

HEYDI NORIEGA GUERRA

**IMPORTÂNCIA DA PROTEASE ADAMTS-1 NA INVASÃO
LOCAL E SISTÊMICA DE CÉLULAS DO FIBROSSARCOMA**

Tese apresentada ao Departamento de Biologia Celular e do Desenvolvimento do Instituto de Ciências Biomédicas da Universidade de São Paulo, para a obtenção do Título de Doutor em Ciências.

Área de concentração: Biologia Celular e Tecidual

Orientadora: Profa. Dra. Vanessa Morais Freitas

Versão original

São Paulo
2017

RESUMO

GUERRA, H. N. **Importância da protease ADAMTS-1 na invasão local e sistêmica de células do fibrossarcoma.** 2017. 93 f. Tese (Doutorado em Biologia Celular e Tecidual) - Instituto de Ciências Biomédicas, Universidade de São Paulo, São Paulo, 2017.

O crescimento e a malignidade de um tumor são ditados pelo microambiente circundante. A matriz extracelular, que faz parte do microambiente tumoral, serve como depósito para fatores biologicamente ativos, como fatores de crescimento e proteases, os quais influenciam no comportamento das células tumorais. A ADAMTS-1 (uma desintegrina e metaloproteinase com motivos trombospondina) é uma protease secretada, conhecida por sua capacidade de modificar a matriz extracelular durante os processos fisiológicos e patológicos. Neste trabalho, avaliamos o papel da ADAMTS-1 na regulação das atividades estimuladas pelos fatores de crescimento (HGF e TGF- β 1), sobre a linhagem celular de fibrossarcoma humano (HT1080). Para tanto, foram geradas células HT1080 e HEK293T que superexpressam a protease ADAMTS-1. Ensaios de incorporação de BrdU e expressão de Ki67 demonstraram que a superexpressão de ADAMTS-1 diminuiu a proliferação das células HT1080, na presença do HGF. Ademais, as células HT1080 que superexpressam ADAMTS-1 apresentaram uma velocidade de migração de $6.120 \pm 0.416 \mu\text{m/hora}$, a qual foi 2 vezes menor que a velocidade de migração das células controle ($13.763 \pm 1,421 \mu\text{m/hora}$), ambas na presença do HGF. Da mesma forma, observamos que a proliferação e a velocidade de migração das células HT1080 (tipo selvagem) diminuiu quando estas foram tratadas com o HGF acrescido ao meio condicionado de outro tipo celular (HEK293T) que superexpressa ADAMTS-1. A seguir, demonstramos que a superexpressão da ADAMTS-1 interrompe a ativação do receptor c-Met, após estimulação com o HGF. Conseqüentemente, as vias de sinalização *downstream* ERK1/2 e FAK foram perturbadas. Por outro lado, a superexpressão de ADAMTS-1 não afetou a atividade do TGF- β 1 associada à proliferação celular e velocidade de migração das células HT1080. Embora observamos uma diminuição na fosforilação das smad2/3 nas células que superexpressam ADAMTS-1, esta diminuição não foi suficiente para perturbar as vias de sinalização ERK1/2 e Akt, após o tratamento com o TGF- β 1. Adicionalmente, através de cultura 3D em *Hanging drop* mostramos que na presença do HGF, a superexpressão de ADAMTS-1 diminuiu o diâmetro das fibrosarcoesferas ($\sim 0,7 \text{ mm}$), quando comparado com o controle (diâmetro das fibrosarcoesferas $\sim 1,3 \text{ mm}$). Nos estudos *in vivo* observamos que na presença do HGF, a superexpressão de ADAMTS-1 perturbou a formação de microtumores. E os poucos microtumores formados, incluindo as células individuais, apresentaram características morfológicas de lesões menos invasivas. Nossos dados sugerem que a ADAMTS-1 está envolvida na regulação das atividades estimuladas pelo HGF, nas células do fibrossarcoma. Esta protease pode então representar um mecanismo endógeno no controle da biodisponibilidade de diferentes fatores de crescimento com influência direta sobre o comportamento das células tumorais.

Palavras-chave: Fibrossarcoma. Matriz extracelular. Metaloproteinases da matriz. ADAMTS1. Fator de crescimento do hepatócito. Fator de crescimento transformante beta.

ABSTRACT

GUERRA, H. N. **Importance of ADAMTS-1 protease in local and systemic invasion of fibrosarcoma.** 2017. 93 p. PhD Thesis (Cell and Tissue Biology) - Instituto de Ciências Biomédicas, Universidade de São Paulo, São Paulo, 2017.

The growth and malignancy of a tumor are dictated by the surrounding microenvironment. The extracellular matrix forms part of the tumor microenvironment, and serves as a reservoir for biologically active factors, such as growth factors and proteases that influence the tumor cell behavior. ADAMTS-1 (a disintegrin and metalloproteinase with thrombospondin motifs) is a secreted protease that has the ability to modify the extracellular matrix during physiological and pathological processes. Here, we addressed the role played by ADAMTS-1 regulating HGF and TGF- β 1 activities in the high-grade fibrosarcoma cell line (HT1080). For this, we generated HT1080 and HEK293T cells that overexpress ADAMTS-1. BrdU incorporation and Ki67 expression assays demonstrated that ADAMTS-1 overexpression induced a significant decrease of HT1080 cell proliferation, in the presence of HGF. In addition, the HT1080 cells overexpressing ADAMTS-1 presented a migration velocity of $6.120 \pm 0.416 \mu\text{m}/\text{hour}$, which was at least 2 times lower than the control cell migration velocity ($13.763 \pm 1.421 \mu\text{m}/\text{hour}$), both in presence of HGF. Likewise, when HT1080 cells were treated with HGF added into the conditioned medium of other cell type (HEK293T) that overexpressed ADAMTS-1, we observed a decrease of cell proliferation and migration velocity. Next, we demonstrate that ADAMTS-1 overexpression interrupts c-Met activation upon HGF stimulation. Consequently, the downstream ERK1/2 and FAK signaling pathways are disturbed. On the other hand, ADAMTS-1 overexpression failed to affect TGF- β 1 activity associated with HT1080 cell proliferation and migration velocity. Although we observed a decrease of smad2/3 phosphorylation in HT1080 cells overexpressing ADAMTS-1, this partial reduction was not sufficient to alter ERK1/2 and Akt activation, after treatment with TGF- β 1. Additionally, through 3D culture in Hanging drop we showed that in presence of HGF, ADAMTS-1 overexpression decreased the fibrosarcospheres diameter ($\sim 0.7 \text{ mm}$), as compared to the control (fibrosarcospheres diameter of $\sim 1.3 \text{ mm}$). *In vivo* studies showed that in presence of HGF, ADAMTS-1 overexpression disrupted the formation of microtumors. These microtumors, including individual cells, presented characteristics of low invasive tumor cells (rounded morphology). Our results suggest that ADAMTS-1 is involved in regulating HGF-related functions on fibrosarcoma cells. This protease may then represent an endogenous mechanism in controlling the bioavailability of different growth factors that have a direct influence on tumor cell behavior.

Keywords: Fibrosarcoma. Extracellular matrix. Matrix metalloproteinases. ADAMTS1. Hepatocyte growth factor. Transforming growth factor beta.

INTRODUÇÃO

O fibrossarcoma é um câncer de origem mesenquimal, de alta malignidade e agressividade. Deriva-se do tecido conjuntivo denso e se caracteriza pela presença de fibroblastos malignos (imaturos); os quais apresentam uma divisão celular rápida e desorganizada, formando assim tumores sólidos (JAYAMATHI et al., 2010; NIKITOVIC et al., 2013). Na clínica, o fibrossarcoma apresenta elevado índice de recorrência local e baixa incidência de metástase hematogênica e/ou metástase em linfonodos regionais (KOTRASHETTI et al., 2012; NIKITOVIC et al., 2013).

A progressão do câncer não depende apenas de novas habilidades adquiridas pelas células tumorais, mas também da interação com seu microambiente. Como é sabido, a matriz extracelular (MEC) faz parte do microambiente tumoral; e esta matriz é constituída por uma rede complexa de macromoléculas, tais como glicoproteínas, colágenos, glicosaminoglicanos e proteoglicanos (BRESNICK; WEBER; ZIMMER, 2015; DECLERCK et al., 2004; KIM; TURNBULL; GUIMOND, 2011).

No microambiente tumoral, o papel da MEC não está limitado a atuar somente como uma barreira física contra a invasão tumoral. Também serve como depósito para fatores biologicamente ativos, como fatores de crescimento, proteases, hormônios, entre outros, os quais influenciam no comportamento das células tumorais (DECLERCK et al., 2004).

Encontra-se bem estabelecido na literatura que um evento chave na invasão e metástase dos tumores é a clivagem ou degradação dos componentes da MEC por ação de proteases (FOLKMAN; SHING, 1992). Entre estas proteases temos a ADAMTS-1, o primeiro membro identificado da família ADAMTS (uma desintegrina e metaloproteinase com motivos trombospondina). Esta protease se caracteriza por apresentar um peptídeo sinalizador, um pró-domínio, um domínio metaloproteinase dependente de zinco, um domínio semelhante a desintegrina, três motivos trombospondina tipo I (TSP-1), um domínio rico em cisteína e uma região espaçadora. É importante mencionar que todos os membros da família ADAMTS não apresentam o domínio transmembrânico e são, portanto, secretadas na matriz (KUNO et al., 2004).

