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**EFEITOS DO CONSUMO DE FRUTOSE DURANTE A
GESTAÇÃO E LACTAÇÃO E SUA REPERCUSSÃO NA VIDA
PÓS-NATAL: ESTUDO DE PROGRAMAÇÃO METABÓLICA
EM RATOS MACHOS**

Tese apresentada ao Programa de Pós-Graduação em Ciências Morfofuncionais do Instituto de Ciências Biomédicas da Universidade de São Paulo, para obtenção do Título de Doutor em Ciências Morfofuncionais.

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RESUMO

SILVA, R. J. **Efeitos do consumo de frutose durante a gestação e lactação e sua repercussão na vida pós-natal: estudo de programação metabólica em ratos machos.** 175 f. Tese (Doutorado em Ciências Morfofuncionais) - Instituto de Ciências Biomédicas, Universidade de São Paulo, São Paulo, 2017.

INTRODUÇÃO: A exposição materna à frutose durante a gestação e lactação pode contribuir para o desenvolvimento de doenças crônicas na vida adulta da prole, mediado por alterações periféricas relacionadas ao metabolismo hepático glicídico e lipídico, e na via de sinalização de insulina, podendo provocar alterações centrais relacionadas à via de sinalização da leptina e de neuropeptídeos envolvidos no controle do comportamento alimentar. **OBJETIVO:** Estudar os efeitos da ingestão de frutose durante a gestação e lactação sobre a vida pós-natal no que diz respeito às alterações metabólicas, de expressão gênica e ao comportamento alimentar em ratos machos. **METODOLOGIA:** Foram utilizados ratos machos da linhagem Sprague-Dawley, sacrificados nos 14^o e 90^o dias de vida pós-natal completos, provenientes de mães alimentadas com ração controle (AIN-93) e de mães alimentadas com ração rica em frutose (60%), ambas isocalóricas, durante (i) a gestação; (ii) a lactação e (iii) gestação e lactação. Após o desmame, a prole foi alimentada com ração controle (AIN-93) até os 91 dias de vida. Foram analisados os seguintes parâmetros: biométricos, dietéticos e metabólicos (glicemia, concentração sérica de leptina, de insulina) e de neuropeptídeos envolvidos no controle do comportamento alimentar (AgRP, CART e MCH), sensibilidade periférica à ação da insulina, expressão de proteínas envolvidas com a via de sinalização da insulina e deposição hepática de lipídios. **RESULTADOS:** Frente a dieta materna rica em frutose na gestação e na lactação, houve redução do índice de Lee no nascimento e hipoleptinemia aos 90 dias de vida (apenas nas proles que as mães consumiram frutose na gestação), intolerância à glicose na vida pós-natal desde o desmame, concomitante com alterações na sensibilidade à ação da insulina na vida adulta, hiperinsulinemia, dislipidemia (elevação da concentração de TG e de LDL-colesterol e redução da concentração de HDL-colesterol), deposição hepática de TG, estímulo hedônico do comportamento alimentar alterado aos 14 dias de vida pós-natal via núcleo *accumbens shell* com maior expressão gênica de CART. **CONCLUSÃO:** Proles provenientes de mães que consumiram ração rica em frutose durante a gestação e/ou lactação, possuem possível marca epigenética que pode contribuir para o prejuízo na produção e ação da insulina, ocasionando alterações na homeostase do metabolismo glicídico e lipídico hepáticos, levando a deposição hepática de gordura, alterações no perfil lipídico e no controle hedônico central do comportamento alimentar.

Palavras-chave: Programação metabólica. Frutose. Risco cardiometabólico. Comportamento alimentar. Genômica nutricional.

ABSTRACT

SILVA, R. J. **Effects of fructose consumption during gestation and lactation and its repercussion on postnatal life: a metabolic programming study in male rats.** 175 p. Thesis (Ph.D. in Morphofunctional Sciences) - Institute of Biomedical Sciences, University of São Paulo, São Paulo, 2017.

INTRODUCTION: Maternal exposure to fructose during gestation and lactation can contribute to the development of chronic diseases in the adult life of offspring, mediated by peripheral alterations related to glucose and lipid metabolism of the liver, and in the insulin signaling pathway, which may lead to central alterations related to the signaling pathway of leptin and neuropeptides involved in the control of eating behavior. **OBJECTIVE:** To study the effects of ingestion of fructose during gestation and lactation on postnatal life with respect to metabolic changes, gene expression and feeding behavior in male rats. **METHODS:** Male Sprague-Dawley rats were sacrificed on the 14th and 90th days of postnatal life, from mothers fed with control rat chow (AIN-93) and from mothers fed a high fructose diet (60%). Both isocaloric, during (i) gestation; (ii) lactation and (iii) gestation and lactation. After weaning, the offspring were fed with control rat chow (AIN-93) until the 91st days of life. The following parameters were analyzed: biometric, dietary, metabolic (glycemia, serum concentration of leptin, insulin) and neuropeptides involved in food behavior control (AgRP, CART and MCH), peripheral sensitivity to insulin action, expression of proteins involved with the insulin signaling pathway and hepatic deposition of lipid. **RESULTS:** A maternal diet high in fructose during gestation and lactation had a reduction in the Lee index at birth and hypoleptinemia at 90 days of age (only in the offspring that the mothers consumed fructose during pregnancy), glucose intolerance in postnatal life since with changes in insulin sensitivity in adult life, hyperinsulinemia, dyslipidemia (elevation of TG and LDL-cholesterol, reduction of HDL-cholesterol), hepatic deposition of TG, and in the central hedonic control of dietary behavior altered at 14th days of postnatal life via the nucleus accumbens shell with greater gene expression of CART. **CONCLUSION:** The offspring from mothers who consumed fructose rat chow during gestation and / or lactation have a possible epigenetic mark that may contribute to the impairment in the production and action of insulin, leading to alterations in hepatic glucose and lipid metabolism homeostasis, leading to hepatic fat deposition and alterations in the lipid profile, and in the central hedonic control of dietary behavior.

Keywords: Metabolic programming. Fructose. Cardiometabolic risk. Food behavior. Nutritional genomics.

1 INTRODUÇÃO

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A ocidentalização da alimentação é um dos fatores da transição nutricional que ocorre em todo o mundo, com aumento da disponibilidade de alimentos de elevado teor calórico e ricos em frutose, como refrigerantes e alimentos pré-preparados (processados e ultraprocessados), que contribuem para o aumento da prevalência de síndrome metabólica (SM) (SILVA et al., 2011; MINISTÉRIO DA SAÚDE, 2014).

Evidências demonstram que doenças crônicas não transmissíveis (DCNT), como diabetes melito tipo 2 (DM 2), hipertensão arterial sistêmica (HAS), aterosclerose e doenças cardiovasculares (DCV), podem ter origem em alterações ocorridas no útero e/ou durante o início da vida pós-natal (JOSHI et al., 2003).

A maioria dos estudos sobre programação metabólica é realizada com modelos animais que induzem subnutrição materna, ou por meio do consumo de ração hiperlipídica pelas mães, o que aumenta o risco de obesidade, resistência à ação da insulina (RI) e diminuição da sensibilidade à leptina pela prole, independentemente do tipo de ração ingerida pós-parto (HOWIE et al., 2009; VICKERS, 2011a). No entanto, há escassez de estudos utilizando a frutose e seu impacto na programação metabólica. Neste contexto, os efeitos da ingestão materna de frutose sobre a saúde da prole ainda são pouco conhecidos, apesar do recente aumento do consumo de bebidas e alimentos com alto teor de frutose (ALZAMENDI et al., 2016; VICKERS, 2011a).

Tendo em vista que a SM é uma condição clínica que vem tomando proporções epidêmicas nos últimos anos, mostra-se de suma importância o estudo das consequências do consumo de frutose desde a sua exposição precoce.

Frente ao exposto, este trabalho busca entender melhor os mecanismos envolvidos no desenvolvimento da SM, ocasionado pelo consumo materno de frutose, uma vez que, a prole pode sofrer alterações irreversíveis tanto metabólicas, como no e no comportamento alimentar, podendo culminar no desenvolvimento de DCNT na vida adulta.

1.1 Justificativa

Tendo em vista que a SM é uma doença que vem tomando proporções epidêmicas nos últimos anos, mostra-se de suma importância o estudo das consequências do consumo de frutose desde a sua exposição no período perinatal.

A maior parte dos nossos conhecimentos sobre a ação de nutrientes e compostos bioativos dos alimentos sobre a programação metabólica é oriundo de estudos com restrição alimentar, restrição proteica, ou dietas ricas em lipídios totais ou saturados.

