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Mais do que um apoio:
a conexão da uma liana a suportes leva a
profundas mudanças na anatomia do xilema, de
condutividade hidráulica, e do perfil
transcricional do câmbio

More Than Supportive:
Liana Attachment to Supports Lead to Profound
Changes in Xylem Anatomy, Hydraulic
Conductivity and Cambium Transcriptional
Profile

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Resumo

- O xilema secundário desempenha duas funções cruciais: o suporte mecânico e condução de água e minerais. Muitas lianas mostram duas fases anatômicas contrastantes em seu xilema, uma inicial, onde o xilema é homogêneo, fibroso e apresenta vasos de pequeno diâmetro, denominada xilema autoportante, e posteriormente o xilema lianescente, anatomicamente complexo, com vasos grandes e dimórficos e menos tecido de suporte. Não se sabe, no entanto, o que leva à mudança abrupta da formação do xilema.

- Neste estudo, abordamos as questões de o que inicia e quais são os determinantes genéticos das profundas mudanças durante a transição do xilema autossuporte para o xilema lianescente na liana *Bignonia magnifica*, Bignoniaceae. Para esse fim, analisamos primeiramente as taxas de crescimento de plantas cultivadas com e sem suportes, como os parâmetros hidráulicos variavam ao longo do caule e descrevemos em detalhes a anatomia do xilema autossuportante e lianescente. Em seguida, construímos o transcriptoma do câmbio e do xilema em diferenciação dessas duas fases e realizamos uma análise de expressão diferencial dos dados gerados por RNA-Seq.

- Nosso trabalho mostra que a presença de suportes leva a alterações no padrão de crescimento e nos parâmetros anatômicos ao longo do caule, aumentando a condutividade potencial e promovendo a formação de xilemas lianescentes. Essas alterações estão associadas à expressão diferencial de genes relacionados à divisão celular e à biossíntese da parede celular, sobre-expressos na fase autoportante, e de fatores de transcrição, defesa / morte celular e genes responsivos a hormônios, sobre-expressos na fase lianescente.

- Concluímos que a anatomia mais complexa na fase lianescente é o resultado de uma regulação transcricional mais complexa no câmbio e xilema em diferenciação.

Palavras-chave: xilema; xilogênese; transcriptoma; anatomia funcional; condutividade hídrica

Summary

- Secondary xylem performs two crucial functions, namely mechanical support and water and mineral conduction. Many lianas show two contrasting xylem anatomy phases, the initial homogeneous, fibrous and small vesselled self-supporting xylem, and the later lianescent xylem, which is anatomically complex, with large and dimorphic vessels and less supportive tissue. It is not known, however, what leads to the abrupt change of xylem formation.
- In this study, we address the question of what triggers and which are the genetic determinants of the profound changes during the transition from self-supporting to the lianescent xylem in the liana *Bignonia magnifica*, Bignoniaceae. For this purpose, we first analyzed growth rates of plants grown with and without supports, how hydraulic parameters varied along the stem and described in detail self-supporting and lianescent xylem anatomy. We then constructed cambium and differentiating xylem transcriptome of these two phases and conducted a differential expression analysis of RNA-Seq generated data.
- Our work shows that the presence of supports leads to changes in growth pattern and anatomical parameters along the stem, increasing potential conductivity, and promotes lianescent xylem formation. These changes are associated to differential expression of genes related to cell division and cell wall biosynthesis, overregulated in self-supporting phase, and of transcription factors, defense/cell death, and hormone-responsive genes, overregulated in lianescent phase.
- We conclude that the more complex anatomy in lianescent phase is the result from a more complex transcriptional regulation in cambium and wood forming tissues.

Keywords: xylem; xylogenesis; transcriptome; functional anatomy; hydraulic conductivity.

Introduction

Secondary xylem, or simply wood, performs two crucial functions for woody plants survival and reproductive success: mechanical support, optimizing light uptake for photosynthesis and exposure of reproductive organs; and the conduction of water and nutrients from the soil and between source and sink organs, serving yet as storage tissue during unfavorable periods (Baas *et al.*, 2004). In contrast to the homogeneous, almost entirely tracheid composed gymnosperm wood, cellular specialization evolution in angiosperm xylem led to a division of labor among the different cell types composing it, with support being carried out by fibers, conduction of water and minerals carried out mainly by vessels, and storage by axial and radial parenchyma (Bailey & Tupper, 1918; Tyree & Zimmermann, 2002; Evert, 2006). Despite the established relationship of cell types and their functions, there is a great diversity of arrangements, compositions and dimensions of these cell types in the xylem of plants. In turn, this anatomical structural diversity are under genetic control, which regulation has been unveiled with the studies in model species such as *Arabidopsis thaliana* and *Populus* (J. Zhang *et al.*, 2014; Ye & Zhong, 2015; Sundell *et al.*, 2017; J. Zhang *et al.*, 2019), and define how plants cope with the different functions (Zieminska *et al.*, 2013; 2015; Beekman, 2016). Overall, relations between genetic control, anatomy structure and functional implications are still poorly understood for wood plants.