O mecanismo de ação da ADAMTS-1 na progressão tumoral ainda não está claramente definido. Embora a molécula inteira de ADAMTS-1 promova a metástase,

os fragmentos da molécula que não possuem atividade catalítica e apresentam um motivo TSP1 na sua estrutura, mostraram um efeito antimetastático. Portanto, dependendo do local de auto-clivagem proteolítica, a ADAMTS-1 pode estar relacionada à atividade antitumoral (PORTER et al., 2005) ou atividade pró-tumoral e estimuladora de metástase (LIU; XU; YU, 2006; LU et al., 2009).

Além disso, ADAMTS-1 possui um efeito anti-angiogênico, devido à ligação direta desta protease com o fator de crescimento do endotélio vascular (VEGF). Esta interação impede então ao VEGF de se ligar e ativar seu respectivo receptor (KUNO et al., 2000; RUSSELL et al., 2003; SANDY et al., 2001). Por outro lado, ADAMTS-1 promove a angiogênese e invasão tumoral, através da sua atividade *shedase* sobre os precursores transmembrânicos do fator de crescimento epidermal ligado a heparina (HB-EGF), e a subsequente ativação do seu respectivo receptor (LIU; XU; YU, 2006). Dessa forma, ADAMTS-1 representa um mecanismo endógeno no controle da biodisponibilidade dos diferentes fatores de crescimento (LIU; XU; YU, 2006; MARGOSIO et al., 2003).

Neste trabalho, buscamos elucidar o papel da protease ADAMTS-1 e sua íntima ligação com o microambiente tumoral do fibrossarcoma. Nosso enfoque foi estudar o efeito da protease ADAMTS-1 na regulação das atividades estimuladas pelos fatores de crescimento (HGF e TGF- β 1), sobre a linhagem celular derivada de fibrossarcoma.

Os ensaios foram então realizados a partir de células de fibrossarcoma humano (HT1080) transduzidas para superexpressar ADAMTS-1, e as quais foram tratadas apenas com os fatores de crescimento de interesse. Por outro lado, células de fibrossarcoma não transduzidas (selvagens) foram tratadas com os fatores de crescimento de interesse adicionados ao meio condicionado coletado de outro tipo celular, o qual também foi transduzido para superexpressar ADAMTS-1. Desta forma, o meio condicionado enriquecido com ADAMTS-1, mimetiza a protease secretada por outros tipos celulares presentes no microambiente tumoral.

A proliferação celular foi analisada através dos ensaios de incorporação de Bromodesoxiuridina (BrdU) e expressão de Ki67. A velocidade de migração foi determinada utilizando-se ensaios de *time-lapse* e o plugin *MTrackJ* do *software Fiji/ImageJ*. Investigamos também os mecanismos através dos quais ADAMTS-1 interfere com as atividades estimuladas por HGF ou TGF- β 1. Além disso, estudamos por meio de ensaios de cultura 3D em *Hanging Drop*, a capacidade de formação de

fibrosarcoesferas em diferentes condições. Por outro lado, a capacidade de formação de microtumores *in vivo* foi analisada utilizando-se o ensaio de microinjeção de células tumorais em embriões de *zebrafish* transgênicos Tg (fli1:EGFP).

CONCLUSÃO

Baseados nos resultados dos experimentos realizados, concluímos que a superexpressão de ADAMTS-1:

- i.) afeta os processos celulares de proliferação e migração das células de fibrossarcoma (HT1080), estimulados pelo fator de crescimento de hepatócitos (HGF);
- ii.) não afeta as respostas celulares de proliferação e migração em células HT1080, estimuladas pelo fator de crescimento transformante beta (TGF- β 1);
- iii.) interrompe a ativação do receptor c-Met, além das suas vias de sinalização *downstream* ERK1/2 e FAK, nas células HT1080 após 15 minutos de estimulação com o HGF;
- iv.) prejudica a formação de fibrosarcoesferas a partir das células HT1080 estimuladas com HGF;
- v.) perturba a formação de microtumores de HT1080 *in vivo*, após estimulação com o HGF. Os microtumores e as células que não tiveram a capacidade de formar microtumores apresentaram características morfológicas de lesões menos invasivas.

REFERÊNCIAS*

AKHURST, R. J.; HATA, A. Targeting the TGF β signalling pathway in disease. **Nat Rev Drug Discov**, v. 11, n. 10, p. 790-811, Oct 2012.

ALEXANDER, S.; WEIGELIN, B.; WINKLER, F.; FRIEDL, P. Preclinical intravital microscopy of the tumour-stroma interface: invasion, metastasis, and therapy response. **Curr Opin Cell Biol**, v. 25, n. 5, p. 659-671, Oct 2013.

ALJABAB, A. S.; NASON, R. W.; KAZI, R.; PATHAK, K. A. Head and neck soft tissue sarcoma. **Indian J Surg Oncol**, v. 2, n. 4, p. 286-290, Dec 2011.

ALLEN, M.; LOUISE JONES, J. Jekyll and Hyde: the role of the microenvironment on the progression of cancer. **J Pathol**, v. 223, n. 2, p. 162-176, Jan 2011.

APTE, S. S. A disintegrin-like and metalloprotease (reprolysin type) with thrombospondin type 1 motifs: the ADAMTS family. **Int J Biochem Cell Biol**, v. 36, n. 6, p. 981-985, Jun 2004.

_____. A disintegrin-like and metalloprotease (reprolysin-type) with thrombospondin type 1 motif (ADAMTS) superfamily: functions and mechanisms. **J Biol Chem**, v. 284, n. 46, p. 31493-31497, Nov 2009.

ARRIBAS, J.; BECH-SERRA, J. J.; SANTIAGO-JOSEFAT, B. ADAMs, cell migration and cancer. **Cancer Metastasis Rev**, v. 25, n. 1, p. 57-68, Mar 2006.

BAHRAMI, A.; FOLPE, A. L. Adult-type fibrosarcoma: A reevaluation of 163 putative cases diagnosed at a single institution over a 48-year period. **Am J Surg Pathol**, v. 34, n. 10, p. 1504-1513, Oct 2010.

BASILICO, C.; ARNESANO, A.; GALLUZZO, M.; COMOGLIO, P. M.; MICHIELI, P. A high affinity hepatocyte growth factor-binding site in the immunoglobulin-like region of Met. **J Biol Chem**, v. 283, n. 30, p. 21267-21277, Jul 2008.

BHOWMICK, N. A.; NEILSON, E. G.; MOSES, H. L. Stromal fibroblasts in cancer initiation and progression. **Nature**, v. 432, n. 7015, p. 332-337, Nov 2004.

BIANCO, R.; MELISI, D.; CIARDIELLO, F.; TORTORA, G. Key cancer cell signal transduction pathways as therapeutic targets. **Eur J Cancer**, v. 42, n. 3, p. 290-294, Feb 2006.

BIRCHMEIER, C.; BIRCHMEIER, W.; GHERARDI, E.; VANDE WOUDE, G. F. Met, metastasis, motility and more. **Nat Rev Mol Cell Biol**, v. 4, n. 12, p. 915-925, Dec 2003.

BLOBEL, C. P. Metalloprotease-disintegrins: links to cell adhesion and cleavage of TNF alpha and Notch. **Cell**, v. 90, n. 4, p. 589-592, Aug 1997.

BOBROVNIKOVA-MARJON, E. V.; MARJON, P. L.; BARBASH, O.; VANDER JAGT, D. L.; ABCOUWER, S. F. Expression of angiogenic factors vascular endothelial growth factor and

* De acordo com:

ASSOCIAÇÃO BRASILEIRA DE NORMAS TÉCNICAS. **NBR 6023**: informação e documentação: referências: elaboração. Rio de Janeiro, 2002.

interleukin-8/CXCL8 is highly responsive to ambient glutamine availability: role of nuclear factor-kappaB and activating protein-1. **Cancer Res**, v. 64, n. 14, p. 4858-4869, Jul 2004.

BORNSTEIN, P.; SAGE, E. H. Matricellular proteins: extracellular modulators of cell function. **Curr Opin Cell Biol**, v. 14, n. 5, p. 608-616, Oct 2002.

BOTTARO, D. P.; RUBIN, J. S.; FALETTO, D. L.; CHAN, A. M.; KMIECIK, T. E.; VANDE WOUDE, G. F.; AARONSON, S. A. Identification of the hepatocyte growth factor receptor as the c-met proto-oncogene product. **Science**, v. 251, n. 4995, p. 802-804, Feb 1991.

BOUCK, N.; STELLMACH, V.; HSU, S. C. How tumors become angiogenic. **Adv Cancer Res**, v. 69, p. 135-174, 1996.

BOURD-BOITTIN, K.; BONNIER, D.; LEYME, A.; MARI, B.; TUFFERY, P.; SAMSON, M.; EZAN, F.; BAFFET, G.; THERET, N. Protease profiling of liver fibrosis reveals the ADAM metalloproteinase with thrombospondin type 1 motif, 1 as a central activator of transforming growth factor beta. **Hepatology**, v. 54, n. 6, p. 2173-2184, Dec 2011.

BRASILEIRO, G. **Bogliolo Patologia Geral**. 3. Rio de Janeiro: Guanabara Koogan, 2004.

BREMNES, R. M.; DØNNEM, T.; AL-SAAD, S.; AL-SHIBLI, K.; ANDERSEN, S.; SIRERA, R.; CAMPS, C.; MARINEZ, I.; BUSUND, L. T. The role of tumor stroma in cancer progression and prognosis: emphasis on carcinoma-associated fibroblasts and non-small cell lung cancer. **J Thorac Oncol**, v. 6, n. 1, p. 209-217, Jan 2011.

BRESNICK, A. R.; WEBER, D. J.; ZIMMER, D. B. S100 proteins in cancer. **Nat Rev Cancer**, v. 15, n. 2, p. 96-109, Feb 2015.