Dessa forma, na busca de um modelo experimental adequado ao estudo da SM/RCM, o presente trabalho foi delineado para analisar a tolerância à glicose, a sensibilidade à insulina, perfil lipídico e alterações neuroendócrinas em ratos descendentes de mães mantidas com ração rica em frutose nos períodos gestacional e lactacional, uma vez que, a frutose tem sido um nutriente associado ao desenvolvimento dos fatores de risco associado.

1.2 Hipótese

A exposição materna à frutose durante a gestação e lactação contribui para o desenvolvimento de DCNT na vida adulta da prole, mediado por alterações periféricas relacionadas ao metabolismo hepático glicídico e lipídico e na via de sinalização de insulina, bem como por meio de alterações centrais relacionadas a via de sinalização de leptina e de neuropeptídeos envolvidos no controle do comportamento alimentar.

7 CONCLUSÃO

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Frente aos objetivos propostos e aos resultados obtidos, podemos concluir que a ingestão de frutose pelas mães durante os períodos críticos de desenvolvimento fetal e infantil favorece na descendência:

- ✓ Redução do índice de Lee (peso/comprimento) no nascimento apenas nas proles que as mães consumiram frutose na gestação;
- ✓ Intolerância à glicose na vida pós-natal desde o desmame, concomitante com alterações na sensibilidade à ação da insulina na vida adulta;
- ✓ Hiperinsulinemia;
- ✓ Hipoleptinemia apenas nas proles que as mães consumiram frutose na gestação;
- ✓ Dislipidemia (elevação de TG e LDL-colesterol, com redução do HDL-colesterol);
- ✓ Comprometimento hepático com deposição hepática de TG;
- ✓ Controle hedônico do comportamento alimentar alterado aos 14 dias de vida via núcleo AcbSh.

REFERÊNCIAS

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ADKINS, A. et al. Higher Insulin Concentrations Are Required to Suppress Gluconeogenesis Than Glycogenolysis in Nondiabetic Humans. **Diabetes**, v. 52, n. 9, p. 2213-2220, 2003.

ANDREOLLO, N. A. et al. Idade dos ratos versus idade humana: qual é a relação? **ABCD Arq Bras Cir Dig.**, v25, n. 1, p. 49-51, 2012.

AHIMA, R. S. et al. Role of leptin in the neuroendocrine response to fasting. **Nature**, v. 382, n. 6588, p. 250-252, 1996.

AHMED, K. T. et al. Liver diseases in pregnancy: Diseases unique to pregnancy. **World J. Gastroenterol.**, v. 21, n. 19 (43), p. 7639-7646, 2013.

ALZAMENDI, A. et al. High risk of metabolic and adipose tissue dysfunctions in adult male progeny, due to prenatal and adulthood malnutrition induced by fructose rich diet. **Nutrients**, v. 8, n. 3, p. 178-193, 2016.

ALZAMENDI, A. et al. Increased male offspring's risk of metabolic neuroendocrine dysfunction and overweight after fructose-rich diet intake by the lactating mother. **Endocrinology**, v. 151, n. 9, p. 4214–4223, 2010b.

ALZAMENDI, A. et al. Oral metformin treatment prevents enhanced insulin demand and placental dysfunction in the pregnant rat fed a fructose-rich diet. **ISRN Endocrinol.**, v. 2012: n. 757913, p. 1-8, 2012.

ALZAMENDI, A. et al. Parametrial adipose tissue and metabolic dysfunctions induced by fructose-rich diet in normal and neonatal-androgenized adult female rats. **Obesity (Silver Spring)**, v. 18, n. 3, p. 441-448, 2010a.

* De acordo com:

ASSOCIAÇÃO BRASILEIRA DE NORMAS TÉCNICAS. **NBR 6023**: informação e documentação: referências: elaboração. Rio de Janeiro, 2002.

ARAÚJO, E. P. et al. Short-term in vivo inhibition of insulin receptor substrate-1 expression leads to insulin resistance, hyperinsulinemia, and increased adiposity. **Endocrinology**, v. 146, n. 3, p. 1428-1437, 2005.

ARMITAGE, J. A.; TAYLOR, P. D.; POSTON, L. Experimental models of developmental programming: consequences of exposure to an energy rich diet during development. **J. Physiol.**, v. 565, n. Pt 1, p. 3-8, 2005.

ASARIAN, L.; BÄCHLER, T. Neuroendocrine control of satiation. **Horm. Mol. Biol. Clin. Investig.**, v. 19, n. 3, p. 163-192, 2014.

ASGHAR, Z. A. et al. Maternal fructose drives placental uric acid production leading to adverse fetal outcomes. **Sci. Rep.**, v. 6, n. 25091, p. 1-11, 2016.

BADO, A. et al. The stomach is a source of leptin. **Nature**, v. 395, p. 790-793, 1998.

BAHJAOUI-BOUHADDI, M. et al. Insulin treatment stimulates the rat melanin-concentrating hormone-producing neurons. **Neuropeptides**, v. 27, n. 4, p. 251-258, 1994.

BAINS, R. K. et al. Visceral obesity without insulin resistance in late-onset obesity rats. **Endocrinology**, v. 145, n. 6, p. 2666-2679, 2004.

BAKOS, G. Spectrophotometric Evaluation of Protein-Determination by the Biuret Method. **Orv. Hetil.**, v. 105, p. 1077-1079, 1964.

BALBUS, J. M. et al. Early-life prevention of non-communicable diseases. **Lancet**, v. 381, p. 3-4, 2013.

BALONAN, L. C.; SHENG, H. P. Perinatal feedings adversely affect lipogenic activities but not glucose handling in adult rats. **Pediatr. Res.**, v. 48, n. 5, p. 668-673, 2000.

BARKER, D. J. et al. Type 2 (non-insulin-dependent) diabetes mellitus, hypertension and hyperlipidemia (syndrome X): relation to reduced fetal growth. **Diabetologia**, v. 36, p. 62–67, 1993.

BARKER, D. J. et al., Fetal origins of adult disease: strength of effects and biological basis. **Int. J. Epidemiol.**, v. 31, p. 1235–1239, 2002.

BARREIROS, R. C; BOSSOLAN, G. TRINDADE, C. E. P. Frutose em humanos: efeitos metabólicos, utilização clínica e erros inatos associados. **Rev. Nutr.**, v. 18, n. 3, p. 377-389, 2005.

BART, B.; WIT, J. M. Catch-up growth. **Endocr. Ver.**, v. 18, p. 646–661, 1997.

BASARANOGU, M. et al. Fructose as a key player in the development of fatty liver disease. **World J. Gastroenterol.**, v. 19, n. 8, p. 1166-1172, 2013.

BASCIANO, H.; FEDERICO, L.; ADELI, K. Fructose, insulin resistance, and metabolic dyslipidemia. **Nutr. Metab. (Lond)**., v. 2, n. 1, p. 1-5, 2005.

BECK, B. Early and persistent up-regulation of hypothalamic orexigenic peptides in rat offspring born to dams fed a high-carbohydrate supplement during gestation. **Brain Res.**, v. 5, n. 1477, p. 10-18, 2012.

BERG, J. P. Leptin is a potent anti-diabetic in mice with lipodystrophy and insulin resistance. **Eur. J. Endocrinol.**, v. 142, n. 2, p. 114-116, 2000.

BERNARDIS, L. L.; PATTERSON, B. D. Correlation between 'Lee index' and carcass fat content in weanling and adult female rats with hypothalamic lesions. **J. Endocrinol.**, v. 40, p. 527-538, 1968.

BEZERRA, R. M. et al. A high-fructose diet induces insulin resistance but not blood pressure changes in normotensive rats. **Braz. J. Med. Biol. Res.**, v. 34, n. 9, p. 1155-1160, 2001.

BIRN, I. et al. The association between preoperative symptoms of obesity in knee and hip joints and the change in quality of life after laparoscopic Roux-en-y gastric bypass. **Obes Surg.**, v. 26, n. 5, p. 950-6, 2016.

BLACKMORE, H. L.; OZANNE, S. E. Maternal diet-induced obesity and offspring cardiovascular health. **J. Dev. Orig. Health Dis.**, v. 4, n. 5, p. 338-347, 2013.

BONORA, E. et al. Estimates of in vivo insulin action in man: comparison of insulin tolerance tests with euglycemic and hyperglycemic glucose clamp studies. **J Clin Endocrinol Metab.**, v. 68, n. 2, p. 374-378, 1989.

BOURET, S. G.; SIMERLY, R. B. Minireview: Leptin and development of hypothalamic feeding circuits. **Endocrinology**, v.145, n. 6, p. 2621-2626, 2004.

BOURET, S. G.; DRAPER, S. J.; SIMERLY, R. B. Formation of projection pathways from the arcuate nucleus of the hypothalamus to hypothalamic regions implicated in the neural control of feeding behavior in mice. **J. Neurosci.**, v. 17, n. 11, p. 2797-2805, 2004.

BRADFORD, M. M. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. **Anal. Biochem.**, v. 7, n. 72, p. 248-254.1976.