Mechanical properties of fibers are controlled by the cell wall thickness, cell wall/lumen ratio (Zieminska *et al.*, 2013), as well as by cell wall hemicellulose and lignin composition, and cellulose amount and microfibril angle (Bergander & Salmén, 2002; Li *et al.*, 2009; MacMillan *et al.*, 2010; Bourmaud *et al.*, 2013). Water conductivity (K), in turn, is directly related to vessels diameter (D) raised to the fourth power and inversely related to the length (L) that the water must travel between roots and leaves. The relation

of K with D^4 shows that even small increments of diameter profound considerable impacts on conductivity, while growth in L could lead to smaller water supply for distally located leaves and limitation in length growth due to an increase in conductivity resistance (Petit & Anfodillo, 2009; Anfodillo *et al.*, 2013). However, the increase of vessels diameter found along the stem allows for a compensatory effect on the path length resistance (West *et al.*, 1999), an hypothesis that was further reinforced by later works (Becker *et al.*, 2000; Sperry *et al.*, 2012). Olson *et al.* (2014), analyzing the stem xylem anatomy at the base and at the top of 257 angiosperm species, found a vessel widening ratio of $D = L^{0.22}$, independent of habit and habitats. This proportion between L and D describes a fast growth in diameter near the apex and a progressive lesser increase toward the base, and its value is consistent with previous models (Becker *et al.*, 2000). The controversy, as well as the insight, of the model lies in the assumption that length is the main factor, if not the only one, determining vessel diameter.

Nevertheless, different proportions of the different cell types that compose secondary xylem may lead to changes in stem properties and how they cope with support and conductivity (Zieminska *et al.*, 2013; 2015; Bittencourt *et al.*, 2016; Gerolamo *et al.* unpublished). Lianas, or woody vines, are an extreme example of reduced amount of supporting cells, namely xylem fibers, that accompanied the evolution of this life form, which uses other plants as supports. Although this iconic component of tropical forests has evolved several times in different groups (Gentry, 1991; Angyalossy *et al.*, 2015), the life form shows a series of striking converging anatomical features, as the presence of huge vessels, up to 500 micrometers in diameter (Angyalossy *et al.*, 2015), that are associated with small vessels (vessel dimorphism, Carlquist, 1981). Moreover, the reduction of fibers and increase of parenchyma in comparison to phylogenetically related species, and the presence of cambial variations, that are different arrangements of

secondary vascular tissues (Carlquist, 1985; 2001; Angyalossy *et al.*, 2012) are characteristic of lianescent habit. All these features are collectively called lianescent vascular syndrome (Angyalossy *et al.*, 2015) and are observed in both existing and extinct liana species (Burnham, 2009).

Nevertheless, many lianas show a denser xylem at the beginning of their development, which is more fibrous and shows smaller vessels, resembling the xylem of self-supporting species (Schenck, 1893; Obaton, 1960; Caballé, 1998; Gallenmuller *et al.*, 2001). The transition from this denser xylem phase (termed as self-supporting xylem hereafter) to that showing the lianescent vascular syndrome (named as lianescent xylem hereafter) occurs abruptly, as seen in the adult stem cross-section (Fig. 1), and was observed in 85 % of the more than 13,000 specimens, belonging to 400 liana species of 50 different families analyzed by Caballé (1998). The beginning of lianescent xylem production occurs, in different stems, after the formation of different amounts of the self-supporting xylem (Caballé, 1998), which suggests that it is not as a result of a pre-defined developmental program. It is not known, however, what leads to the abrupt change of xylem formation (Rowe & Speck, 1996; Caballé, 1998). While several reports have investigated the exuberant anatomical architectures formed due to cambial variants (Rajput *et al.*, 2008; Pace *et al.*, 2009; Tamaio *et al.*, 2010; Cabanillas *et al.*, 2017), the sudden change in lianas xylem anatomy has been poorly addressed (Obaton, 1960; Caballé, 1993; 1998; Gallenmüller *et al.*, 2001). The detailed characterization of the two anatomy phases, not only contributes to the better understanding of the control of their formation, the demands suffered and met by lianas xylem and the evolution of this important life form, but also shed light on factors that modulate the development of