BROWN, H. M.; DUNNING, K. R.; ROBKER, R. L.; PRITCHARD, M.; RUSSELL, D. L. Requirement for ADAMTS-1 in extracellular matrix remodeling during ovarian folliculogenesis and lymphangiogenesis. **Dev Biol**, v. 300, n. 2, p. 699-709, Dec 2006.

BURT, D. W.; LAW, A. S. Evolution of the transforming growth factor-beta superfamily. **Prog Growth Factor Res**, v. 5, n. 1, p. 99-118, 1994.

CANALS, F.; COLOMÉ, N.; FERRER, C.; PLAZA-CALONGE, M. E. C.; RODRÍGUEZ-MANZANEQUE, J. C. Identification of substrates of the extracellular protease ADAMTS1 by DIGE proteomic analysis. **Proteomics**, v. 6 Suppl 1, p. S28-35, Apr 2006.

CASAL, C.; TORRES-COLLADO, A. X.; PLAZA-CALONGE, M. E. C.; MARTINO-ECHARRI, E.; RAMÓN Y CAJAL, S.; ROJO, F.; GRIFFIOEN, A. W.; RODRÍGUEZ-MANZANEQUE, J. C. ADAMTS1 contributes to the acquisition of an endothelial-like phenotype in plastic tumor cells. **Cancer Res**, v. 70, n. 11, p. 4676-4686, Jun 2010.

CAUSSINUS, E.; COLOMBELLI, J.; AFFOLTER, M. Tip-cell migration controls stalk-cell intercalation during Drosophila tracheal tube elongation. **Curr Biol**, v. 18, n. 22, p. 1727-1734, Nov 2008.

CAVALLARO, U.; CHRISTOFORI, G. Cell adhesion and signalling by cadherins and Ig-CAMs in cancer. **Nat Rev Cancer**, v. 4, n. 2, p. 118-132, Feb 2004.

CHAMBERS, A. F.; MATRISIAN, L. M. Changing views of the role of matrix metalloproteinases in metastasis. **J Natl Cancer Inst**, v. 89, n. 17, p. 1260-1270, Sep 1997.

CHANG, C.; WERB, Z. The many faces of metalloproteases: cell growth, invasion, angiogenesis and metastasis. **Trends Cell Biol**, v. 11, n. 11, p. S37-43, Nov 2001.

CHEN, X.; XIAO, W.; WANG, W.; LUO, L.; YE, S.; LIU, Y. The complex interplay between ERK1/2, TGF β /Smad, and Jagged/Notch signaling pathways in the regulation of epithelial-mesenchymal transition in retinal pigment epithelium cells. **PLoS One**, v. 9, n. 5, p. e96365, 2014.

CHRISTIAN, L. M. The ADAM family: Insights into Notch proteolysis. **Fly (Austin)**, v. 6, n. 1, p. 30-34, 2012 Jan-Mar 2012.

CLARK, A. G.; VIGNJEVIC, D. M. Modes of cancer cell invasion and the role of the microenvironment. **Curr Opin Cell Biol**, v. 36, p. 13-22, Oct 2015.

COLLINI, P.; SORENSEN, P. H.; PATEL, S.; BLAY, J. Y.; ISSELS, R. D.; MAKI, R. G.; ERIKSSON, M.; DEL MURO, X. G. Sarcomas with spindle cell morphology. **Semin Oncol**, v. 36, n. 4, p. 324-337, Aug 2009.

COMOGLIO, P. M.; GIORDANO, S.; TRUSOLINO, L. Drug development of MET inhibitors: targeting oncogene addiction and expedience. **Nat Rev Drug Discov**, v. 7, n. 6, p. 504-516, Jun 2008.

CONERY, A. R.; CAO, Y.; THOMPSON, E. A.; TOWNSEND, C. M.; KO, T. C.; LUO, K. Akt interacts directly with Smad3 to regulate the sensitivity to TGF-beta induced apoptosis. **Nat Cell Biol**, v. 6, n. 4, p. 366-372, Apr 2004.

COOPER, C. S. The met oncogene: from detection by transfection to transmembrane receptor for hepatocyte growth factor. **Oncogene**, v. 7, n. 1, p. 3-7, Jan 1992.

COOPER, G. The Development and Causes of Cancer. In: 2 (Ed.). **The Cell: A Molecular Approach**. Sunderland (MA): Sinauer Associates, 2000.

CORSO, S.; COMOGLIO, P. M.; GIORDANO, S. Cancer therapy: can the challenge be MET? **Trends Mol Med**, v. 11, n. 6, p. 284-292, Jun 2005.

COUSSENS, L. M.; WERB, Z. Matrix metalloproteinases and the development of cancer. **Chem Biol**, v. 3, n. 11, p. 895-904, Nov 1996.

CRAGO, A. M.; BRENNAN, M. F. Principles in Management of Soft Tissue Sarcoma. **Adv Surg**, v. 49, p. 107-122, 2015.

DE HERDT, M. J.; BAATENBURG DE JONG, R. J. HGF and c-MET as potential orchestrators of invasive growth in head and neck squamous cell carcinoma. **Front Biosci**, v. 13, p. 2516-2526, Jan 2008.

DE VISSER, K. E.; KORETS, L. V.; COUSSENS, L. M. De novo carcinogenesis promoted by chronic inflammation is B lymphocyte dependent. **Cancer Cell**, v. 7, n. 5, p. 411-423, May 2005.

DECLERCK, Y. A.; MERCURIO, A. M.; STACK, M. S.; CHAPMAN, H. A.; ZUTTER, M. M.; MUSCHEL, R. J.; RAZ, A.; MATRISIAN, L. M.; SLOANE, B. F.; NOEL, A.; HENDRIX, M. J.; COUSSENS, L.; PADARATHSINGH, M. Proteases, extracellular matrix, and cancer: a workshop of the path B study section. **Am J Pathol**, v. 164, n. 4, p. 1131-1139, Apr 2004.

DERKSEN, P. W.; LIU, X.; SARIDIN, F.; VAN DER GULDEN, H.; ZEVENHOVEN, J.; EVERS, B.; VAN BEIJNUM, J. R.; GRIFFIOEN, A. W.; VINK, J.; KRIMPENFORT, P.; PETERSE, J. L.; CARDIFF, R. D.; BERNS, A.; JONKERS, J. Somatic inactivation of E-cadherin and p53 in mice leads to metastatic lobular mammary carcinoma through induction of anoikis resistance and angiogenesis. **Cancer Cell**, v. 10, n. 5, p. 437-449, Nov 2006.

DERYNCK, R.; ZHANG, Y. E. Smad-dependent and Smad-independent pathways in TGF-beta family signalling. **Nature**, v. 425, n. 6958, p. 577-584, Oct 2003.

DERYUGINA, E. I.; RATNIKOV, B. I.; YU, Q.; BACIU, P. C.; ROZANOV, D. V.; STRONGIN, A. Y. Prointegrin maturation follows rapid trafficking and processing of MT1-MMP in Furin-Negative Colon Carcinoma LoVo Cells. **Traffic**, v. 5, n. 8, p. 627-641, Aug 2004.

DEVY, L.; BLACHER, S.; GRIGNET-DEBRUS, C.; BAJOU, K.; MASSON, V.; GERARD, R. D.; GILS, A.; CARMELIET, G.; CARMELIET, P.; DECLERCK, P. J.; NÖEL, A.; FOIDART, J. M. The pro- or antiangiogenic effect of plasminogen activator inhibitor 1 is dose dependent. **FASEB J**, v. 16, n. 2, p. 147-154, Feb 2002.

DIRAT, B.; BOCHET, L.; DABEK, M.; DAVIAUD, D.; DAUVILLIER, S.; MAJED, B.; WANG, Y. Y.; MEULLE, A.; SALLES, B.; LE GONIDEC, S.; GARRIDO, I.; ESCOURROU, G.; VALET, P.; MULLER, C. Cancer-associated adipocytes exhibit an activated phenotype and contribute to breast cancer invasion. **Cancer Res**, v. 71, n. 7, p. 2455-2465, Apr 2011.

EDWARDS, D. R.; HANDSLEY, M. M.; PENNINGTON, C. J. The ADAM metalloproteinases. **Mol Aspects Med**, v. 29, n. 5, p. 258-289, Oct 2008.

ENGELLAU, J.; ANDERSON, H.; RYDHOLM, A.; BAUER, H. C.; HALL, K. S.; GUSTAFSON, P.; AKERMAN, M.; MEIS-KINDBLOM, J.; ALVEGÅRD, T. A.; NILBERT, M.; GROUP, S. S. Time dependence of prognostic factors for patients with soft tissue sarcoma: a Scandinavian Sarcoma Group Study of 338 malignant fibrous histiocytomas. **Cancer**, v. 100, n. 10, p. 2233-2239, May 2004.

FICHER, C.; VAN DEN BERG, E.; MOLENAAR, W. Adult Fibrosarcoma. In: FLETCHER, C. M.; UNNI, K., *et al* (Ed.). **World Health Organization Classification of Tumours - Pathology and Genetics of Tumours of Soft Tissue and Bone**: Lyon: IARC Press, 2002. p.100-101.

FLUHRER, R.; HAASS, C. Signal peptide peptidases and gamma-secretase: cousins of the same protease family? **Neurodegener Dis**, v. 4, n. 2-3, p. 112-116, 2007.

FOLKMAN, J. Tumor angiogenesis: therapeutic implications. **N Engl J Med**, v. 285, n. 21, p. 1182-1186, Nov 1971.