BRAY, G. A.; NIELSEN, S. J.; POPKIN, B. M. Consumption of high-fructose corn syrup in beverages may play a role in the epidemic of obesity. **Am. J. Clin. Nutr.**, v. 79, n. 4, p. 537-543, 2004.

BUCHANAN, T. A. et al. Insulin sensitivity and B-cell responsiveness to glucose during late pregnancy in lean and moderately obese women with normal glucose tolerance or mild gestational diabetes. **Am. J. Obstet. Gynecol.** v. 162, p.1008-1014, 1990.

CAMBRI, L. T. et al. Recovery of rat growth and lipid profiles in adult rats subjected to fetal protein malnutrition with a fructose-rich diet. **Nutr. Res.**, v. 30, p. 156–162, 2010.

CAMPBELL, W. J.; BRENDENUHL, J. H.; BAZER, F. W. Effect of fructose consumption during lactation on sow and litter performance and sow plasma constituents. **J. Anim. Sci.**, v. 68, p. 1378–1388, 1990.

CARRETERO, I. B. et al. Leptina: implicaciones fisiológicas y clínicas. **Anales Medicina Interna**, v.18, n.3, p.152-160, 2001.

CARVALHEIRA, J. B. et al. Insulin modulates leptin-induced STAT-3 activation in rat hypothalamus. **FEBS. Lett.**, v. 500, p. 119-124, 2001.

CASTELLANO, C.; OLIVERIO, A. Early malnutrition and postnatal changes in brain and behavior in the mouse. **Brain. Res.**, v. 101, n. 2, p. 317-325, 1976.

CASTELLO, A. et al. Regulation of GLUT5 gene expression in rat intestinal mucosa: Regional distribution, circadian rhythm, perinatal development and effect of diabetes. **Biochem. J.**, v. 309, p. 271-277, 1995.

CESARETTI, M. L.; KOHLMANN, JUNIOR O. Experimental models of insulin resistance and obesity: lessons learned. **Arq. Bras. Endocrinol. Metabol.**, v. 50, n. 2, p. 190-197, 2006.

CHA, S. H. et al. Differential effects of central fructose and glucose on hypothalamic malonyl-CoA and food intake. **Proc. Natl. Acad. Sci. USA.**, v. 105, p. 16871-16875, 2008.

CHING, R. H. et al. Supplementation of bitter melon to rats fed a high-fructose diet during gestation and lactation ameliorates fructose-induced dyslipidemia and hepatic oxidative stress in male offspring. **J. Nutr.**, v. 141, n. 9, p. 1664-1672, 2011.

CINTRA, D. E. et al. Interleukin-10 is a protective factor against diet-induced insulin resistance in liver. **J Hepatol.**, v. 48, n. 4, p. 628-37, 2008.

CLAYTON, Z. E. et al. Early Life Exposure to Fructose Alters Maternal, Fetal and Neonatal Hepatic Gene Expression and Leads to Sex-Dependent Changes in Lipid Metabolism in Rat Offspring. **PLoS One**, v. 12, n.11, p. e0141962, 2015.

CUNHA, T. S. et al. Relationship between renal and cardiovascular changes in a murine model of glucose intolerance. **Regul. Pept.**, v. 139, n. 3, p. 1-4, 2007.

DAMIANI, D.; D, DAMIANI. Sinalização cerebral do apetite. **Rev Bras Clin Med.**, v. 9, n. 2, p. 138-45, 2011.

DAVIDOWA, H.; LI, Y.; PLAGEMANN, A. Altered responses to orexigenic (AGRP, MCH) and anorexigenic (alpha-MSH, CART) neuropeptides of paraventricular hypothalamic neurons in early postnatally overfed rats. **Eur. J. Neurosci.**, v. 18, n. 3, p. 613-621, 2003.

DEARDEN, L. OZANNE, S. E. Early life origins of metabolic disease: developmental programming of hypothalamic pathways controlling energy homeostasis. **Frontiers in Neuroendocrinology**, v. 39, p. 3-16, 2015.

DEBASSIO, W. A. et al. Prenatal malnutrition effect on pyramidal and granule cell generation in the hippocampal formation. **Brain Res. Bull**, v. 35, p. 57-61, 1994.

DEKKER, M. J. et al. Fructose: a highly lipogenic nutrient implicated in insulin resistance, hepatic steatosis, and the metabolic syndrome. **Am. J. Physiol. Endocrinol. Metab.**, v. 299, n. 5, p. 685-694, 2010.

DESAI, M. et al. Permanent reduction in heart and kidney organ growth in offspring of undernourished rat dams. **Am. J. Obstet. Gynecol.**, v. 193, p. 1224-1232, 2005.

DONATO, J. JR. et al. Hypothalamic sites of leptin action linking metabolism and reproduction. **Neuroendocrinology**, v. 93, n. 1, p. 9-18, 2011.

DONATO, J. JR. et al. Lesions of the ventral premammillary nucleus disrupt the dynamic changes in Kiss1 and GnRH expression characteristic of the proestrus-estrus transition. **Neuroscience**, v. 25, n. 241, p. 67-79, 2013.

DORNAS, W. C. et al. High dietary salt decreases antioxidant defenses in the liver of fructose-fed insulin-resistant rats. **J. Nutr. Biochem.** v. 24, n. 12, p. 2016-2022, 2013.

ELIAS, C. F. et al. Leptin activates hypothalamic CART neurons projecting to the spinal cord. **Neuron**, v. 21, n 6, p. 1375-1385, 1998.

ELIAS, C. F. et al. Leptin differentially regulates NPY and POMC neurons projecting to the lateral hypothalamic area. **Neuron**, v. 23, n. 4, p. 775-786, 1999.

ELLIOTT, S. S. et al. Fructose, weight gain, and the insulin resistance syndrome. **Am. J. Clin. Nutr.**, v. 76, n. 5, p. 911-922, 2002.

ELMQUIST, J. K. et al. From lesions to leptin: hypothalamic control of food intake and body weight. **Neuron**, v. 22, n. 2, p. 221-232, 1999.

ENG, L. F.; GHIRNIKAR, R. S.; LEE, Y. L. Glial Fibrillary Acidic Protein: GFAP-Thirty-one years (1969-2000). **Neurochemical Research**, v. 25, p. 439-451, 2000.

ERIKSSON, J. G. et al. Childhood growth and hypertension in later life. **Hypertension**, v. 49, n. 6, p. 1415-1421, 2007.

ERLANSON-ALBERTSSON, C. How palatable food disrupts appetite regulation. **Basic. Clin. Pharmacol. Toxicol.**, v. 97, p. 61-73, 2005.

FAEH, D. et al. Effect of fructose overfeeding and fish oil administration on hepatic de novo lipogenesis and insulin sensitivity in healthy men. **Diabetes**, v. 54, n. 7, p. 1907-1913, 2005.

FARAH, V. et al. Nocturnal hypertension in mice consuming a high fructose diet. *Auton Neurosci.* 2006; 130(1-2): 41-50.

FERGUSSON, M. A.; KOSKI, K. G. Comparison of effects of dietary glucose versus fructose during pregnancy on fetal growth and development in rats. **J. Nutr.**, v. 120, p. 1312-1319, 1990.

FOLCH, J. et al. A simple method for the isolation and purification of total lipides from animal tissues. **J. Biol. Chem.**, v. 226, n. 1, p. 497-509, 1957.

FORSEN, T. et al. The fetal and childhood growth of persons who develop type 2 diabetes. **Ann. Intern. Med.**, v. 133, n. 3, p. 176-182, 2000.

FRANCK, N. et al. Insulin-induced GLUT4 translocation to the plasma membrane is blunted in large compared with small primary fat cells isolated from the same individual. **Diabetologia**, v. 50, p. 1716–1722, 2007.

GALE, E. Should we dump the metabolic syndrome? Yes. **BMJ**, v. 22, n. 336 (7645), p. 640-646, 2008.

GAO, S. et al. Leptin activates hypothalamic acetyl–CoA carboxylase to inhibit food intake. **Proc. Natl. Acad. Sci. USA**, v. 104, p. 17358-17363, 2007.

GARCÍA, M. C. et al. Hypothalamic levels of NPY, MCH, and prepro-orexin mRNA during pregnancy and lactation in the rat: role of prolactin. **FASEB J.**, v. 17, n. 11, p. 1392-1400, 2003.

GERICH, J. E. Physiology of glucose homeostasis. **Diabetes Obes. Metab.**, v. 2, n. 6, p. 345-350, 2000.

GHEZZI, A.C. et al. Impact of early fructose intake on metabolic profile and aerobic capacity of rats. **Lipids Health Dis.**, v. 10, n. 3, p. 1-21, 2011.