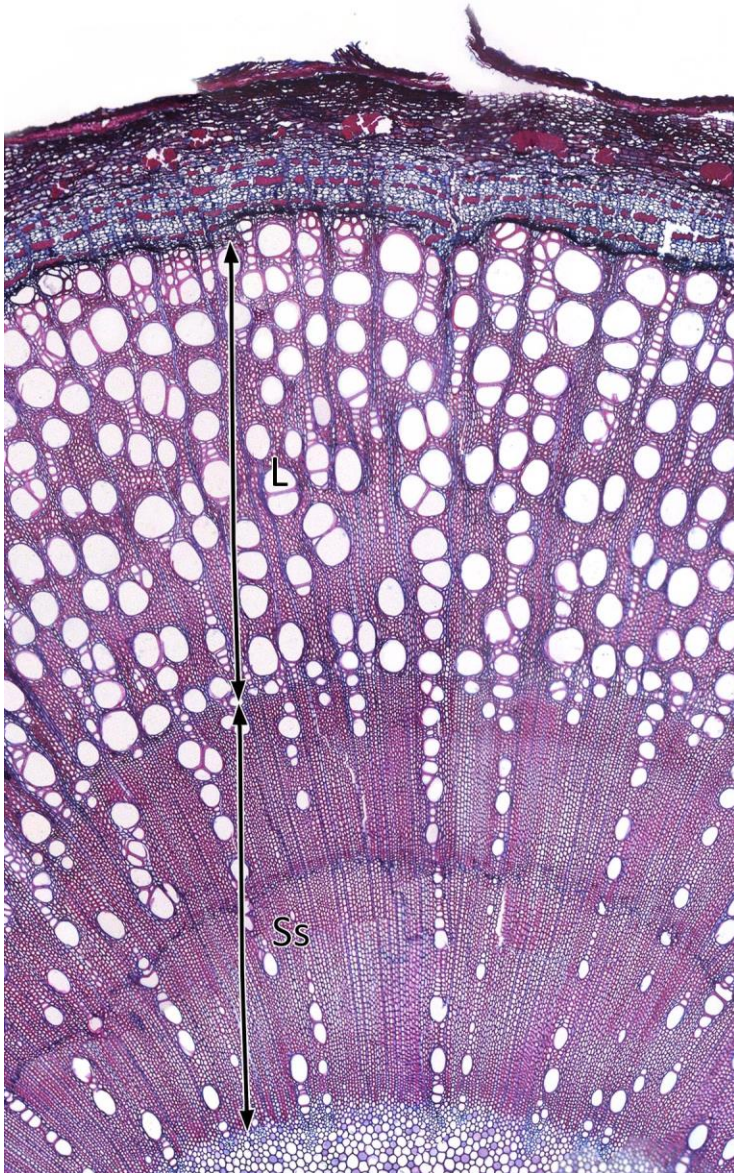


Fig. 1. *B. magnifica* stem cross section. As *B. magnifica*, many lianas show the formation of a denser xylem at the beginning of the secondary growth, composed by a high proportion of fibers and with small vessels, denominated here self-supporting xylem (Ss), and a sudden change to a xylem showing the lianescent vascular syndrome, *i.e.* with less fibers and large vessels, which are associated with small vessels, denominated lianescent xylem (L).

secondary xylem, with underlying genetic traits and the implications for hydraulic conductivity and mechanical performance, in plants as a whole.

The use of model species, as *Arabidopsis thaliana* and *Populus*, has provided invaluable advances in the understanding of cambium installation and activity, as well as xylem formation and differentiation (Groover, 2005; Hidakawa *et al.*, 2010; Agusti *et al.*, 2011; Robischon *et al.*, 2011; J. Zhang *et al.*, 2014; 2019; Ye & Zhong, 2015; Sundell *et al.*, 2017). The signaling pathway regulating cambial

maintenance and proliferation is conserved from herbaceous to woody species, as described in both *A. thaliana* and *Populus*, and is controlled by the module TDIF/CLE41/CLE44-TDR/PXY-WOX4 (Zhang *et al.*, 2014). TRACHEARY DIFFERENTIATION INHIBITORY FACTOR (TDIF) peptides, which are products of

the posttranslational process of CLAVATA/EMBRYO SURROUNDING REGION (CLE41 /CLE44) protein are produced in the phloem and perceived in the (pro)cambium by the receptor-like kinase TDIF RECEPTOR/PHLOEM INTERCALATED WITH XYLEM (TDR/PXY). This interaction induces the expression of the *WUSCHEL-RELATED HOMEODOMAIN* gene (*WOX4*), which in turn regulates cell proliferation (Hirakawa et al., 2010; Etchells & Turner, 2010). Secondary cell wall (SCW) biosynthesis regulation by the primary and secondary master switches NAM/ATAF1/CUC2 (NAC) and MYB (V-MYB protein from avian myeloblastosis virus) transcription factors, respectively, was also described in *A. thaliana* (Kubo et al., 2005; Zhong et al., 2006; McCarthy et al., 2009; Ko et al. 2009). The elaborated transcriptional network controlling fiber and vessels differentiation was further characterized in numerous other species, such as *Populus trichocarpa*, *Oryza sativa*, *Brachypodium distachyon*, *Eucalyptus grandis* and *Zea mays* (Hu et al., 2010; Nuruzzaman et al., 2010; Valdivia et al., 2013; Soler et al., 2015; Zhong et al., 2011).

Molecular studies in non-model species, on the other hand, have become possible in an unprecedented way thanks to the use of new technologies, such as next-generation sequencing (Wang *et al.*, 2009), and allowed to unravel many unique features and processes absent in model species (Carpentier *et al.*, 2008). In this study, we made an integrative approach to address the question of what triggers and which are the genetic determinants of the profound changes during the transition from self-supporting to the lianescent xylem in the liana *Bignonia magnifica* W.Bull (Bignoniaceae). For this purpose, we first analyzed growth rates of plants grown with and without supports, how xylem parameters varied along the stem, described in detail self-supporting and lianescent xylem anatomy and what are the implications for hydraulic conductivity. We then

constructed cambium and differentiating xylem transcriptome of these two phases and conducted a differential expression analysis of RNA-Seq generated data.