_____. Fundamental concepts of the angiogenic process. **Curr Mol Med**, v. 3, n. 7, p. 643-651, Nov 2003.

FOLKMAN, J.; SHING, Y. Angiogenesis. **J Biol Chem**, v. 267, n. 16, p. 10931-10934, Jun 1992.

FOLPE, A. L. Fibrosarcoma: a review and update. **Histopathology**, v. 64, n. 1, p. 12-25, Jan 2014.

FOULDS, L. The experimental study of tumor progression: a review. **Cancer Res**, v. 14, n. 5, p. 327-339, Jun 1954.

FREITAS, V. M.; DO AMARAL, J. B.; SILVA, T. A.; SANTOS, E. S.; MANGONE, F. R.; PINHEIRO, J. E. J.; JAEGER, R. G.; NAGAI, M. A.; MACHADO-SANTELLI, G. M. Decreased expression of ADAMTS-1 in human breast tumors stimulates migration and invasion. **Mol Cancer**, v. 12, p. 2, 2013.

FRIEDL, P.; ALEXANDER, S. Cancer invasion and the microenvironment: plasticity and reciprocity. **Cell**, v. 147, n. 5, p. 992-1009, Nov 2011.

FRISCH, S. M.; FRANCIS, H. Disruption of epithelial cell-matrix interactions induces apoptosis. **J Cell Biol**, v. 124, n. 4, p. 619-626, Feb 1994.

FURGE, K. A.; ZHANG, Y. W.; VANDE WOUDE, G. F. Met receptor tyrosine kinase: enhanced signaling through adapter proteins. **Oncogene**, v. 19, n. 49, p. 5582-5589, Nov 2000.

FUSHIMI, K.; TROEBERG, L.; NAKAMURA, H.; LIM, N. H.; NAGASE, H. Functional differences of the catalytic and non-catalytic domains in human ADAMTS-4 and ADAMTS-5 in aggrecanolytic activity. **J Biol Chem**, v. 283, n. 11, p. 6706-6716, Mar 2008.

GANDINO, L.; DI RENZO, M. F.; GIORDANO, S.; BUSSOLINO, F.; COMOGLIO, P. M. Protein kinase-c activation inhibits tyrosine phosphorylation of the c-met protein. **Oncogene**, v. 5, n. 5, p. 721-725, May 1990.

GANDINO, L.; LONGATI, P.; MEDICO, E.; PRAT, M.; COMOGLIO, P. M. Phosphorylation of serine 985 negatively regulates the hepatocyte growth factor receptor kinase. **J Biol Chem**, v. 269, n. 3, p. 1815-1820, Jan 1994.

GANDINO, L.; MUNARON, L.; NALDINI, L.; FERRACINI, R.; MAGNI, M.; COMOGLIO, P. M. Intracellular calcium regulates the tyrosine kinase receptor encoded by the MET oncogene. **J Biol Chem**, v. 266, n. 24, p. 16098-16104, Aug 1991.

GANTT, K. R.; SCHULTZ-CHERRY, S.; RODRIGUEZ, N.; JERONIMO, S. M.; NASCIMENTO, E. T.; GOLDMAN, T. L.; RECKER, T. J.; MILLER, M. A.; WILSON, M. E. Activation of TGF-beta by Leishmania chagasi: importance for parasite survival in macrophages. **J Immunol**, v. 170, n. 5, p. 2613-2620, Mar 2003.

GAO, C. F.; VANDE WOUDE, G. F. HGF/SF-Met signaling in tumor progression. **Cell Res**, v. 15, n. 1, p. 49-51, Jan 2005.

GENDRON, C.; KASHIWAGI, M.; LIM, N. H.; ENGHILD, J. J.; THØGERSEN, I. B.; HUGHES, C.; CATERSON, B.; NAGASE, H. Proteolytic activities of human ADAMTS-5: comparative studies with ADAMTS-4. **J Biol Chem**, v. 282, n. 25, p. 18294-18306, Jun 2007.

GERDES, M. J.; SOOD, A.; SEVINSKY, C.; PRIS, A. D.; ZAVODSZKY, M. I.; GINTY, F. Emerging understanding of multiscale tumor heterogeneity. **Front Oncol**, v. 4, p. 366, 2014.

GERHARDT, S.; HASSALL, G.; HAWTIN, P.; MCCALL, E.; FLAVELL, L.; MINSHULL, C.; HARGREAVES, D.; TING, A.; PAUPTIT, R. A.; PARKER, A. E.; ABBOTT, W. M. Crystal structures of human ADAMTS-1 reveal a conserved catalytic domain and a disintegrin-like domain with a fold homologous to cysteine-rich domains. **J Mol Biol**, v. 373, n. 4, p. 891-902, Nov 2007.

GHEBRANIOUS, N.; DONEHOWER, L. A. Mouse models in tumor suppression. **Oncogene**, v. 17, n. 25, p. 3385-3400, Dec 1998.

GIANNONI, E.; FIASCHI, T.; RAMPONI, G.; CHIARUGI, P. Redox regulation of anoikis resistance of metastatic prostate cancer cells: key role for Src and EGFR-mediated pro-survival signals. **Oncogene**, v. 28, n. 20, p. 2074-2086, May 2009.

GKRETSI, V.; STYLIANOU, A.; PAPAGEORGIS, P.; POLYDOROU, C.; STYLIANOPOULOS, T. Remodeling Components of the Tumor Microenvironment to Enhance Cancer Therapy. **Front Oncol**, v. 5, p. 214, 2015.

GOMIS-RÜTH, F. X. Catalytic domain architecture of metzincin metalloproteases. **J Biol Chem**, v. 284, n. 23, p. 15353-15357, Jun 2009.

GRASSO, G.; BONNET, S. Metal complexes and metalloproteases: targeting conformational diseases. **Metallomics**, v. 6, n. 8, p. 1346-1357, Aug 2014.

GRESSNER, O. A.; WEISKIRCHEN, R.; GRESSNER, A. M. Evolving concepts of liver fibrogenesis provide new diagnostic and therapeutic options. **Comp Hepatol**, v. 6, p. 7, Jul 2007.

GRIVENNIKOV, S. I.; GRETEN, F. R.; KARIN, M. Immunity, inflammation, and cancer. **Cell**, v. 140, n. 6, p. 883-899, Mar 2010.

GÜNTHER, W.; SKAFTNESMO, K. O.; ARNOLD, H.; BJERKVIG, R.; TERZIS, A. J. Distribution patterns of the anti-angiogenic protein ADAMTS-1 during rat development. **Acta Histochem**, v. 107, n. 2, p. 121-131, 2005.

GUO, B. H.; FENG, Y.; ZHANG, R.; XU, L. H.; LI, M. Z.; KUNG, H. F.; SONG, L. B.; ZENG, M. S. Bmi-1 promotes invasion and metastasis, and its elevated expression is correlated with an advanced stage of breast cancer. **Mol Cancer**, v. 10, n. 1, p. 10, Jan 2011.

GUO, M.; MATHIEU, P. A.; LINEBAUGH, B.; SLOANE, B. F.; REINERS, J. J. Phorbol ester activation of a proteolytic cascade capable of activating latent transforming growth factor-betaL a process initiated by the exocytosis of cathepsin B. **J Biol Chem**, v. 277, n. 17, p. 14829-14837, Apr 2002.

GUO, Y.; XIE, J.; RUBIN, E.; TANG, Y. X.; LIN, F.; ZI, X.; HOANG, B. H. Frzb, a secreted Wnt antagonist, decreases growth and invasiveness of fibrosarcoma cells associated with inhibition of Met signaling. **Cancer Res**, v. 68, n. 9, p. 3350-3360, May 2008.

GUPTA, K.; GUPTA, P.; WILD, R.; RAMAKRISHNAN, S.; HEBBEL, R. P. Binding and displacement of vascular endothelial growth factor (VEGF) by thrombospondin: effect on human microvascular endothelial cell proliferation and angiogenesis. **Angiogenesis**, v. 3, n. 2, p. 147-158, 1999.

GUSTAFSON, P. Soft tissue sarcoma. Epidemiology and prognosis in 508 patients. **Acta Orthop Scand Suppl**, v. 259, p. 1-31, Jun 1994.

HAAS, P.; GILMOUR, D. Chemokine signaling mediates self-organizing tissue migration in the zebrafish lateral line. **Dev Cell**, v. 10, n. 5, p. 673-680, May 2006.

HANAHAN, D.; COUSSENS, L. M. Accessories to the crime: functions of cells recruited to the tumor microenvironment. **Cancer Cell**, v. 21, n. 3, p. 309-322, Mar 2012.

HANAHAN, D.; FOLKMAN, J. Patterns and emerging mechanisms of the angiogenic switch during tumorigenesis. **Cell**, v. 86, n. 3, p. 353-364, Aug 1996.

HANAHAN, D.; WEINBERG, R. A. Hallmarks of cancer: the next generation. **Cell**, v. 144, n. 5, p. 646-674, Mar 2011.

HANDSLEY, M. M.; EDWARDS, D. R. Metalloproteinases and their inhibitors in tumor angiogenesis. **Int J Cancer**, v. 115, n. 6, p. 849-860, Jul 2005.

HARPER, E.; BLOCH, K. J.; GROSS, J. The zymogen of tadpole collagenase. **Biochemistry**, v. 10, n. 16, p. 3035-3041, Aug 1971.

HARRIS, C. C. p53 tumor suppressor gene: from the basic research laboratory to the clinic--an abridged historical perspective. **Carcinogenesis**, v. 17, n. 6, p. 1187-1198, Jun 1996.