GLUCKMAN, P. D.; HANSON, M. A. Maternal constraint of fetal growth and its consequences. **Semin. Fetal Neonatal Med.**, v. 9, n. 5, p. 419-425, 2004.

GONZA´LEZ, J. A. et al. Metabolism-independent sugar sensing in central orexin neurons. **Diabetes**, v. 57, p. 2569–2576, 2008.

GORAN, M. I. et al. The obesogenic effect of high fructose exposure during early development. **Nat. Ver. Endocrinol.**, v. 9, n. 8, p. 494-500, 2013.

GRAY, C. et al. Excess maternal salt or fructose intake programmes sex-specific, stress- and fructose-sensitive hypertension in the offspring. **Br. J. Nutr.**, v. 28; n. 115 (4), p. 594-604, 2016.

GRAY, C. et al. Maternal fructose and/or salt intake and reproductive outcome in the rat. Effects on growth, fertility, sex ratio, and birth order. **Biol Reprod.**, v. 89, n. 3 (51), p. 1-8, 2013.

GRESSENS, P. et al. Maternal protein restriction early in rat pregnancy alters brain development in the progeny. **Brain Res. Dev. Brain Res.**, v. 103, p. 21-35, 1997.

GRISSOM, N. M. et al. Dissociable Deficits of Executive Function Caused by Gestational Adversity are Linked to Specific Transcriptional Changes in the Prefrontal Cortex. **Neuropsychopharmacology**, v. 40, n. 6, p. 1353-1363, 2014b.

GRISSOM, N. M. et al. Epigenetic programming of reward function in offspring: a role for maternal diet. *Mammalian genome: official journal of the International Mammalian Genome Society*, v. 25, p. 41-48, 2014a.

GROSS, D. N.; WAN, M.; BIRNBAUM, M. J. The role of FOXO in the regulation of metabolism. **Curr. Diab. Rep.**, v. 9, n. 3, p. 208-214, 2009.

GUGLIUCCI, A. Formation of Fructose-Mediated Advanced Glycation End Products and Their Roles in Metabolic and Inflammatory Diseases. **Adv Nutr.**, v. 8, p. 1, p. 54-62, 2017.

GUGUSHEFF, J. R.; ONG, Z. Y.; MUHLHAUSLER, B. S. Naloxone treatment alters gene expression in the mesolimbic reward system in 'junk food' exposed offspring in a

sex-specific manner but does not affect food preferences in adulthood. **Physiol. Behav.**, v. 22, n. 133, p. 14-21, 2014.

HA, V. et al. Fructose-containing sugars, blood pressure, and cardiometabolic risk: A critical review. **Curr. Hypertens. Rep.**, v. 15, p. 281-297, 2013.

HAEMMERLE, C. A.; CAMPOS, A. M.; BITTENCOURT, J. C. Melanin-concentrating hormone inputs to the nucleus accumbens originate from distinct hypothalamic sources and are apposed to GABAergic and cholinergic cells in the Long-Evans rat brain. **Neuroscience**, v.19, n. 289, p. 392-405, 2015.

HALES, C. N.; BARKER, D. J. et al. Type 2 (non-insulin-dependent) diabetes mellitus: the thrifty phenotype hypothesis. **Diabetologia**, v. 35n. 7, p. 595-601, 1992.

HALL, R. K. et al. Regulation of phosphoenolpyruvate carboxykinase and insulin-like growth factor-binding protein-1 gene expression by insulin. The role of winged helix/forkhead proteins. **J. Biol. Chem.**, v. 29, n. 275(39), p. 30169-30175, 2000.

HALLDORSSON, T. I. et al. Intake of artificially sweetened soft drinks and risk of preterm delivery: A prospective cohort study in 59,334 Danish pregnant women. **Am. J. Clin. Nutr.**, v. 92, p. 626-633, 2010.

HALLFRISCH, J. et al. Insulin and glucose responses in rats fed sucrose or starch. **Am. J. Clin. Nutr.**, v. 32, n. 4, p. 787-7793, 1979.

HEERWAGEN, M. J. et al. Maternal obesity and fetal metabolic programming: A fertile epigenetic soil. **Am. J. Physiol. Regul. Integr. Comp. Physiol.**, v. 299, p. R711–R722, 2010.

HEGYI, K. et al. Leptin-induced signal transduction pathways. **Cell Biol. Int.**, v. 28, n. 3, p. 159-169, 2004.

HOLEMANS, K. et al. Diet-induced obesity in the rat: a model for gestational diabetes mellitus. **Am. J. Obstet. Gynecol.**, v. 190, n. 3, p. 858-865, 2004.

HOUSEKNECHT, K. L. et al. The biology of leptin: a review. **J. Anim. Sci.**, v. 76, n. 5, p. 1405-1420, 1998.

HOWIE G. J. et al. Maternal nutritional history predicts obesity in adult offspring independent of postnatal diet. **J. Physiol.**, v. 587, n. (Pt 4), p. 905-915, 2009.

HSU, C. N. et al. Aliskiren Administration during Early Postnatal Life Sex-Specifically Alleviates Hypertension Programmed by Maternal High Fructose Consumption. **Front Physiol.**, v. 7, n. 299, p. 1-20, 2016.

HUXLEYM, R. R. et al. The role of size at birth and postnatal catch-up growth in determining systolic blood pressure: a systematic review of the literature. **J. Hypertens.**, v. 18, n. 7, p. 815-831, 2000.

International Diabetes Federation. **The IDF consensus worldwide definition of metabolic syndrome. 2006** Disponível em: https://www.idf.org/webdata/docs/IDF_Meta_def_final.pdf. Acesso em: 13 de jul de 2016.

ISHII, Y.; BOURET, S. G. Embryonic birthdate of hypothalamic leptin-activated neurons in mice. **Endocrinology**, v. 153, n. 8, p. 3657-3667, 2012.

JEN, K. L. et al. Fructose and sucrose feeding during pregnancy and lactation in rats changes maternal and pup fuel metabolism. **J. Nutr.**, v. 121, n. 12, p. 1999-2005, 1991.

JESUS, G. C. et al. Principais patologias e biomarcadores das alterações hepáticas. **Estudos**, v. 41, n. 3, p. 525-537, 2014.

JIANG, L.; FERRARIS, R. P. Developmental reprogramming of rat GLUT-5 requires de novo mRNA and protein synthesis. **Am. J. Physiol. Gastrointest. Liver Physiol.**, v. 280, p. G113–G120, 2001.

JOSHI, S. et al. Maternal protein restriction before pregnancy affects vital organs of offspring in Wistar rats. **Metabolism**, v. 52, n. 1, p. 13-18, 2003.

KAC, G. et al. Breastfeeding and postpartum weight retention in a cohort of Brazilian women. **Am. J. Clin. Nutr.**, v. 79, n. 3, p. 487-493, 2004.

KANETO, H. et al. Pdx-1 and MafA Play a Crucial Role in Pancreatic beta-Cell Differentiation and Maintenance of Mature beta-Cell Function. **Endocrine Journal**, v. 55, p. 235-252, 2008.

KAPLAN, N. M. The deadly quartet. Upper-body obesity, glucose intolerance, hypertriglyceridemia and hypertension. **Arch. Intern. Med.**, v. 149, p. 1514-1520, 1989.

KATZ, A. et al. Quantitative insulin sensitivity check index: a simple, accurate method for assessing insulin sensitivity in humans. **J. Clin. Endocrinol. Metab.**, v. 85, p. 2402-2410, 2000.

KAULZKY-WILLER, A. et al. Pronounced insulin resistance and inadequate B-cell secretion characterize lean gestational diabetes during and after pregnancy. **Diabetes Care**, v. 20, p. 1717-1723, 1997.

KELLEY, A. E. et al. Corticostriatal-hypothalamic circuitry and food motivation: integration of energy, action and reward. **Physiol. Behav.**, v. 86, p. 773-795, 2005.

KIM, C.; NEWTON, K. M.; KNOOP, R. H. Gestational diabetes and the incidence of type 2 diabetes. **Diabetes Care**, v. 25, p. 1862-1868, 2002.

KIM, J. et al. Functional roles of fructose. **Proc. Natl. Acad. Sci. USA**, v. 109, p. E1619-E1628, 2012.

KINOTE, A. et al. Fructose-induced hypothalamic AMPK activation stimulates hepatic PEPCK and gluconeogenesis due to increased corticosterone levels. **Endocrinology**, v. 153, n. 8, p. 3633-3645, 2012.

KNOLLEMA, S. et al. Novel hypothalamic and preoptic sites of prepro-melanin-concentrating hormone messenger ribonucleic Acid and Peptide expression in lactating rats. **J Neuroendocrinol.**, v. 4, n. 6, p. 709-717, 1992.

KOLETZKO, S. H. et al. Mixed Expectations: Effects of Goal Ambivalence during Pregnancy on Maternal Well-Being, Stress, and Coping. **Appl. Psychol. Health Well Being.**, v. 7, n. 3, p. 249-274, 2015.