Our work shows that the presence of supports leads to changes in growth pattern and anatomical parameters along the stem, increasing specific conductivity, and promotes lianescent xylem formation. These changes are associated to differential expression of genes related to cell division, cell wall, transcription factors, defense/cell death, and hormone-responsive genes.

Conclusions

In the present study, we verified profound impacts of support on the liana *B. magnifica* shoot development, increasing growth in length, decreasing growth in thickness and promoting the formation of the lianescent xylem. The detailed xylem anatomical characterization showed that the onset of lianescent phase is characterized by much larger vessels, whose production is not anymore restricted to the front of protoxylem poles, that drastically increase specific conductivity. The comprehensive integration of anatomical and differential expression analysis data allows us to propose a model to characterize the molecular control of the lianescent vascular syndrome establishment (Fig 11). Our model shows that the more complex lianescent xylem reflects an also more intricate transcriptional regulation network, involving a more diverse repertory of transcription factors and hormone responsive genes.

References

- Agusti, J., Herold, S., Schwarz, M., Sanchez, P., Ljung, K., Dun, E. A., ... & Greb, T. 2011. Strigolactone signaling is required for auxin-dependent stimulation of secondary growth in plants. *Proceedings of the National Academy of Sciences*, 108(50), 20242-20247.
- Aloni, R., & Jacobs, W. P. 1977. Polarity of tracheary regeneration in young internodes of *Coleus* (Labiatae). *American Journal of Botany*, 64(4), 395-403.
- Aloni, R., Schwalm, K., Langhans, M., & Ullrich, C. I. 2003. Gradual shifts in sites of free-auxin production during leaf-primordium development and their role in vascular differentiation and leaf morphogenesis in *Arabidopsis*. *Planta*, 216(5), 841-853.
- Anfodillo, T., Petit, G., & Crivellaro, A. 2013. Axial conduit widening in woody species: a still neglected anatomical pattern. *Iawa Journal*, 34(4), 352-364.
- Angyalossy, V., Angeles, G., Pace, M. R., Lima, A. C., Dias-Leme, C. L., Lohmann, L. G., & Madero-Vega, C. 2012. An overview of the anatomy, development and evolution of the vascular system of lianas. *Plant Ecology & Diversity*, 5, 167-182.
- Angyalossy, V., Pace, M. R., & Lima, A. C. 2015. Liana anatomy: a broad perspective on structural evolution of the vascular system. In Schnitzer, S. A., Bongers, F., Burnham, R. J. & Putz, F. E. (eds.). *Ecology of Lianas*. John Wiley & Sons Ltd. Chichester.
- Antosch, M., Schubert, V., Holzinger, P., Houben, A., & Grasser, K. D. 2015. Mitotic lifecycle of chromosomal 3x HMG-box proteins and the role of their N-terminal domain in the association with r DNA loci and proteolysis. *New Phytologist*, 208(4), 1067-1077.
- Baas, P., Ewers, F. W., Davis, S. D., & Wheeler, E. A. 2004. Evolution of xylem physiology. In Hemsley, A. R., Poole, I. (eds.). *The Evolution of Plant Physiology*. Elsevier Academic Press, London.
- Baas, P., Schmid, R., & van Heuven, B. J. 1986. Wood anatomy of *Pinus longaeva* (bristlecone pine) and the sustained length-on-age increase of its tracheids. *IAWA Journal*, 7(3), 221-228.
- Bailey, I. W., & Tupper, W. W. 1918. Size variation in tracheary cells: I. A comparison between the secondary xylems of vascular cryptogams, gymnosperms and angiosperms. *Proceedings of the American Academy of Arts and Sciences*, 54 (2), 149-204.
- Baima, S., Possenti, M., Matteucci, A., Wisman, E., Altamura, M. M., Ruberti, I., & Morelli, G. 2001. The *Arabidopsis* ATHB-8 HD-zip protein acts as a differentiation-promoting transcription factor of the vascular meristems. *Plant physiology*, 126(2), 643-655.
- Barbosa, A. C. F., Pace, M. R., Witovsk, L., & Angyalossy, V. 2010. A new method to obtain good anatomical slides of heterogeneous plant parts. *IAWA Journal*, 31, 373-383.
- Becker, P., Gribben, R. J., & Lim, C. M. 2000. Tapered conduits can buffer hydraulic conductance from path-length effects. *Tree Physiology*, 20(14), 965-967.
- Beeckman, H. 2016. Wood anatomy and trait-based ecology. *IAWA journal*, 37(2), 127-151.