HELDIN, C. H.; MIYAZONO, K.; TEN DIJKE, P. TGF-beta signalling from cell membrane to nucleus through SMAD proteins. **Nature**, v. 390, n. 6659, p. 465-471, Dec 1997.

HELMAN, L. J.; MELTZER, P. Mechanisms of sarcoma development. **Nat Rev Cancer**, v. 3, n. 9, p. 685-694, Sep 2003.

HEMLER, M. E.; RUTISHAUSER, U. Cell-to-cell contact and extracellular matrix. Editorial overview. **Curr Opin Cell Biol**, v. 12, n. 5, p. 539-541, Oct 2000.

HU, L.; JONSSON, K. B.; ANDERSÉN, H.; EDENRO, A.; BOHLOOLY-Y, M.; MELHUS, H.; LIND, T. Over-expression of Adamts1 in mice alters bone mineral density. **J Bone Miner Metab**, v. 30, n. 3, p. 304-311, May 2012.

HUA, H.; LI, M.; LUO, T.; YIN, Y.; JIANG, Y. Matrix metalloproteinases in tumorigenesis: an evolving paradigm. **Cell Mol Life Sci**, v. 68, n. 23, p. 3853-3868, Dec 2011.

HUANG, T. F. What have snakes taught us about integrins? **Cell Mol Life Sci**, v. 54, n. 6, p. 527-540, Jun 1998.

HYNES, R. O. The extracellular matrix: not just pretty fibrils. **Science**, v. 326, n. 5957, p. 1216-1219, Nov 2009.

IKEJIRI, M.; BERNARDO, M. M.; BONFIL, R. D.; TOTH, M.; CHANG, M.; FRIDMAN, R.; MOBASHERY, S. Potent mechanism-based inhibitors for matrix metalloproteinases. **J Biol Chem**, v. 280, n. 40, p. 33992-34002, Oct 2005.

ILASLAN, H.; SCHILS, J.; NAGEOTTE, W.; LIETMAN, S. A.; SUNDARAM, M. Clinical presentation and imaging of bone and soft-tissue sarcomas. **Cleve Clin J Med**, v. 77 Suppl 1, p. S2-7, Mar 2010.

INOKI, I.; SHIOMI, T.; HASHIMOTO, G.; ENOMOTO, H.; NAKAMURA, H.; MAKINO, K.; IKEDA, E.; TAKATA, S.; KOBAYASHI, K.; OKADA, Y. Connective tissue growth factor binds vascular endothelial growth factor (VEGF) and inhibits VEGF-induced angiogenesis. **FASEB J**, v. 16, n. 2, p. 219-221, Feb 2002.

IRUELA-ARISPE, M. L.; CARPIZO, D.; LUQUE, A. ADAMTS1: a matrix metalloprotease with angioinhibitory properties. **Ann N Y Acad Sci**, v. 995, p. 183-190, May 2003.

JAKOWLEW, S. B. Transforming growth factor-beta in cancer and metastasis. **Cancer Metastasis Rev**, v. 25, n. 3, p. 435-457, Sep 2006.

JAYAMATHI, P.; KEERTHIDAA, G.; VIDYALAKHSMI, K.; BHAVANI, G.; RUKMANI DEVI, S. Antioxidant Property of Plumbagin on Fibrosarcoma Induced Rats. **Recent Research in**

Science and Technology, v. 2, n. 11, 2010.

JIANG, W.; HISCOX, S.; MATSUMOTO, K.; NAKAMURA, T. Hepatocyte growth factor/scatter factor, its molecular, cellular and clinical implications in cancer. **Crit Rev Oncol Hematol**, v. 29, n. 3, p. 209-248, Feb 1999.

KALLURI, R.; ZEISBERG, M. Fibroblasts in cancer. **Nat Rev Cancer**, v. 6, n. 5, p. 392-401, May 2006.

KARAGIANNIS, E. D.; POPEL, A. S. Anti-angiogenic peptides identified in thrombospondin type I domains. **Biochem Biophys Res Commun**, v. 359, n. 1, p. 63-69, Jul 2007.

KELLER, E. T.; LI, L. Y. The first Tianjin, China forum on tumor microenvironment. **Cancer Res**, v. 71, n. 2, p. 310-313, Jan 2011.

KELLER, K. E.; BRADLEY, J. M.; ACOTT, T. S. Differential effects of ADAMTS-1, -4, and -5 in the trabecular meshwork. **Invest Ophthalmol Vis Sci**, v. 50, n. 12, p. 5769-5777, Dec 2009.

KESSENBROCK, K.; PLAKS, V.; WERB, Z. Matrix metalloproteinases: regulators of the tumor microenvironment. **Cell**, v. 141, n. 1, p. 52-67, Apr 2010.

KHALIL, N. TGF-beta: from latent to active. **Microbes Infect**, v. 1, n. 15, p. 1255-1263, Dec 1999.

KIM, S. H.; TURNBULL, J.; GUIMOND, S. Extracellular matrix and cell signalling: the dynamic cooperation of integrin, proteoglycan and growth factor receptor. **J Endocrinol**, v. 209, n. 2, p. 139-151, May 2011.

KOTRASHETTI, V. S.; KALE, A. D.; HALLIKEREMATH, S. R.; MANE, D. R.; V ANGADI, P.; BHATT, P. Intraosseous fibrosarcoma of maxilla in an HIV patient. **Arch Iran Med**, v. 15, n. 1, p. 59-62, Jan 2012.

KRAUSE, D. S.; VAN ETTEN, R. A. Tyrosine kinases as targets for cancer therapy. **N Engl J Med**, v. 353, n. 2, p. 172-187, Jul 2005.

KRETZSCHMAR, M.; MASSAGUÉ, J. SMADs: mediators and regulators of TGF-beta signaling. **Curr Opin Genet Dev**, v. 8, n. 1, p. 103-111, Feb 1998.

KUBOTA, S.; FRIDMAN, R.; YAMADA, Y. Transforming growth factor-beta suppresses the invasiveness of human fibrosarcoma cells in vitro by increasing expression of tissue inhibitor of metalloprotease. **Biochem Biophys Res Commun**, v. 176, n. 1, p. 129-136, Apr 1991.

KUNDU, J. K.; SURH, Y. J. Inflammation: gearing the journey to cancer. **Mutat Res**, v. 659, n. 1-2, p. 15-30, 2008 Jul-Aug 2008.

KUNO, K.; BANNAI, K.; HAKOZAKI, M.; MATSUSHIMA, K.; HIROSE, K. The carboxyl-terminal half region of ADAMTS-1 suppresses both tumorigenicity and experimental tumor metastatic potential. **Biochem Biophys Res Commun**, v. 319, n. 4, p. 1327-1333, Jul 2004.

KUNO, K.; KANADA, N.; NAKASHIMA, E.; FUJIKI, F.; ICHIMURA, F.; MATSUSHIMA, K. Molecular cloning of a gene encoding a new type of metalloproteinase-disintegrin family protein with thrombospondin motifs as an inflammation associated gene. **J Biol Chem**, v. 272, n. 1, p. 556-562, Jan 1997.

KUNO, K.; OKADA, Y.; KAWASHIMA, H.; NAKAMURA, H.; MIYASAKA, M.; OHNO, H.; MATSUSHIMA, K. ADAMTS-1 cleaves a cartilage proteoglycan, aggrecan. **FEBS Lett**, v. 478, n. 3, p. 241-245, Aug 2000.

KURNIAWAN, N. A.; CHAUDHURI, P. K.; LIM, C. T. Mechanobiology of cell migration in the context of dynamic two-way cell-matrix interactions. **J Biomech**, v. 49, n. 8, p. 1355-1368, May 2016.

KUSZYK, B. S.; CORL, F. M.; FRANANO, F. N.; BLUEMKE, D. A.; HOFMANN, L. V.; FORTMAN, B. J.; FISHMAN, E. K. Tumor transport physiology: implications for imaging and imaging-guided therapy. **AJR Am J Roentgenol**, v. 177, n. 4, p. 747-753, Oct 2001.

KWAK, H. J.; PARK, M. J.; CHO, H.; PARK, C. M.; MOON, S. I.; LEE, H. C.; PARK, I. C.; KIM, M. S.; RHEE, C. H.; HONG, S. I. Transforming growth factor-beta1 induces tissue inhibitor of metalloproteinase-1 expression via activation of extracellular signal-regulated kinase and Sp1 in human fibrosarcoma cells. **Mol Cancer Res**, v. 4, n. 3, p. 209-220, Mar 2006.

LAI, Y.; SHEN, Y.; LIU, X. H.; ZHANG, Y.; ZENG, Y.; LIU, Y. F. Interleukin-8 induces the endothelial cell migration through the activation of phosphoinositide 3-kinase-Rac1/RhoA pathway. **Int J Biol Sci**, v. 7, n. 6, p. 782-791, 2011.

LANDRISCINA, M.; MADDALENA, F.; LAUDIERO, G.; ESPOSITO, F. Adaptation to oxidative stress, chemoresistance, and cell survival. **Antioxid Redox Signal**, v. 11, n. 11, p. 2701-2716, Nov 2009.

LAWLER, J. Thrombospondin-1 as an endogenous inhibitor of angiogenesis and tumor growth. **J Cell Mol Med**, v. 6, n. 1, p. 1-12, 2002 Jan-Mar 2002.

LAWRENCE, D. A. Transforming growth factor-beta: a general review. **Eur Cytokine Netw**, v. 7, n. 3, p. 363-374, Sep 1996.

LE GOFF, C.; CORMIER-DAIRE, V. The ADAMTS(L) family and human genetic disorders. **Hum Mol Genet**, v. 20, n. R2, p. R163-167, Oct 2011.