KÖNNER, A. C. et al. Insulin action in AgRP-expressing neurons is required for suppression of hepatic glucose production. **Cell Metab.**, v. 5, n. 6, p. 438-449, 2007.

KÖNNER, A. C. et al. Role for insulin signaling in catecholaminergic neurons in control of energy homeostasis. **Cell Metab.**, v. 8, n. 13(6), p. 720-728, 2011.

KÖNNER, A. C.; BRÜNING, J.C. Selective insulin and leptin resistance in metabolic disorders. **Cell Metab.**, v. 8, n. 16(2), p. 144-152, 2012.

KUHL, C. Insulin secretion and insulin resistance in pregnancy and GDM: implications for diagnosis and management. **Diabetes**, v. 40, p. 18-24, 1991.

LADYMAN, S. R. Hormone interactions regulating energy balance during pregnancy. **J. Neuroendocrinol.**, v. 22, n. 7, p. 805-817, 2010.

LAEMMLI, U. K. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. **Nature**, v. 227, n. 5259, p. 680-685, 1970.

LAGER, S. et al. Diet-induced obesity in mice reduces placental efficiency and inhibits placental mTOR signaling. **Physiol. Rep.**, v. 26, n. 2(2), p. e00242, 2014.

LAWRENCE, J. M. et al. Trends in the prevalence of preexisting diabetes and gestational diabetes mellitus among a racially/ethnically diverse population of pregnant women, 1999-2005. **Diabetes Care**, v. 31, p. 899-904, 2008.

LEE, J. E.; GE, K. Transcriptional and epigenetic regulation of PPAR γ expression during adipogenesis. **Cell Biosci.**, v. 4, n. 29, p. 1-32, 2014.

LIEDTKE, W. et al. GFAP is necessary for the integrity of CNS white matter architecture and long-term maintenance of myelination. **Neuron**, v. 17, p. 607-615, 1996.

LIM, J. S. et al. The role of fructose in the pathogenesis of NAFLD and the metabolic syndrome. **Nat. Rev. Gastroenterol. Hepatol.**, v. 7, n. 5, p. 251-264, 2010.

LINEKER, C. et al. High fructose consumption in pregnancy alters the perinatal environment without increasing metabolic disease in the offspring. **Reprod. Fertil. Dev.**, v. 28, n. 125, p. 2007-2015, 2015.

LIU, X. et al. Effect of pioglitazone on insulin resistance in fructose-drinking rats correlates with AGEs/RAGE inhibition and block of NADPH oxidase and NF kappa B activation. **Eur. J. Pharmacol.**, v. 629, n.1-3, p. 153-158, 2010.

LOPEZ, M. et al. A possible role of neuropeptide Y, agouti-related protein and leptin receptor isoforms in hypothalamic programming by perinatal feeding in the rat. **Diabetologia**, v. 48, p. 140-148, 2005.

LOZANO, I. et al. High-fructose and high-fat diet-induced disorders in rats: impact on diabetes risk, hepatic and vascular complications. **Nutrition & Metabolism**, v. 13, n. 15, p. 1-13, 2016.

LUCA, C.; OLEFSKY, J. M. Inflammation and insulin resistance. **FEBS Lett.**, v. 9, n. 582(1), p. 97-105, 2008.

LUO, D. et al. Effect of pioglitazone on altered expression of Abeta metabolism-associated molecules in the brain of fructose-drinking rats, a rodent model of insulin resistance. **Eur. J. Pharmacol.**, v. 664, n. 1-3, p. 14-19, 2011.

LUO, S. et al. Differential effects of fructose versus glucose on brain and appetitive responses to food cues and decisions for food rewards. **Proc. Natl. Acad. Sci. USA**, v. 19, n. 112 (20), p. 6509-6514, 2015.

LUSTING, R. H. Fructose: metabolic, hedonic, and societal parallels with ethanol. **J. Am. Diet. Assoc.**, v. 110, n. 9, p.1307-1321, 2010.

MALO, E. et al. Plasma lipid levels and body weight altered by intrauterine growth restriction and postnatal fructose diet in adult rats. **Pediatr. Res.**, v. 73, p. 155-162, 2013.

MANTZOROS, C. S. The role of leptin in human obesity and disease: a review of current evidence. **Ann. Intern. Med.**, v. 130, n. 8, p. 671-680, 1999.

MARTINEZ, F. J.; RIZZA, R. A.; ROMERO, J. C. High-fructose feeding elicits insulin resistance, hyperinsulinism, and hypertension in normal mongrel dogs. **Hypertension**, v. 23, n. 4, p. 456-463, 1994.

MASUZAKI, H. et al. Nonadipose tissue production of leptin: leptin as a novel placenta-derived hormone in humans. **Nat. Med.**, v. 3, n. 9, p. 1029-1033, 1997.

MATTHEWS, D. R. et al. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. **Diabetologia**, v. 28, n. 7, p. 412-419, 1985.

MCARDLE, M. A. et al. Mechanisms of obesity-induced inflammation and insulin resistance: insights into the emerging role of nutritional strategies. **Front. Endocrinol. (Lausanne)**, v. 4, n. 52, p. 1-79, 2013.

MCCORMICK, C. M.; Mathews, I. Z. HPA function in adolescence: role of sex hormones in its regulation and the enduring consequences of exposure to stressors. **Pharmacology, biochemistry and behavior**, v. 86, p. 220-233, 2007.

MCMILLEN, I. C.; ROBINSON, J. S. Developmental origins of the metabolic syndrome: prediction, plasticity, and programming. **Physiol. Rev.**, v. 85, n. 2, p. 571-633, 2005.

MELANSON, K. J. et al., High-fructose corn syrup, energy intake, and appetite regulation. **Am. J. Clin. Nutr.**, v. 88, n. 6, p. 1738S-1744S, 2008.

MINISTÉRIO DA SAÚDE. Secretaria de Assistência à Saúde. Departamento de Atenção Básica, Coordenação da Política de Alimentação e Nutrição. **Pesquisa de Orçamentos Familiares (POF), 2008-9**. Pesquisa do IBGE. Disponível em <<http://www.ministeriodasaude.org.br>>. Acesso em: 26 mar. 2013

MINISTÉRIO DA SAÚDE. Secretaria de Atenção à Saúde. Departamento de Atenção Básica. **Guia alimentar para a população brasileira**/Ministério da Saúde, Secretaria de Atenção à Saúde, Departamento de Atenção Básica. – 2. ed., 1. reimpr. – Brasília: Ministério da Saúde, 2014.

MOORE, J. B.; GUNN, P. J.; FIELDING, B. A. The role of dietary sugars and de novo lipogenesis in non-alcoholic fatty liver disease. **Nutrients**, v. 10, n. 6(12), p. 5679-5703, 2014.

MORRIS, M. J.; CHEN, H. Established maternal obesity in the rat reprograms hypothalamic appetite regulators and leptin signaling at birth. **Int. J. Obes. (Lond)**., v. 33, n. 1, p. 115-122, 2009.

MUKAI, Y.; KUMAZAWA, M.; SATO, S. Fructose intake during pregnancy up-regulates the expression of maternal and fetal hepatic sterol regulatory element-binding protein-1c in rats. **Endocrine**, v. 44, p. 79-86, 2013.

NAEF, L. et al. Maternal high-fat intake alters presynaptic regulation of dopamine in the nucleus accumbens and increases motivation for fat rewards in the offspring. **Neuroscience**, v.176, p. 225-236, 2011.

NAEF, L. et al. Reduced anticipatory dopamine responses to food in rats exposed to high fat during early development. **Int. J. Obes. (Lond)**., v. 37, p. 885-888, 2013.

NAGAI, Y. et al. Amelioration of high fructose-induced metabolic derangements by activation of PPARalpha. **Am. J. Physiol. Endocrinol. Metab.**, v. 282, n. 5, p. E1180-E1190, 2002.

NEGRÃO, A. et al. Diferenças na secreção diária de leptina em homens e mulheres: possíveis implicações na neuroendocrinologia dos transtornos alimentares. **Ver. de Psiq. Clín.**, v. 25, n. 4, p. 184-190, 1998.

NEHRING, I. et al. Gestational weight gain and long-term postpartum weight retention: a meta-analysis. **Am. J. Clin. Nutr.**, v. 94, n. 5, p.1225-1231, 2011.

NERY, C. S. et al. Medidas murinométricas e eficiência alimentar em ratos provenientes de ninhadas reduzidas na lactação e submetidos ou não ao exercício de natação. **Rev. Bras. Med. Esporte**, v.17, n. 1, p. 49-55, 2011.