- Bergander, A., & Salmén, L. 2002. Cell wall properties and their effects on the mechanical properties of fibers. *Journal of materials science*, 37(1), 151-156.
- Berlyn, G. P., & Miksche J. P. 1976. Botanical microtechnique and cytochemistry. Iowa State University Press, Ames.
- Beyer, M., Nazareno, A. G., & Lohmann, L. G. 2019. Development of nuclear microsatellite markers in *Stizophyllum* (Bignoniaceae) using next-generation sequencing. *Plant Genetic Resources*, 1-4.
- Bittencourt, P. R., Pereira, L., Oliveira, R. S. 2016. On xylem hydraulic efficiencies, wood space-use and the safety–efficiency tradeoff: comment on Gleason *et al.* (2016) ‘Weak tradeoff between xylem safety and xylem-specific hydraulic efficiency across the world’s woody plant species’. *New Phytologist* 211:1152–1155.
- Björklund, S., Antti, H., Uddestrand, I., Moritz, T., & Sundberg, B. 2007. Cross-talk between gibberellin and auxin in development of *Populus* wood: gibberellin stimulates polar auxin transport and has a common transcriptome with auxin. *The Plant Journal*, 52(3), 499-511.
- BLAST® Command Line Applications User Manual [Internet]. Bethesda (MD): National Center for Biotechnology Information (US); 2008-. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK279690/>
- Boerjan, W., Ralph, J., & Baucher, M. 2003. Lignin biosynthesis. *Annual review of plant biology*, 54(1), 519-546.
- Bollhöner, B., Prestele, J., & Tuominen, H. 2012. Xylem cell death: emerging understanding of regulation and function. *Journal of Experimental Botany*, 63(3), 1081-1094.
- Bourmaud, A., Morvan, C., Bouali, A., Placet, V., Perre, P., & Baley, C. 2013. Relationships between micro-fibrillar angle, mechanical properties and biochemical composition of flax fibers. *Industrial Crops and Products*, 44, 343-351.
- Bukatsch, F. 1972. Bemerkungem zur Doppelfärbung Astra-blau-Safranin. *Mikrokosmos* 6:255.
- Burnham, R.J. 2009. An overview of the fossil record of climbers: bejucos, sogas, trepadoras, lianas, cipós, and vines. *Revista Brasileira de Paleontologia*, 12, 149–160.
- Caballé, G. U. Y. 1993. Liana structure, function and selection: a comparative study of xylem cylinders of tropical rainforest species in Africa and America. *Botanical Journal of the Linnean Society*, 113(1), 41-60.
- Caballé, G. 1998. Le port autoportant des lianes tropicales: une synthèse des stratégies de croissance. *Canadian Journal of Botany*, 76, 1703-1716.
- Cabanillas, P. A., Pace, M. R., & Angyalossy, V. 2017. Structure and ontogeny of the fissured stems of *Callaeum* (Malpighiaceae). *IAWA Journal*, 38(1), 49-66.
- Caño-Delgado, A., Yin, Y., Yu, C., Vafeados, D., Mora-García, S., Cheng, J. C., ... & Chory, J. 2004. BRL1 and BRL3 are novel brassinosteroid receptors that function in vascular differentiation in *Arabidopsis*. *Development*, 131(21), 5341-5351.

- Carlquist, S. 1962. A theory of paedomorphosis in dicotyledonous woods. *Phytomorphology*, 12(1), 30-45.
- Carlquist, S. 1981. Wood anatomy of Nepenthaceae. *Bulletin of the Torrey Botanical Club*, 108, 324-330.
- Carlquist, S. 1985. Observations on functional wood histology of vines and lianas: vessel dimorphism, tracheids, vasicentric tracheids, narrow vessels, and parenchyma. *Aliso* 11, 139-157.
- Carlquist, S. 1991. Anatomy of vine and liana stems: a review and synthesis. In Putz, F. E. & Mooney, H.A. (eds.). *The Biology of Vines*. Cambridge University Press, Cambridge.
- Carlquist, S. 2001. *Comparative Wood Anatomy*. 2nd ed. Springer, Berlin.
- Carpentier, S. C., Panis, B., Vertommen, A., Swennen, R., Sergeant, K., Renaut, J., ... & Devreese, B. 2008. Proteome analysis of non-model plants: a challenging but powerful approach. *Mass spectrometry reviews*, 27(4), 354-377.
- Cassan-Wang, H., Goué, N., Saidi, M. N., Legay, S., Sivadon, P., Goffner, D., & Grima-Pettenati, J. 2013. Identification of novel transcription factors regulating secondary cell wall formation in Arabidopsis. *Frontiers in plant science*, 4, 189.
- Choat, B., Brodie, T. W., Cobb, A. R., Zwieniecki, M. A., & Holbrook, N. M. 2006. Direct measurements of intervessel pit membrane hydraulic resistance in two angiosperm tree species. *American journal of botany*, 93(7), 993-1000.
- Conesa, A., Götz, S., García-Gómez, J. M., Terol, J., Talón, M., & Robles, M. 2005. Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. *Bioinformatics*, 21(18), 3674-3676.
- Cordeiro, J. M., Kaehler, M., Souza, G., & Felix, L. P. 2017. Karyotype analysis in Bignoniaceae (Bignoniaceae): chromosome numbers and heterochromatin. *Anais da Academia Brasileira de Ciências*, 89(4), 2697-2706.
- Costa, S. L., Brito, I. J. N., Lohmann, L. G., & de Melo, J. I. M. 2019. New records of the tribe Bignoniaceae (Bignoniaceae) for Paraíba state, northeastern Brazil. *Acta Brasiliensis*, 3(3), 89-96.
- Crawley, M. J. 2007. *The R Book*. London, John Wiley & Sons Ltd..
- Darwin, C. 1875. *The movements and habits of climbing plants*. J. Murray, London, UK.
- De Storme, N., & Geelen, D. 2011. The Arabidopsis mutant jason produces unreduced first division restitution male gametes through a parallel/fused spindle mechanism in meiosis II. *Plant physiology*, 155(3), 1403-1415.
- Dos Santos, G. M. A. 1995. Wood anatomy, chloroplast DNA, and flavonoids of the tribe Bignoniaceae (Bignoniaceae). PhD thesis, University of Reading, Reading, UK.
- Duarte, M. O., Mendes-Rodrigues, C., Alves, M. F., Oliveira, P. E., & Sampaio, D. S. 2017. Mixed pollen load and late-acting self-incompatibility flexibility in Adenocalymma