LESKO, E.; MAJKA, M. The biological role of HGF-MET axis in tumor growth and development of metastasis. **Front Biosci**, v. 13, p. 1271-1280, Jan 2008.

LI, N.; LORINCZI, M.; IRETON, K.; ELFERINK, L. A. Specific Grb2-mediated interactions regulate clathrin-dependent endocytosis of the cMet-tyrosine kinase. **J Biol Chem**, v. 282, n. 23, p. 16764-16775, Jun 2007.

LIANG, H.; O'REILLY, S.; LIU, Y.; ABOUNADER, R.; LATERRA, J.; MAHER, V. M.; MCCORMICK, J. J. Sp1 regulates expression of MET, and ribozyme-induced down-regulation of MET in fibrosarcoma-derived human cells reduces or eliminates their tumorigenicity. **Int J Oncol**, v. 24, n. 5, p. 1057-1067, May 2004.

LIPINSKI, M. M.; JACKS, T. The retinoblastoma gene family in differentiation and development. **Oncogene**, v. 18, n. 55, p. 7873-7882, Dec 1999.

LIU, W. D.; ZHANG, T.; WANG, C. L.; MENG, H. M.; SONG, Y. W.; ZHAO, Z.; LI, Z. M.; LIU, J. K.; PAN, S. H.; WANG, W. B. Sphere-forming tumor cells possess stem-like properties in human fibrosarcoma primary tumors and cell lines. **Oncol Lett**, v. 4, n. 6, p. 1315-1320, Dec 2012.

LIU, Y. J.; XU, Y.; YU, Q. Full-length ADAMTS-1 and the ADAMTS-1 fragments display pro- and antimetastatic activity, respectively. **Oncogene**, v. 25, n. 17, p. 2452-2467, Apr 2006.

LODISH, H.; BERK, A.; ZIPURSKY, S. L.; MATSUDAIRA, P.; BALTIMORE, D.; DARNELL, J. Section 24.2, Proto-Oncogenes and Tumor-Suppressor Genes. In: COMPANY, W. H. F. A. (Ed.). **Molecular Cell Biology**. New York, 2000.

LU, P.; TAKAI, K.; WEAVER, V. M.; WERB, Z. Extracellular matrix degradation and remodeling in development and disease. **Cold Spring Harb Perspect Biol**, v. 3, n. 12, Dec 2011.

LU, P.; WEAVER, V. M.; WERB, Z. The extracellular matrix: a dynamic niche in cancer progression. **J Cell Biol**, v. 196, n. 4, p. 395-406, Feb 2012.

LU, X.; WANG, Q.; HU, G.; VAN POZNAK, C.; FLEISHER, M.; REISS, M.; MASSAGUÉ, J.; KANG, Y. ADAMTS1 and MMP1 proteolytically engage EGF-like ligands in an osteolytic signaling cascade for bone metastasis. **Genes Dev**, v. 23, n. 16, p. 1882-1894, Aug 2009.

LUQUE, A.; CARPIZO, D. R.; IRUELA-ARISPE, M. L. ADAMTS1/METH1 inhibits endothelial cell proliferation by direct binding and sequestration of VEGF165. **J Biol Chem**, v. 278, n. 26, p. 23656-23665, Jun 2003.

LYNCH, C. C.; MATRISIAN, L. M. Matrix metalloproteinases in tumor-host cell communication. **Differentiation**, v. 70, n. 9-10, p. 561-573, Dec 2002.

MARGOSIO, B.; MARCHETTI, D.; VERGANI, V.; GIAVAZZI, R.; RUSNATI, M.; PRESTA, M.; TARABOLETTI, G. Thrombospondin 1 as a scavenger for matrix-associated fibroblast growth factor 2. **Blood**, v. 102, n. 13, p. 4399-4406, Dec 2003.

MARKOWITZ, S. D.; ROBERTS, A. B. Tumor suppressor activity of the TGF-beta pathway in human cancers. **Cytokine Growth Factor Rev**, v. 7, n. 1, p. 93-102, Jun 1996.

MASSAGUÉ, J. TGF-beta signal transduction. **Annu Rev Biochem**, v. 67, p. 753-791, 1998.

MASSAGUÉ, J.; ATTISANO, L.; WRANA, J. L. The TGF-beta family and its composite receptors. **Trends Cell Biol**, v. 4, n. 5, p. 172-178, May 1994.

MASUI, T.; HOSOTANI, R.; TSUJI, S.; MIYAMOTO, Y.; YASUDA, S.; IDA, J.; NAKAJIMA, S.; KAWAGUCHI, M.; KOBAYASHI, H.; KOIZUMI, M.; TOYODA, E.; TULACHAN, S.; ARII, S.; DOI, R.; IMAMURA, M. Expression of METH-1 and METH-2 in pancreatic cancer. **Clin Cancer Res**, v. 7, n. 11, p. 3437-3443, Nov 2001.

MATSUMOTO, K.; NAKAMURA, T. Hepatocyte growth factor and the Met system as a mediator of tumor-stromal interactions. **Int J Cancer**, v. 119, n. 3, p. 477-483, Aug 2006.

MATSUMOTO, K.; OKAZAKI, H.; NAKAMURA, T. Novel function of prostaglandins as inducers of gene expression of HGF and putative mediators of tissue regeneration. **J Biochem**, v. 117, n. 2, p. 458-464, Feb 1995.

MAULIK, G.; MADHIWALA, P.; BROOKS, S.; MA, P. C.; KIJIMA, T.; TIBALDI, E. V.; SCHAEFER, E.; PARMAR, K.; SALGIA, R. Activated c-Met signals through PI3K with dramatic effects on cytoskeletal functions in small cell lung cancer. **J Cell Mol Med**, v. 6, n. 4, p. 539-553, 2002 Oct-Dec 2002.

MBEUNKUI, F.; JOHANN, D. J. Cancer and the tumor microenvironment: a review of an essential relationship. **Cancer Chemother Pharmacol**, v. 63, n. 4, p. 571-582, Mar 2009.

MINN, A. J.; KANG, Y.; SERGANOVA, I.; GUPTA, G. P.; GIRI, D. D.; DOUBROVIN, M.; PONOMAREV, V.; GERALD, W. L.; BLASBERG, R.; MASSAGUÉ, J. Distinct organ-specific metastatic potential of individual breast cancer cells and primary tumors. **J Clin Invest**, v. 115, n. 1, p. 44-55, Jan 2005.

MINOBE, K.; ONO, R.; MATSUMINE, A.; SHIBATA-MINOSHIMA, F.; IZAWA, K.; OKI, T.; KITAURA, J.; IINO, T.; TAKITA, J.; IWAMOTO, S.; HORI, H.; KOMADA, Y.; UCHIDA, A.; HAYASHI, Y.; KITAMURA, T.; NOSAKA, T. Expression of ADAMTS4 in Ewing's sarcoma. **Int J Oncol**, v. 37, n. 3, p. 569-581, Sep 2010.

MORRISON, B. A. Soft tissue sarcomas of the extremities. **Proc (Bayl Univ Med Cent)**, v. 16, n. 3, p. 285-290, Jul 2003.

MOUSTAKAS, A.; HELDIN, C. H. Non-Smad TGF-beta signals. **J Cell Sci**, v. 118, n. Pt 16, p. 3573-3584, Aug 2005.

MUELLER, M. M.; FUSENIG, N. E. Friends or foes - bipolar effects of the tumour stroma in cancer. **Nat Rev Cancer**, v. 4, n. 11, p. 839-849, Nov 2004.

MUNGER, J. S.; HARPEL, J. G.; GLEIZES, P. E.; MAZZIERI, R.; NUNES, I.; RIFKIN, D. B. Latent transforming growth factor-beta: structural features and mechanisms of activation. **Kidney Int**, v. 51, n. 5, p. 1376-1382, May 1997.

MUNGER, J. S.; HUANG, X.; KAWAKATSU, H.; GRIFFITHS, M. J.; DALTON, S. L.; WU, J.; PITTET, J. F.; KAMINSKI, N.; GARAT, C.; MATTHAY, M. A.; RIFKIN, D. B.; SHEPPARD, D. The integrin alpha v beta 6 binds and activates latent TGF beta 1: a mechanism for regulating pulmonary inflammation and fibrosis. **Cell**, v. 96, n. 3, p. 319-328, Feb 1999.

NABESHIMA, K.; INOUE, T.; SHIMAO, Y.; SAMESHIMA, T. Matrix metalloproteinases in tumor invasion: role for cell migration. **Pathol Int**, v. 52, n. 4, p. 255-264, Apr 2002.

NAKAMURA, M.; SONE, S.; TAKAHASHI, I.; MIZOGUCHI, I.; ECHIGO, S.; SASANO, Y. Expression of versican and ADAMTS1, 4, and 5 during bone development in the rat mandible and hind limb. **J Histochem Cytochem**, v. 53, n. 12, p. 1553-1562, Dec 2005.

NAKAMURA, T.; MATSUMOTO, K.; KIRITOSHI, A.; TANO, Y. Induction of hepatocyte growth factor in fibroblasts by tumor-derived factors affects invasive growth of tumor cells: in vitro analysis of tumor-stromal interactions. **Cancer Res**, v. 57, n. 15, p. 3305-3313, Aug 1997.

NALDINI, L.; VIGNA, E.; NARSIMHAN, R. P.; GAUDINO, G.; ZARNEGAR, R.; MICHALOPOULOS, G. K.; COMOGLIO, P. M. Hepatocyte growth factor (HGF) stimulates the tyrosine kinase activity of the receptor encoded by the proto-oncogene c-MET. **Oncogene**, v. 6, n. 4, p. 501-504, Apr 1991.