NISWENDER, K. D.; BASKIN, D. G.; SCHWARTZ, M. W. Insulin and its evolving partnership with leptin in the hypothalamic control of energy homeostasis. **Trends Endocrinol. Metab.**, v. 15, n. 8, p. 362-369, 2004.

NOVELLI, E. L. et al. Anthropometrical parameters and markers of obesity in rats. **Lab. Anim.**, v. 41, n. 1, p. 111-119, 2007.

OKADA, Y. et al. Effects of the K⁺ channel opener KRN4884 on the cardiovascular metabolic syndrome model in rats. **J. Cardiovasc. Pharmacol.**, v. 35, n. 2, p. 287-293, 2000.

OLIVERIO, A.; CASTELLANO, C.; ALLEGRA, S. P. Effects of genetic and nutritional factors on post-natal reflex and behavioral development in the mouse. **Exp. Aging. Res.**, v. 1, n. 1, p. 41-56, 1975.

ORON-HERMAN, M.; SELA, B. A.; ROSENTHAL, T. Risk reduction therapy for syndrome X: comparison of several treatments. **Am. J. Hypertens.**, v. 18, n. 3, p. 372-378, 2005.

OUNSTED, M.; SCOTT. A.; OUNSTED, C. Transmission through the female line of a mechanism constraining human fetal growth. **Int. J. Epidemiol.**, v. 37, n. 2, p. 245-250, 2008.

OZOUGWU, J. C. et al. The pathogenesis and pathophysiology of type 1 and type 2 diabetes mellitus. *Journal of Physiology and Pathophysiology*, v. 4, n. 4, p. 46-57, 2013.

PADILLA, S. L.; CARMODY, J. S.; ZELTSER, L. M. Pomc-expressing progenitors give rise to antagonistic neuronal populations in hypothalamic feeding circuits. **Nature Medicine**, v. 16, p. 403-405, 2010.

PAGE, K. A. et al. Effects of fructose vs glucose on regional cerebral blood flow in brain regions involved with appetite and reward pathways. **JAMA**, v. 309, p. 63-71, 2013.

PARK, J. H. et al. Development of type 2 diabetes following intrauterine growth retardation in rats is associated with progressive epigenetic silencing of Pdx1. **J. Clin. Invest.**, v. 118, p. 2316-2324, 2008.

PELCHAT, M. L. Of human bondage: Food craving, obsession, compulsion, and addiction. **Physiol. Behav.**, v. 76, p. 347-352, 2002.

PELLET, P. L.; YOUNG, V. R. Evaluation of protein quality in experimental animals. In: Nutritional Evaluation of Protein Foods. Tokio: **The United Nations University**, p. 41-57, 1980.

PETRY, C. J. et al. Diabetes in old male offspring of rat dams fed a reduced protein diet. **Int. J. Exp. Diabetes Res.**, v. 2, n. 2, p. 139-143, 2001.

PHILLIPS, D. I. Insulin resistance as a programmed response to fetal undernutrition. **Diabetologia**, v. 39, n. 9, p. 1119-1122, 1996.

PLAGEMANN, A. et al. Elevation of hypothalamic neuropeptide Y-neurons in adult offspring of diabetic mother rats. **Neuroreport.**, v. 19, n. 10(15), p. 3211-3216, 1999.

PLAGEMANN, A. et al. Hypothalamic nuclei are malformed in weanling offspring of low protein malnourished rat dams. **J. Nutr.**, v. 130, n. 10, p. 2582-2589, 2000.

PLAGEMANN, A. Perinatal nutrition and hormone-dependent programming of food intake. **Horm. Res.**, v. 65, Suppl 3, p. 83-89, 2006.

PORTHA, B.; CHAVEY, A.; MOVASSAT, J. Early-life origins of type 2 diabetes: fetal programming of the beta-cell mass. **Exp. Diabetes Res.**, v. 2011, n. 105076, p. 1-39, 2011.

PRADA, O. et al. Western diet modulates insulin signaling, c-Jun N-terminal kinase activity, and insulin receptor substrate-1ser307 phosphorylation in a tissue-specific fashion. **Endocrinology**, v. 146, n. 3, p. 1576-1587, 2005.

PRESSE, F. et al. Melanin-concentrating hormone is a potent anorectic peptide regulated by food-deprivation and glucopenia in the rat. **Neuroscience**, v. 71, n. 3, p. 735-745, 1996.

QU, D. et al. A role for melanin-concentrating hormone in the central regulation of feeding behaviour. **Nature**, v. 380, n. 6571, p. 243-247, 1996.

RAWANA S. et al. Low dose fructose ingestion during gestation and lactation affects carbohydrate metabolism in rat dams and their offspring. **J. Nutr.**, v. 123, n. 12, p. 2158-2165, 1993.

REAVEN, G.M. Role of insulin resistance in human disease. **Diabetes**, v. 37, p. 1595-1607, 1988.

REAVEN, G.M. The metabolic syndrome: requiescat in pace. **Clin. Chem.**, v. 51, n. 6, p. 931-938, 2005.

REEVES, P.G.; NIELSEN, F.H.; FAHEY, G.C. JR. AIN-93 purified diets for laboratory rodents: final report of the American Institute of Nutrition ad hoc writing committee on the reformulation of the AIN-76A rodent diet. **J. Nutr.**, v. 123, n. 11, p. 1939-1951, 1993.

REGNAULT, T. R. et al. Fructose, pregnancy and later life impacts. **Clin. Exp. Pharmacol. Physiol.**, v. 40, n. 11, p. 824-837, 2013.

REYNOLDS, C. M. Early Life Nutrition and Energy Balance Disorders in Offspring in Later Life. **Nutrients**, v. 21, n. 7(9), p. 8090-8111, 2015.

RIBEIRO, S. M. L. et al. Leptina: aspectos sobre o balanço energético, exercício físico e amenorreia do esforço. **Arquivo Brasileiro de Endocrinologia e Metabolismo**, v. 51, n. 1, p.11-24, 2007.

RODRÍGUEZ, L. et al. Liquid fructose in pregnancy exacerbates fructose-induced dyslipidemia in adult female offspring. **J. Nutr. Biochem.**, v. 32, p. 115-122, 2016.

RODRÍGUEZ, L. et al. Maternal obesity in the rat programs male offspring exploratory, learning and motivation behavior: prevention by dietary intervention pre-gestation or in gestation. *International journal of developmental neuroscience*. **The official Journal of the International Society for Developmental Neuroscience**, v. 30, p. 75-81, 2012.

ROGERO, M. M.; BORGES, M. C.; PIRES, I. S. O.; TIRAPÉGUI, J. O desmame precoce afeta o ganho de peso e a composição corporal em camundongos adultos?. **Revista de Nutrição**, v. 23, p. 85-93, 2010.

ROGLANS, N. et al. Impairment of hepatic Stat-3 activation and reduction of PPARalpha activity in fructose-fed rats. **Hepatology**, v. 45, p. 778–788, 2007.

RONDINI, T. A. et al. Chemical identity and connections of medial preoptic area neurons expressing melanin-concentrating hormone during lactation. **J. Chem. Neuroanat.**, v. 39, n. 1, p. 51-62, 2010.

RORABAUGH, J. M.; STRATFORD, J. M.; ZAHNISER, N. R. A relationship between reduced nucleus accumbens shell and enhanced lateral hypothalamic orexin neuronal activation in long-term fructose bingeing behavior. **PLoS One**, v. 15, n. 9(4), p. e95019, 2014.

RORABAUGH, J. M.; STRATFORD, J. M.; ZAHNISER, N. R. Differences in bingeing behavior and cocaine reward following intermittent access to sucrose, glucose or fructose solutions. **Neuroscience**, v. 301, p. 213-220, 2015.

SAAD, A. F. et al. High-fructose diet in pregnancy leads to fetal programming of hypertension, insulin resistance, and obesity in adult offspring. **Am. J. Obstet. Gynecol.**, v. 215, n. 3, p. 378-388, 2016.

SANO, H. YOKOI, M. Striatal medium spiny neurons terminate in a distinct region in the lateral hypothalamic area and do not directly innervate orexin/hypocretin- or melanin-concentrating hormone containing neurons. **J. Neurosci.**, v. 27, p. 6948-6955, 2007.

SARI, E. et al. Metabolic and Histopathological Effects of Fructose Intake During Pregestation, Gestation and Lactation in Rats and their Offspring. **J. Clin. Res. Pediatr. Endocrinol.**, v. 7, n. 1, p. 19-26, 2015.

SAWCHENKO, P. E. Toward a new neurobiology of energy balance, appetite, and obesity: the anatomists weigh in. **J. Comp. Neurol.**, v. 402, n. 4, p. 435-441, 1998.

SCHERER, T. et al. Brain insulin controls adipose tissue lipolysis and lipogenesis. **Cell Metab.**, v. 2, n. 13(2), p. 183-194, 2011.