- peregrinum (Miers) LG Lohmann (Bignoniaceae: Bignoniaceae). *Plant Biology*, 19(2), 140-146.
- Eriksson, M. E., Israelsson, M., Olsson, O., & Moritz, T. 2000. Increased gibberellin biosynthesis in transgenic trees promotes growth, biomass production and xylem fiber length. *Nature biotechnology*, 18(7), 784.
- Espinosa-Ruiz, A., Saxena, S., Schmidt, J., Mellerowicz, E., Miskolczi, P., Bakó, L., & Bhalerao, R. P. 2004. Differential stage-specific regulation of cyclin-dependent kinases during cambial dormancy in hybrid aspen. *The Plant Journal*, 38(4), 603-615.
- Evert, R. F. 2006. *Esau's plant anatomy: meristems, cells, and tissues of the plant body: their structure, function, and development*. John Wiley & Sons.
- Ewers, F. W., Ewers, J. M., Jacobsen, A. L., & López-Portillo, J. 2007. Vessel redundancy: modeling safety in numbers. *Iawa Journal*, 28(4), 373-388.
- Ewers, F. W., & Fisher, J. B. 1989. Variation in vessel length and diameter in stems of six tropical and subtropical lianas. *American Journal of Botany*, 76(10), 1452-1459.
- Ewers, F. W., Fisher, J. B., & Chiu, S. T. 1990. A survey of vessel dimensions in stems of tropical lianas and other growth forms. *Oecologia*, 84(4), 544-552.
- Ewers, F. W., Fisher, J. B., Fichtner, K. 1991. Water flux and xylem structure in vines. In Putz, F. E. & Mooney, H. A. (eds.). *The Biology of Vines*. Cambridge University Press, Cambridge.
- Felipo-Benavent, A., Úrbez, C., Blanco-Touriñán, N., Serrano-Mislata, A., Baumberger, N., Achard, P., ... & Alabadí, D. 2018. Regulation of xylem fiber differentiation by gibberellins through DELLA-KNAT1 interaction. *Development*, 145(23).
- Fonseca, L. H. M., Cabral, S. M., Agra, M. D. F., & Lohmann, L. G. 2017. Taxonomic revision of *Dolichandra* (Bignoniaceae, Bignoniaceae). *Phytotaxa*, 301(1), 1-70.
- Fujiyama, K., Hino, T., Kanadani, M., Watanabe, B., Lee, H. J., Mizutani, M., & Nagano, S. 2019. Structural insights into a key step of brassinosteroid biosynthesis and its inhibition. *Nature plants*, 5(6), 589.
- Gallenmüller, F., Müller, U., Rowe, N., & Speck, T. 2001. The Growth Form of *Croton pullei* (Euphorbiaceae)-Functional Morphology and Biomechanics of a Neotropical Liana. *Plant Biology*, 3(1), 50-61.
- Gartner, B. L. 1991a. Relative growth rates of vines and shrubs of western poison oak, *Toxicodendron diversilobum* (Anacardiaceae). *American Journal of Botany*, 78(10), 1345-1353.
- Gartner, B. L. 1995. Patterns of xylem variation within a tree and their hydraulic and mechanical consequences. In *Plant stems* (pp. 125-149). Academic Press.
- Gentry, A. H. 1973. Generic delimitations of central American Bignoniaceae. *Brittonia*, 25(3), 226-242.

