, Matsui J, et al. Disruption of adiponectin causes insulin resistance and neointimal formation. *J Biol Chem*. 2002;277(29):25863-6.
161. Berg AH, Combs TP, Du X, Brownlee M, Scherer PE. The adipocyte-secreted protein Acrp30 enhances hepatic insulin action. *Nat Med*. 2001;7(8):947-53.
162. Fruebis J, Tsao TS, Javorschi S, Ebbets-Reed D, Erickson MR, Yen FT, et al. Proteolytic cleavage product of 30-kDa adipocyte complement-related protein increases fatty acid oxidation in muscle and causes weight loss in mice. *Proc Natl Acad Sci U S A*. 2001;98(4):2005-10.
163. Smith AC, Bruce CR, Dyck DJ. AMP kinase activation with AICAR simultaneously increases fatty acid and glucose oxidation in resting rat soleus muscle. *J Physiol*. 2005;565(Pt 2):537-46.
164. Sullivan JE, Brocklehurst KJ, Marley AE, Carey F, Carling D, Beri RK. Inhibition of lipolysis and lipogenesis in isolated rat adipocytes with AICAR, a cell-permeable activator of AMP-activated protein kinase. *FEBS Lett*. 1994;353(1):33-6.
165. Sztalryd C, Komaromy MC, Kraemer FB. Overexpression of hormone-sensitive lipase prevents triglyceride accumulation in adipocytes. *J Clin Invest*. 1995;95(6):2652-61.
166. Kobayashi T, Fujimori K. Very long-chain-fatty acids enhance adipogenesis through coregulation of Elovl3 and PPAR $\gamma$  in 3T3-L1 cells. *Am J Physiol Endocrinol Metab*. 2012;302(12):E1461-71.
167. Ferreira AV, Parreira GG, Green A, Botion LM. Effects of fenofibrate on lipid metabolism in adipose tissue of rats. *Metabolism*. 2006;55(6):731-5.
168. Lee JY, Hashizaki H, Goto T, Sakamoto T, Takahashi N, Kawada T. Activation of peroxisome proliferator-activated receptor- $\alpha$  enhances fatty acid oxidation in human adipocytes. *Biochem Biophys Res Commun*. 2011;407(4):818-22.
169. Ahmadian M, Duncan RE, Varady KA, Frasson D, Hellerstein MK, Birkenfeld AL, et al. Adipose overexpression of desnutrin promotes fatty acid use and attenuates diet-induced obesity. *Diabetes*. 2009;58(4):855-66.
170. Jensen MD, Ekberg K, Landau BR. Lipid metabolism during fasting. *Am J Physiol Endocrinol Metab*. 2001;281(4):E789-93.

171. Reshef L, Olswang Y, Cassuto H, Blum B, Croniger CM, Kalhan SC, et al. Glyceroneogenesis and the triglyceride/fatty acid cycle. *J Biol Chem.* 2003;278(33):30413-6.
172. Newsholme EA. Reflections on the mechanism of action of hormones. *FEBS Lett.* 1980;117 Suppl:K121-34.
173. Brooks B, Arch JR, Newsholme EA. Effects of hormones on the rate of the triacylglycerol/fatty acid substrate cycle in adipocytes and epididymal fat pads. *FEBS Lett.* 1982;146(2):327-30.
174. Wolfe RR, Klein S, Carraro F, Weber JM. Role of triglyceride-fatty acid cycle in controlling fat metabolism in humans during and after exercise. *Am J Physiol.* 1990;258(2 Pt 1):E382-9.
175. Gauthier MS, Miyoshi H, Souza SC, Cacicedo JM, Saha AK, Greenberg AS, et al. AMP-activated protein kinase is activated as a consequence of lipolysis in the adipocyte: potential mechanism and physiological relevance. *J Biol Chem.* 2008;283(24):16514-24.
176. Su CL, Sztalryd C, Contreras JA, Holm C, Kimmel AR, Londos C. Mutational analysis of the hormone-sensitive lipase translocation reaction in adipocytes. *J Biol Chem.* 2003;278(44):43615-9.
177. Yin W, Mu J, Birnbaum MJ. Role of AMP-activated protein kinase in cyclic AMP-dependent lipolysis in 3T3-L1 adipocytes. *J Biol Chem.* 2003;278(44):43074-80.
178. Moule SK, Denton RM. The activation of p38 MAPK by the beta-adrenergic agonist isoproterenol in rat epididymal fat cells. *FEBS Lett.* 1998;439(3):287-90.
179. Kurebayashi S, Hirose T, Miyashita Y, Kasayama S, Kishimoto T. Thiazolidinediones downregulate stearoyl-CoA desaturase 1 gene expression in 3T3-L1 adipocytes. *Diabetes.* 1997;46(12):2115-8.