NIKITOVIC, D.; KOUVIDI, K.; KARAMANOS, N. K.; TZANAKAKIS, G. N. The roles of hyaluronan/RHAMM/CD44 and their respective interactions along the insidious pathways of fibrosarcoma progression. **Biomed Res Int**, v. 2013, p. 929531, 2013.

NORTON, W. H.; LEDIN, J.; GRANDEL, H.; NEUMANN, C. J. HSPG synthesis by zebrafish Ext2 and Extl3 is required for Fgf10 signalling during limb development. **Development**, v. 132, n. 22, p. 4963-4973, Nov 2005.

ORGAN, S. L.; TSAO, M. S. An overview of the c-MET signaling pathway. **Ther Adv Med Oncol**, v. 3, n. 1 Suppl, p. S7-S19, Nov 2011.

OVERALL, C. M. Molecular determinants of metalloproteinase substrate specificity: matrix metalloproteinase substrate binding domains, modules, and exosites. **Mol Biotechnol**, v. 22, n. 1, p. 51-86, Sep 2002.

OVERALL, C. M.; BLOBEL, C. P. In search of partners: linking extracellular proteases to substrates. **Nat Rev Mol Cell Biol**, v. 8, n. 3, p. 245-257, Mar 2007.

OZBEK, S.; BALASUBRAMANIAN, P. G.; CHIQUET-EHRISMANN, R.; TUCKER, R. P.; ADAMS, J. C. The evolution of extracellular matrix. **Mol Biol Cell**, v. 21, n. 24, p. 4300-4305, Dec 2010.

PAGE-MCCAW, A.; EWALD, A. J.; WERB, Z. Matrix metalloproteinases and the regulation of tissue remodelling. **Nat Rev Mol Cell Biol**, v. 8, n. 3, p. 221-233, Mar 2007.

PARAMESWARAN, K.; WILLEMS-WIDYASTUTI, A.; ALAGAPPAN, V. K.; RADFORD, K.; KRANENBURG, A. R.; SHARMA, H. S. Role of extracellular matrix and its regulators in human airway smooth muscle biology. **Cell Biochem Biophys**, v. 44, n. 1, p. 139-146, 2006.

PEEK, R. M.; MOHLA, S.; DUBOIS, R. N. Inflammation in the genesis and perpetuation of cancer: summary and recommendations from a national cancer institute-sponsored meeting. **Cancer Res**, v. 65, n. 19, p. 8583-8586, Oct 2005.

PETRELLI, A.; GILESTRO, G. F.; LANZARDO, S.; COMOGLIO, P. M.; MIGONE, N.; GIORDANO, S. The endophilin-CIN85-Cbl complex mediates ligand-dependent downregulation of c-Met. **Nature**, v. 416, n. 6877, p. 187-190, Mar 2002.

PETRIE, R. J.; GAVARA, N.; CHADWICK, R. S.; YAMADA, K. M. Nonpolarized signaling reveals two distinct modes of 3D cell migration. **J Cell Biol**, v. 197, n. 3, p. 439-455, Apr 2012.

PIEK, E.; HELDIN, C. H.; TEN DIJKE, P. Specificity, diversity, and regulation in TGF-beta superfamily signaling. **FASEB J**, v. 13, n. 15, p. 2105-2124, Dec 1999.

PORTER, S.; CLARK, I. M.; KEVORKIAN, L.; EDWARDS, D. R. The ADAMTS metalloproteinases. **Biochem J**, v. 386, n. Pt 1, p. 15-27, Feb 2005.

PORTER, S.; SCOTT, S. D.; SASSOON, E. M.; WILLIAMS, M. R.; JONES, J. L.; GIRLING, A. C.; BALL, R. Y.; EDWARDS, D. R. Dysregulated expression of adamalysin-thrombospondin genes in human breast carcinoma. **Clin Cancer Res**, v. 10, n. 7, p. 2429-2440, Apr 2004.

PRASAD, M.; MONTELL, D. J. Cellular and molecular mechanisms of border cell migration analyzed using time-lapse live-cell imaging. **Dev Cell**, v. 12, n. 6, p. 997-1005, Jun 2007.

PRIMAKOFF, P.; MYLES, D. G. The ADAM gene family: surface proteins with adhesion and protease activity. **Trends Genet**, v. 16, n. 2, p. 83-87, Feb 2000.

RA, H. J.; PARKS, W. C. Control of matrix metalloproteinase catalytic activity. **Matrix Biol**, v. 26, n. 8, p. 587-596, Oct 2007.

RAIBORG, C.; RUSTEN, T. E.; STENMARK, H. Protein sorting into multivesicular endosomes. **Curr Opin Cell Biol**, v. 15, n. 4, p. 446-455, Aug 2003.

REHN, A. P.; BIRCH, M. A.; KARLSTRÖM, E.; WENDEL, M.; LIND, T. ADAMTS-1 increases the three-dimensional growth of osteoblasts through type I collagen processing. **Bone**, v. 41, n. 2, p. 231-238, Aug 2007.

REISS, K.; SAFTIG, P. The "a disintegrin and metalloprotease" (ADAM) family of sheddases: physiological and cellular functions. **Semin Cell Dev Biol**, v. 20, n. 2, p. 126-137, Apr 2009.

REMACLE, A.; MURPHY, G.; ROGHI, C. Membrane type I-matrix metalloproteinase (MT1-MMP) is internalised by two different pathways and is recycled to the cell surface. **J Cell Sci**, v. 116, n. Pt 19, p. 3905-3916, Oct 2003.

RICCIARDELLI, C.; FREWIN, K. M.; TAN, I. E. A.; WILLIAMS, E. D.; OPESKIN, K.; PRITCHARD, M. A.; INGMAN, W. V.; RUSSELL, D. L. The ADAMTS1 protease gene is required for mammary tumor growth and metastasis. **Am J Pathol**, v. 179, n. 6, p. 3075-3085, Dec 2011.

RIDGE, S. M.; SULLIVAN, F. J.; GLYNN, S. A. Mesenchymal stem cells: key players in cancer progression. **Mol Cancer**, v. 16, n. 1, p. 31, Feb 2017.

ROCKS, N.; PAULISSEN, G.; EL HOUR, M.; QUESADA, F.; CRAHAY, C.; GUEDERS, M.; FOIDART, J. M.; NOEL, A.; CATALDO, D. Emerging roles of ADAM and ADAMTS metalloproteinases in cancer. **Biochimie**, v. 90, n. 2, p. 369-379, Feb 2008.

RODRIGUES, G. A.; PARK, M. Autophosphorylation modulates the kinase activity and oncogenic potential of the Met receptor tyrosine kinase. **Oncogene**, v. 9, n. 7, p. 2019-2027, Jul 1994.

RODRIGUEZ-MANZANEQUE, J. C.; MILCHANOWSKI, A. B.; DUFOUR, E. K.; LEDUC, R.; IRUELA-ARISPE, M. L. Characterization of METH-1/ADAMTS1 processing reveals two distinct active forms. **J Biol Chem**, v. 275, n. 43, p. 33471-33479, Oct 2000.

ROWE, R. G.; WEISS, S. J. Navigating ECM barriers at the invasive front: the cancer cell-stroma interface. **Annu Rev Cell Dev Biol**, v. 25, p. 567-595, 2009.

ROZANOV, D. V.; DERYUGINA, E. I.; MONOSOV, E. Z.; MARCHENKO, N. D.; STRONGIN, A. Y. Aberrant, persistent inclusion into lipid rafts limits the tumorigenic function of membrane type-1 matrix metalloproteinase in malignant cells. **Exp Cell Res**, v. 293, n. 1, p. 81-95, Feb 2004.

RUOSLAHTI, E.; REED, J. C. Anchorage dependence, integrins, and apoptosis. **Cell**, v. 77, n. 4, p. 477-478, May 1994.

RUSSELL, D. L.; DOYLE, K. M.; OCHSNER, S. A.; SANDY, J. D.; RICHARDS, J. S. Processing and localization of ADAMTS-1 and proteolytic cleavage of versican during cumulus matrix expansion and ovulation. **J Biol Chem**, v. 278, n. 43, p. 42330-42339, Oct 2003.

SAHAI, E.; MARSHALL, C. J. Differing modes of tumour cell invasion have distinct requirements for Rho/ROCK signalling and extracellular proteolysis. **Nat Cell Biol**, v. 5, n. 8, p. 711-719, Aug 2003.

SAM, M. R.; ELLIOTT, B. E.; MUELLER, C. R. A novel activating role of SRC and STAT3 on HGF transcription in human breast cancer cells. **Mol Cancer**, v. 6, p. 69, Oct 2007.

SANDY, J. D.; WESTLING, J.; KENAGY, R. D.; IRUELA-ARISPE, M. L.; VERSCHAREN, C.; RODRIGUEZ-MAZANEQUE, J. C.; ZIMMERMANN, D. R.; LEMIRE, J. M.; FISCHER, J. W.; WIGHT, T. N.; CLOWES, A. W. Versican V1 proteolysis in human aorta in vivo occurs at the Glu441-Ala442 bond, a site that is cleaved by recombinant ADAMTS-1 and ADAMTS-4. **J Biol Chem**, v. 276, n. 16, p. 13372-13378, Apr 2001.

SHARMA, M.; SAH, P.; SHARMA, S. S.; RADHAKRISHNAN, R. Molecular changes in invasive front of oral cancer. **J Oral Maxillofac Pathol**, v. 17, n. 2, p. 240-247, May 2013.