SCHINNER, S. et al. Molecular mechanisms of insulin resistance. **Diabet Med.**, v. 22, n. 6, p. 674-682, 2005.

SCHMIDT, M. I. et al. Gestational diabetes mellitus diagnosed with a 2-h 75-g oral glucose tolerance test and adverse pregnancy outcomes. **Diabetes Care**, v. 24, p. 1151-1160, 2001.

SCHMIDT, M. I. et al. Prevalence of gestational diabetes mellitus – do the new WHO criteria make a difference? **Diabet. Med.**, v. 17, p. 376-380, 2000.

SCHWARTZ, M. W. et al. Central nervous system control of food intake. **Nature**, v. 404, n. 6778, p. 661-671, 2000.

SECO, S.; MATIAS, A. Origem fetal das doenças do adulto: revisitando a teoria de Barker. **Acta Obstet. Ginecol. Port.**, v. 3, n. 3, p. 158-168, 2009.

SFERRUZZI-PERRI, A. N.; CAMM, E. J. The Programming Power of the Placenta. **Front. Physiol.**, v. 14, n. 7, p. 33-43, 2016.

SHAPIRO, A. et al. Fructose-induced leptin resistance exacerbates weight gain in response to subsequent high-fat feeding. **Am. J. Physiol. Regul. Integr. Comp. Physiol.**, v. 295, p. R1370-R1375, 2008.

SHI, X. et al. Fructose transport mechanisms in humans. **Gastroenterology**, v. 113, n. 4, p. 1171-1179, 1997.

SHIMADA, M.; NAKAMURA, T. Time of neuron origin in mouse hypothalamic nuclei. **Exp. Neurol.**, v. 41, n. 1, p. 163-173, 1973.

SHIMOMURA, I. et al. Leptin reverses insulin resistance and diabetes mellitus in mice with congenital lipodystrophy. **Nature**, v. 401, n. 6748, p. 73-76, 1999.

SIEMELINK, M. et al. Dietary fatty acid composition during pregnancy and lactation in the rat programs growth and glucose metabolism in the offspring. **Diabetologia**, v. 45, n. 10, p. 1397-1403, 2002.

SILVA, R.J. et al. Simvastatin-induced cardiac autonomic control improvement in fructose-fed female rats. **Clinics**, v. 66, n. 10, p. 1793-1796, 2011.

SILVA, R.J.; DE ANGELIS, K. Aumento no consumo de frutose como fator de risco para o desenvolvimento de síndrome metabólica. **Revista Eletrônica de Iniciação Científica da USJT**, v. 1, n. 1, p. 10-15, 2007.

SINGH, B. S.; WESTFALL, T. C.; DEVASKAR, S. U. Maternal diabetes-induced hyperglycemia and acute intracerebral hyperinsulinism suppress fetal brain neuropeptide Y concentrations. **Endocrinology**, v. 138, p. 963-969, 1997.

SIVITZ, W. I. et al. Effects of leptin on insulin sensitivity in normal rats. **Endocrinology**, v. 138, p. 3395-3401, 1997.

SKURK, T. et al. Relationship between adipocyte size and adipokine expression and secretion. **J. Clin. Endocrinol. Metab.**, v. 92, p. 1023-1033, 2007.

SMITH, P. M. et al. The subfornical organ: a central nervous system site for actions of circulating leptin. **Am. J. Physiol. Regul. Integr. Comp. Physiol.**, v. 296, n. 3, p. R512-R520, 2009.

SNOECK, A. et al. Effect of a low protein diet during pregnancy on the fetal rat endocrine pancreas. **Biol. Neonate**, v. 57, n. 2, p. 107-118, 1990.

SOCIEDADE BRASILEIRA DE DIABETES. **Diretrizes da Sociedade Brasileira de Diabetes**. A.C. Farmacêutica, 2016.

SOFTIC, S.; COHEN, D. E.; KAHN, C. R. Role of Dietary Fructose and Hepatic De Novo Lipogenesis in Fatty Liver Disease. **Dig. Dis. Sci.**, v. 61, n. 5, p. 1282-1293, 2016.

SPANGLER, R. et al. Opiate-like effects of sugar on gene expression in reward areas of the rat brain. **Brain Res. Mol. Brain Res.**, v. 19, n. 124(2), p. 134-142, 2004.

SPEAR, L. P. The adolescent brain and age-related behavioral manifestations. **Neuroscience and biobehavioral reviews**, v. 24, p. 417-463, 2000.

SPENCER, L. et al. The effect of weight management interventions that include a diet component on weight-related outcomes in pregnant and postpartum women: a systematic review protocol. **JBI Database System Ver. Implement Rep.**, v. 13, n. 1, p. 88-98, 2015.

SRINIVASAN, M. et al. Maternal obesity and fetal programming: effects of a high-carbohydrate nutritional modification in the immediate postnatal life of female rats. **Am. J. Physiol. Endocrinol. Metab.**, v. 295, p. E895-E903, 2008.

STANHOPE, K. L. Sugar consumption, metabolic disease and obesity: The state of the controversy. **Crit. Rev. Clin. Lab. Sci.**, v. 53, n. 1, p. 52-67, 2016.

STEINBERGER, J. et al. Progress and challenges in metabolic syndrome in children and adolescents: a scientific statement from the American Heart Association Atherosclerosis, Hypertension, and Obesity in the Young Committee of the Council on Cardiovascular Disease in the Young; Council on Cardiovascular Nursing; and Council on Nutrition, Physical Activity, and Metabolism. **Circulation**, v. 119, p. 628-647, 2009.

STRATFORD, T. R.; KELLEY, A. E. GABA in the nucleus accumbens shell participates in the central regulation of feeding behavior. **J. Neurosci.**, v. 17, p. 4434-4440, 1997.

STRINGER, G. L. Repeated seizures increase GFAP and vimentin in the hippocampus. **Brain Research**, v. 717, p. 147-153, 1996.

SU, Q. et al. Apolipoprotein B100 acts as a molecular link between lipid-induced endoplasmic reticulum stress and hepatic insulin resistance. **Hepatology**, v. 50, p. 77-84, 2009.

SULLIVAN, E. L. et al. Maternal high-fat diet programming of the neuroendocrine system and behavior. **Horm. Behav.**, v. 76, p. 153-161, 2015.

SUZUKI, M. et al. Effect of an insulin sensitizer, pioglitazone, on hypertension in fructose-drinking rats. **Jpn. J. Pharmacol.**, v. 74, n. 4, p. 297-302, 1997.

SUZUKI, T. et al. Diet-induced epigenetic regulation in vivo of the intestinal fructose transporter GLUT5 during development of rat small intestine. **Biochem. J.**, v. 435, p. 43-53, 2011.

TAIN, Y. L. et al. Maternal fructose-intake-induced renal programming in adult male offspring. **J. Nutr. Biochem.**, v. 26, n. 6, p. 642-650, 2015a.

TAIN, Y. L. et al. Melatonin prevents maternal fructose intake-induced programmed hypertension in the offspring: roles of nitric oxide and arachidonic acid metabolites. **J. Pineal Res.**, v. 57, n. 1, p. 80-89, 2014.

TAIN, Y. L. et al. PPARs Link Early Life Nutritional Insults to Later Programmed Hypertension and Metabolic Syndrome. **Int. J. Mol. Sci.**, v. 17, n. 1, p. E20-E28, 2015b.

TAPPY, L.; LE, K. A. Metabolic effects of fructose and the worldwide increase in obesity. **Physiol Rev.**, v. 90, n. 1, p. 23-46. 2010.

TARRY-ADKINS, J. L.; OZANNE, S. E. Mechanisms of early life programming: Current knowledge and future directions. **Am. J. Clin. Nutr.**, v. 94, n. (Suppl.), p. S1765-S1771, 2011.

TAYLOR, P. D.; POSTON, L. Developmental programming of obesity in mammals. **Exp. Physiol.**, v. 92, n. 2, p. 287-298, 2007.

TEFF, K. L. et al. Dietary fructose reduces circulating insulin and leptin, attenuates postprandial suppression of ghrelin, and increases triglycerides in women. **J. Clin. Endocrinol. Metab.**, v. 89, n. 6, p. 2963-2972, 2004.

TERRONI, P. L. et al. Expression of agouti-related peptide, neuropeptide Y, pro-opiomelanocortin and the leptin receptor isoforms in fetal mouse brain from pregnant dams on a protein-restricted diet. **Brain Res. Mol. Brain Res.**, v. 140, p. 111-115, 2005.

THANGARATINAM, S. et al. Accuracy of liver function tests for predicting adverse maternal and fetal outcomes in women with preeclampsia: a systematic review. **Acta Obstet. Gynecol. Scand.**, v. 90, n. 6, p. 574-585, 2011.