- Gentry, A. H. 1980. Bignoniaceae: Part I (Crescentieae and Tourrettieae). *Flora Neotropica*, 25(1), 1-130.
- Gentry, A. H. 1986. Species richness and floristic composition of Chocó region plant communities. *Caldasia*, 71-91.
- Gentry, A. H. 1991. The distribution and evolution of climbing plants. In Putz, F. E. & Mooney, H. A. (eds.). *The Biology of Vines*. Cambridge University Press, Cambridge.
- Gerolamo, C. S., & Angyalossy, V. 2017. Wood anatomy and conductivity in lianas, shrubs and trees of Bignoniaceae. *IAWA Journal*, 38(3), 412-432.
- Gray-Mitsumune, M., Blomquist, K., McQueen-Mason, S., Teeri, T. T., Sundberg, B., & Mellerowicz, E. J. 2008. Ectopic expression of a wood-abundant expansin PttEXPA1 promotes cell expansion in primary and secondary tissues in aspen. *Plant biotechnology journal*, 6(1), 62-72.
- Gray-Mitsumune, M., Mellerowicz, E. J., Abe, H., Schrader, J., Winzéli, A., Sterky, F., ... & Sundberg, B. 2004. Expansins abundant in secondary xylem belong to subgroup A of the α -expansin gene family. *Plant Physiology*, 135(3), 1552-1564.
- Groover, A. T. 2005. What genes make a tree a tree?. *Trends in plant science*, 10(5), 210-214.
- Haas, B. J., Papanicolaou, A., Yassour, M., Grabherr, M., Blood, P. D., Bowden, J., ... & MacManes, M. D. 2013. De novo transcript sequence reconstruction from RNA-seq using the Trinity platform for reference generation and analysis. *Nature protocols*, 8(8), 1494.
- Hacke, U. G., Spicer, R., Schreiber, S. G., & Plavcová, L. 2017. An ecophysiological and developmental perspective on variation in vessel diameter. *Plant, cell & environment*, 40(6), 831-845.
- Hirakawa, Y., Kondo, Y., & Fukuda, H. 2010. TDIF peptide signaling regulates vascular stem cell proliferation via the WOX4 homeobox gene in Arabidopsis. *The Plant Cell*, 22(8), 2618-2629.
- Hu, R., Qi, G., Kong, Y., Kong, D., Gao, Q., & Zhou, G. 2010. Comprehensive analysis of NAC domain transcription factor gene family in *Populus trichocarpa*. *BMC plant biology*, 10(1), 145.
- Huang, S., Raman, A. S., Ream, J. E., Fujiwara, H., Cerny, R. E., & Brown, S. M. 1998. Overexpression of 20-oxidase confers a gibberellin-overproduction phenotype in Arabidopsis. *Plant physiology*, 118(3), 773-781.
- Huber, W., Carey, V. J., Gentleman, R., Anders, S., Carlson, M., Carvalho, B. S., ... & Gottardo, R. 2015. Orchestrating high-throughput genomic analysis with Bioconductor. *Nature methods*, 12(2), 115.
- IAWA Committee 1989. IAWA list of microscopic features for hardwood identification.
- Isnard, S., & Silk, W. K. 2009. Moving with climbing plants from Charles Darwin's time into the 21st century. *American Journal of Botany*, 96(7), 1205-1221.

- Israelsson, M., Sundberg, B., & Moritz, T. 2005. Tissue-specific localization of gibberellins and expression of gibberellin-biosynthetic and signaling genes in wood-forming tissues in aspen. *The Plant Journal*, 44(3), 494-504.
- Johansen, D.A. 1940. Plant microtechnique. McGraw - Hill Book Co. Inc., New York.
- Kaehler, M., Michelangeli, F. A., & Lohmann, L. G. 2019. Fine tuning the circumscription of *Fridericia* (Bignoniaceae, Bignoniaceae). *Taxon*, 68(4), 751-770.
- Kanno, Y., Hanada, A., Chiba, Y., Ichikawa, T., Nakazawa, M., Matsui, M., ... & Seo, M. 2012. Identification of an abscisic acid transporter by functional screening using the receptor complex as a sensor. *Proceedings of the National Academy of Sciences*, 109(24), 9653-9658.
- Kans, J. 2018. Entrez direct: E-utilities on the UNIX command line. In, Entrez Programming Utilities Help [Internet]. *National Center for Biotechnology Information (US)*. <https://www.ncbi.nlm.nih.gov/books/NBK179288/>. Published.
- Kim, D., Langmead, B., & Salzberg, S. L. 2015. HISAT: a fast spliced aligner with low memory requirements. *Nature methods*, 12(4), 357.
- Ko, J. H., Han, K. H., Park, S., & Yang, J. 2004. Plant body weight-induced secondary growth in *Arabidopsis* and its transcription phenotype revealed by whole-transcriptome profiling. *Plant Physiology*, 135(2), 1069-1083.
- Ko, J. H., Kim, W. C., & Han, K. H. 2009. Ectopic expression of MYB46 identifies transcriptional regulatory genes involved in secondary wall biosynthesis in *Arabidopsis*. *The Plant Journal*, 60(4), 649-665.
- Koops, P., Pelsler, S., Ignatz, M., Klose, C., Marrocco-Selden, K., & Kretsch, T. 2011. EDL3 is an F-box protein involved in the regulation of abscisic acid signalling in *Arabidopsis thaliana*. *Journal of experimental botany*, 62(15), 5547-5560.
- Kubo, M., Udagawa, M., Nishikubo, N., Horiguchi, G., Yamaguchi, M., Ito, J., ... & Demura, T. 2005. Transcription switches for protoxylem and metaxylem vessel formation. *Genes & development*, 19(16), 1855-1860.
- Kwon, Y., Kim, J. H., Nguyen, H. N., Jikumaru, Y., Kamiya, Y., Hong, S. W., & Lee, H. 2013. A novel *Arabidopsis* MYB-like transcription factor, MYBH, regulates hypocotyl elongation by enhancing auxin accumulation. *Journal of experimental botany*, 64(12), 3911-3922.
- Li, L., Xu, J., Xu, Z. H., & Xue, H. W. 2005). Brassinosteroids stimulate plant tropisms through modulation of polar auxin transport in *Brassica* and *Arabidopsis*. *The Plant Cell*, 17(10), 2738-2753.
- Li, X., Yang, Y., Yao, J., Chen, G., Li, X., Zhang, Q., & Wu, C. 2009. FLEXIBLE CULM 1 encoding a cinnamyl-alcohol dehydrogenase controls culm mechanical strength in rice. *Plant molecular biology*, 69(6), 685-697.