SIERRA, J. R.; TSAO, M. S. c-MET as a potential therapeutic target and biomarker in cancer. **Ther Adv Med Oncol**, v. 3, n. 1 Suppl, p. S21-35, Nov 2011.

SMIRNOV, A. V. [Fibrosarcoma: immunohistochemical study of the extracellular matrix]. **Arkh Patol**, v. 50, n. 12, p. 17-24, 1988.

SOMANNA, A.; MUNDODI, V.; GEDAMU, L. Functional analysis of cathepsin B-like cysteine proteases from *Leishmania donovani* complex. Evidence for the activation of latent transforming growth factor beta. **J Biol Chem**, v. 277, n. 28, p. 25305-25312, Jul 2002.

STAMENKOVIC, I. Matrix metalloproteinases in tumor invasion and metastasis. **Semin Cancer Biol**, v. 10, n. 6, p. 415-433, Dec 2000.

STANTON, H.; MELROSE, J.; LITTLE, C. B.; FOSANG, A. J. Proteoglycan degradation by the ADAMTS family of proteinases. **Biochim Biophys Acta**, v. 1812, n. 12, p. 1616-1629, Dec 2011.

STEFFAN, J. J.; COLEMAN, D. T.; CARDELLI, J. A. The HGF-met signaling axis: emerging themes and targets of inhibition. **Curr Protein Pept Sci**, v. 12, n. 1, p. 12-22, Feb 2011.

STERNLICHT, M. D.; SUNNARBORG, S. W.; KOUROS-MEHR, H.; YU, Y.; LEE, D. C.; WERB, Z. Mammary ductal morphogenesis requires paracrine activation of stromal EGFR via ADAM17-dependent shedding of epithelial amphiregulin. **Development**, v. 132, n. 17, p. 3923-3933, Sep 2005.

STERNLICHT, M. D.; WERB, Z. How matrix metalloproteinases regulate cell behavior. **Annu Rev Cell Dev Biol**, v. 17, p. 463-516, 2001.

SUGAWARA, J.; FUKAYA, T.; MURAKAMI, T.; YOSHIDA, H.; YAJIMA, A. Hepatocyte growth factor stimulated proliferation, migration, and lumen formation of human endometrial epithelial cells in vitro. **Biol Reprod**, v. 57, n. 4, p. 936-942, Oct 1997.

SWARTZ, M. A.; IIDA, N.; ROBERTS, E. W.; SANGALETTI, S.; WONG, M. H.; YULL, F. E.; COUSSENS, L. M.; DECLERCK, Y. A. Tumor microenvironment complexity: emerging roles in cancer therapy. **Cancer Res**, v. 72, n. 10, p. 2473-2480, May 2012.

TAN, I. E. A.; RICCIARDELLI, C.; RUSSELL, D. L. The metalloproteinase ADAMTS1: a comprehensive review of its role in tumorigenic and metastatic pathways. **Int J Cancer**, v. 133, n. 10, p. 2263-2276, Nov 2013.

TEN DIJKE, P.; HILL, C. S. New insights into TGF-beta-Smad signalling. **Trends Biochem Sci**, v. 29, n. 5, p. 265-273, May 2004.

THAI, S. N.; IRUELA-ARISPE, M. L. Expression of ADAMTS1 during murine development. **Mech Dev**, v. 115, n. 1-2, p. 181-185, Jul 2002.

THIAGALINGAM, S. A cascade of modules of a network defines cancer progression. **Cancer Res**, v. 66, n. 15, p. 7379-7385, Aug 2006.

TORTORELLA, M. D.; MALFAIT, F.; BARVE, R. A.; SHIEH, H. S.; MALFAIT, A. M. A review of the ADAMTS family, pharmaceutical targets of the future. **Curr Pharm Des**, v. 15, n. 20, p. 2359-2374, 2009.

TRUSOLINO, L.; BERTOTTI, A.; COMOGLIO, P. M. MET signalling: principles and functions in development, organ regeneration and cancer. **Nat Rev Mol Cell Biol**, v. 11, n. 12, p. 834-848, Dec 2010.

TRUSOLINO, L.; COMOGLIO, P. M. Scatter-factor and semaphorin receptors: cell signalling for invasive growth. **Nat Rev Cancer**, v. 2, n. 4, p. 289-300, Apr 2002.

TUCK, A. B.; PARK, M.; STERNS, E. E.; BOAG, A.; ELLIOTT, B. E. Coexpression of hepatocyte growth factor and receptor (Met) in human breast carcinoma. **Am J Pathol**, v. 148, n. 1, p. 225-232, Jan 1996.

UCHIDA, D.; KAWAMATA, H.; OMOTEHARA, F.; NAKASHIRO KI; KIMURA-YANAGAWA, T.; HINO, S.; BEGUM, N. M.; HOQUE, M. O.; YOSHIDA, H.; SATO, M.; FUJIMORI, T. Role of HGF/c-met system in invasion and metastasis of oral squamous cell carcinoma cells in vitro and its clinical significance. **Int J Cancer**, v. 93, n. 4, p. 489-496, Aug 2001.

VALVERDE, J.; VINAGREIRO, M.; GOUVEIA, P.; KOCH, P.; SOARES, V.; GOMES, T. Sarcoma the great "masquerader" hematoma/deep vein thrombosis manifestation. **Int J Surg Case Rep**, v. 28, p. 348-351, 2016.

VISSE, R.; NAGASE, H. Matrix metalloproteinases and tissue inhibitors of metalloproteinases: structure, function, and biochemistry. **Circ Res**, v. 92, n. 8, p. 827-839, May 2003.

VOLPERT, O. V.; LAWLER, J.; BOUCK, N. P. A human fibrosarcoma inhibits systemic angiogenesis and the growth of experimental metastases via thrombospondin-1. **Proc Natl Acad Sci U S A**, v. 95, n. 11, p. 6343-6348, May 1998.

WEIDNER, K. M.; SACHS, M.; RIETHMACHER, D.; BIRCHMEIER, W. Mutation of juxtamembrane tyrosine residue 1001 suppresses loss-of-function mutations of the met receptor in epithelial cells. **Proc Natl Acad Sci U S A**, v. 92, n. 7, p. 2597-2601, Mar 1995.

WEIS, S. M.; CHERESH, D. A. Tumor angiogenesis: molecular pathways and therapeutic targets. **Nat Med**, v. 17, n. 11, p. 1359-1370, Nov 2011.

WELCH, M. D. Cell migration, freshly squeezed. **Cell**, v. 160, n. 4, p. 581-582, Feb 2015.

WIBMER, C.; LEITHNER, A.; ZIELONKE, N.; SPERL, M.; WINDHAGER, R. Increasing incidence rates of soft tissue sarcomas? A population-based epidemiologic study and literature review. **Ann Oncol**, v. 21, n. 5, p. 1106-1111, May 2010.

WOLF, K.; MAZO, I.; LEUNG, H.; ENGELKE, K.; VON ANDRIAN, U. H.; DERYUGINA, E. I.; STRONGIN, A. Y.; BRÖCKER, E. B.; FRIEDL, P. Compensation mechanism in tumor cell migration: mesenchymal-amoeboid transition after blocking of pericellular proteolysis. **J Cell Biol**, v. 160, n. 2, p. 267-277, Jan 2003.

WOLF, K.; TE LINDERT, M.; KRAUSE, M.; ALEXANDER, S.; TE RIET, J.; WILLIS, A. L.; HOFFMAN, R. M.; FIGDOR, C. G.; WEISS, S. J.; FRIEDL, P. Physical limits of cell migration: control by ECM space and nuclear deformation and tuning by proteolysis and traction force. **J Cell Biol**, v. 201, n. 7, p. 1069-1084, Jun 2013.

WOLFSBERG, T. G.; PRIMAKOFF, P.; MYLES, D. G.; WHITE, J. M. ADAM, a novel family of membrane proteins containing A Disintegrin And Metalloprotease domain: multipotential functions in cell-cell and cell-matrix interactions. **J Cell Biol**, v. 131, n. 2, p. 275-278, Oct 1995.

YANG, C. Y.; CHANALARIS, A.; TROEBERG, L. ADAMTS and ADAM metalloproteinases in osteoarthritis - looking beyond the 'usual suspects'. **Osteoarthritis Cartilage**, Feb 2017.

YOSHINAGA, Y.; MATSUNO, Y.; FUJITA, S.; NAKAMURA, T.; KIKUCHI, M.; SHIMOSATO, Y.; HIROHASHI, S. Immunohistochemical detection of hepatocyte growth factor/scatter factor in human cancerous and inflammatory lesions of various organs. **Jpn J Cancer Res**, v. 84, n. 11, p. 1150-1158, Nov 1993.

ZETTER, B. R. Angiogenesis and tumor metastasis. **Annu Rev Med**, v. 49, p. 407-424, 1998.

ZHANG, X. P.; KAMATA, T.; YOKOYAMA, K.; PUZON-MCLAUGHLIN, W.; TAKADA, Y. Specific interaction of the recombinant disintegrin-like domain of MDC-15 (metargidin, ADAM-15) with integrin alphavbeta3. **J Biol Chem**, v. 273, n. 13, p. 7345-7350, Mar 1998.

ZHANG, Y. W.; WANG, L. M.; JOVE, R.; VANDE WOUDE, G. F. Requirement of Stat3 signaling for HGF/SF-Met mediated tumorigenesis. **Oncogene**, v. 21, n. 2, p. 217-226, Jan 2002.

ZLOTNIK, A.; YOSHIE, O. Chemokines: a new classification system and their role in immunity. **Immunity**, v. 12, n. 2, p. 121-127, Feb 2000.