THEYS, N. et al. Maternal malnutrition programs pancreatic islet mitochondrial dysfunction in the adult offspring. **J. Nutr. Biochem.**, v. 22, n. 10, p. 985-994, 2011.

THORBURN, A. W. et al. Fructose-induced in vivo insulin resistance and elevated plasma triglyceride levels in rats. **Am. J. Clin. Nutr.**, v. 49, n. 6, p. 1155-1163, 1989.

THORPE, L. E. et al. Trends and racial/ethnic disparities in gestational diabetes among pregnant women in New York City, 1990-2001. **Am. J. Public Health.**, v. 95, n. 9, p. 1536-1539, 2005.

TOWBIN, R.; DUNBAR, J. S.; BOVE, K. Antrochoanal polyps. **AJR Am. J. Roentgenol.**, v. 132, n. 1, p. 27-31, 1979.

TRITOS, N. A. et al. Characterization of expression of hypothalamic appetite-regulating peptides in obese hyperleptinemic brown adipose tissue-deficient (uncoupling protein-promoter-driven diphtheria toxin A) mice. **Endocrinology**, v. 139, n. 11, p. 4634-4641, 1998.

TRUJILLO, M. L. Hyperphagia and central mechanisms for leptin resistance during pregnancy. **Endocrinology**, v. 152, n. 4, p. 1355-1365, 2011.

TSAI, J. et al. Inflammatory NF-kappaB activation promotes hepatic apolipoprotein B100 secretion: evidence for a link between hepatic inflammation and lipoprotein production. **Am. J. Physiol. Gastrointest.**, v. 296, n. 6, p. G1287-G1298, 2009.

TSENG, K. Y. The neonatal ventral hippocampal lesion as a heuristic neurodevelopmental model of schizophrenia. **Journal of Behavioral Brain Research.**, v. 391, n. 2, p. 295-305, 2009.

TURNER, R. et al. Insulin deficiency and insulin resistance interaction in diabetes: Estimation of their relative contribution by feedback analysis from basal plasma insulin and glucose concentration in man. **Metabolism.**, v. 28, p. 1086-1096, 1979.

VAN DE WALL, E. et al. Collective and individual functions of leptin receptor modulated neurons controlling metabolism and ingestion. **Endocrinology**, v. 149, n. 4, p. 1773-1785, 2008.

VANHAESEBROECK, B.; ALESSI, D. R. The PI3K-PDK1 connection: more than just a road to PKB. **Biochemical Journal**, v. 346, p. 561-576, 2000.

VELLOSO, L. A. O Controle Hipotalâmico da Fome e da Termogênese – Implicações no Desenvolvimento da Obesidade. **Arq. Bras. Endocrinol. Metab.**, v. 50, n. 2, p. 165-176, 2006.

VESCO, K. K. et al. Excessive gestational weight gain and postpartum weight retention among obese women. **Obstet. Gynecol.**, v. 114, n. 5, p. 1069-1075, 2009.

VICKERS, M. H. Developmental programming of the metabolic syndrome - critical windows for intervention. **World J. Diabetes**, v. 2, n. 9, p. 137-148, 2011a.

VICKERS, M. H. et al. Fetal origins of hyperphagia, obesity, and hypertension and postnatal amplification by hypercaloric nutrition. **Am. J. Physiol. Endocrinol. Metab.**, v. 279, n. 1, p. E83-E87, 2000.

VICKERS, M. H. et al. Maternal fructose intake during pregnancy and lactation alters placental growth and leads to sex-specific changes in fetal and neonatal endocrine function. **Endocrinology**, v. 152, p. 1378-1387, 2011b.

VUCETIC, Z. et al. Maternal high-fat diet alters methylation and gene expression of dopamine and opioid-related genes. **Endocrinology**, v. 151, p. 4756-4764, 2010.

WÅHLEN, K.; SJÖLIN, E.; LÖFGREN, P. Role of fat cell size for plasma leptin in a large population based sample. **Exp. Clin. Endocrinol. Diabetes**, v. 119, p. 291–294, 2011.

WEISINGER, R. S. et al. Angiotensin converting enzyme inhibition lowers body weight and improves glucose tolerance in C57BL/6J mice maintained on a high fat diet. **Physiol. Behav.**, v. 4, n. 98(1-2), p. 192-197, 2009.

WELSH, J. A. et al. Caloric sweetener consumption and dyslipidemia among US adults. **JAMA**, v. 303, n. 15, p. 1490-1497, 2010.

WILSON, B. D.; OLLMANN, M.M.; BARSH, G.S. The role of agouti-related protein in regulating body weight. **Mol. Med. Today**, v. 5, n. 6, p. 250-256, 1999.

WILSON, R. D.; ISLAM, M. S. Fructose-fed streptozotocin-injected rat: An alternative model for type 2 diabetes. **Pharmacol. Rep.**, v. 64, p. 129-139, 2012.

WOLFGANG, M. J. et al. Hypothalamic malonyl-CoA and the control of energy balance. **Mol. Endocrinol.**, v. 22, n. 18, p. 2012–2020, 2008.

WOLFGANG, M. J. et al. Regulation of hypothalamic malonyl-CoA by central glucose and leptin. **Proc. Natl. Acad. Sci. USA**, v. 104, p. 19285-19290, 2007.

WOODSIDE, B. et al. Many mouths to feed: the control of food intake during lactation. **Front. Neuroendocrinol.**, v. 33, n. 3, p. 301-314, 2012.

WORLD HEALTH ORGANIZATION. **Global status report on non communicable diseases 2014**. Disponível em: <http://www.who.int/nmh/publications/ncd-status-report-2014/en/>. Acesso em: 08 out. 2015.

WORLD HEALTH ORGANIZATION. **Obesity and overweight**. Disponível em: <http://www.who.int/dietphysicalactivity/publications/facts/obesity/en/>. Acesso em: 03 out. 2012

XIN, F.; SUSIARJO, M.; BARTOLOMEI, M. S. Multigenerational and transgenerational effects of endocrine disrupting chemicals: A role for altered epigenetic regulation? **Semin. Cell Dev. Biol.**, v. 43, p. 66-75, 2015.

YAJNIK, C. S. et al. Neonatal anthropometry: the thin-fat Indian baby. The Pune Maternal Nutrition Study. **Int. J. Obes. Relat. Metab. Disord.**, v. 27, n. 2, p. 173-180, 2003.

YAMADA-OBARA, N. et al. Maternal exposure to high-fat and high-fructose diet evokes hypoadiponectinemia and kidney injury in rat offspring. **Clin. Exp. Nephrol.**, v. 20, n. 6, p. 853-861, 2016.

YILMAZ, Y. Review article: Fructose in non-alcoholic fatty liver disease. **Aliment. Pharmacol. Ther.**, v. 35, p. 1135-1144, 2012.

ZAMPIERI, T. T. et al. SOCS3 expression within leptin receptor-expressing cells regulates food intake and leptin sensitivity but does not affect weight gain in pregnant mice consuming a high-fat diet. **Physiol. Behav.**, v. 1, n. 157, p. 109-115, 2016.

ZAVARONI, I. et al. Effect of fructose feeding on insulin secretion and insulin action in the rat. **Metabolism.**, v. 29, n. 10, p. 970-973, 1980.

ZHANG, Q. Y. et al. Quercetin inhibits AMPK/TXNIP activation and reduces inflammatory lesions to improve insulin signaling defect in the hypothalamus of high fructose-fed rats. **J. Nutr. Biochem.**, v. 25, n. 4, p. 420-428, 2014.

ZHANG, Y. et al. Positional cloning of the mouse obese gene and its human homologue. **Nature**, v. 372, n. 6505, p. 425-432, 1994.

ZHANG, Z. Y. et al. Supplementation of the maternal diet during pregnancy with chocolate and fructose interacts with the high-fat diet of the young to facilitate the onset of metabolic disorders in rat offspring. **Clin. Exp. Pharmacol. Physiol.**, v. 40, n. 9, p. 652-661, 2013.

ZHAO, Y. et al. Treatment of rats with Jiangzhi Capsule improves liquid fructose-induced fatty liver: modulation of hepatic expression of SREBP-1c and DGAT-2. **J. Transl. Med.**, v. 2, n. 13, p. 174-180, 2015.

ZHENG, J. et al. Early Life Fructose Exposure and Its Implications for Long-Term Cardiometabolic Health in Offspring. **Nutrients**, v. 1, n. 8(11), p. E685-E692, 2016.

ZHENG, J. et al. Maternal and post-weaning high-fat, high-sucrose diet modulates glucose homeostasis and hypothalamic POMC promoter methylation in mouse offspring. **Metab. Brain. Dis.**, v. 30, n. 5, p. 1129-1137, 2015.

ZOU, M. et al. Fructose consumption during pregnancy and lactation induces fatty liver and glucose intolerance in rats. **Nutr. Res.**, v. 32, n. 8, p. 588-598, 2012.