- Little, C. H. A., & Wareing, P. F. 1981. Control of cambial activity and dormancy in *Picea sitchensis* by indol-3-ylacetic and abscisic acids. *Canadian Journal of Botany*, 59(8), 1480-1493.
- Lohmann, L. G. 2006. Untangling the phylogeny of neotropical lianas (Bignoniaceae, Bignoniaceae). *American Journal of Botany*, 93(2), 304-318.
- Lohmann, L. G., Bell, C. D., Calió, M. F., & Winkworth, R. C. 2012. Pattern and timing of biogeographical history in the Neotropical tribe Bignoniaceae (Bignoniaceae). *Botanical Journal of the Linnean Society*, 171(1), 154-170.
- Lohmann, L. G., & Taylor, C. M. 2014. A new generic classification of tribe Bignoniaceae (Bignoniaceae) 1. *Annals of the Missouri Botanical Garden*, 99(3), 348-489.
- Lovisol, C., & Schubert, A. 1998. Effects of water stress on vessel size and xylem hydraulic conductivity in *Vitis vinifera* L. *Journal of experimental botany*, 49(321), 693-700.
- Lu, D., Wang, T., Persson, S., Mueller-Roeber, B., & Schippers, J. H. 2014. Transcriptional control of ROS homeostasis by KUODA1 regulates cell expansion during leaf development. *Nature communications*, 5, 3767.
- Luisi, A., Giovannelli, A., Traversi, M. L., Anichini, M., & Sorce, C. 2014. Hormonal responses to water deficit in cambial tissues of *Populus alba* L. *Journal of plant growth regulation*, 33(3), 489-498.
- MacMillan, C. P., Mansfield, S. D., Stachurski, Z. H., Evans, R., & Southerton, S. G. 2010. Fasciclin-like arabinogalactan proteins: specialization for stem biomechanics and cell wall architecture in *Arabidopsis* and *Eucalyptus*. *The Plant Journal*, 62(4), 689-703.
- Martre, P., Cochard, H., & Durand, J. L. 2001. Hydraulic architecture and water flow in growing grass tillers (*Festuca arundinacea* Schreb.). *Plant, Cell & Environment*, 24(1), 65-76.
- Martre, P., Durand, J. L., & Cochard, H. 2000. Changes in axial hydraulic conductivity along elongating leaf blades in relation to xylem maturation in tall fescue. *The New Phytologist*, 146(2), 235-247.
- Matsumoto-Kitano, M., Kusumoto, T., Tarkowski, P., Kinoshita-Tsujimura, K., Václavíková, K., Miyawaki, K., & Kakimoto, T. 2008. Cytokinins are central regulators of cambial activity. *Proceedings of the National Academy of Sciences*, 105(50), 20027-20031.
- Mauriat, M., & Moritz, T. 2009. Analyses of GA20ox-and GID1-over-expressing aspen suggest that gibberellins play two distinct roles in wood formation. *The Plant Journal*, 58(6), 989-1003.
- Mazur, E., Benková, E., & Friml, J. 2016. Vascular cambium regeneration and vessel formation in wounded inflorescence stems of *Arabidopsis*. *Scientific reports*, 6, 33754.
- Mazur, E., Kurczyńska, E. U., & Friml, J. 2014. Cellular events during interfascicular cambium ontogenesis in inflorescence stems of *Arabidopsis*. *Protoplasma*, 251(5), 1125-1139.

- McCarthy, R. L., Zhong, R., & Ye, Z. H. 2009. MYB83 is a direct target of SND1 and acts redundantly with MYB46 in the regulation of secondary cell wall biosynthesis in *Arabidopsis*. *Plant and Cell Physiology*, *50*(11), 1950-1964.
- Ménard, L., McKey, D., & Rowe, N. 2009. Developmental plasticity and biomechanics of treelets and lianas in *Manihot aff. quinquepartita* (Euphorbiaceae): a branch-angle climber of French Guiana. *Annals of Botany*, *103*(8), 1249-1259.
- Meyer, L., Diniz-Filho, J. A. F., Lohmann, L. G., Hortal, J., Barreto, E., Rangel, T., & Kissling, W. D. 2020. Canopy height explains species richness in the largest clade of Neotropical lianas. *Global Ecology and Biogeography*, *29*(1), 26-37.
- Muñiz, L., Minguet, E. G., Singh, S. K., Pesquet, E., Vera-Sirera, F., Moreau-Courtois, C. L., ... & Tuominen, H. 2008. ACAULIS5 controls *Arabidopsis* xylem specification through the prevention of premature cell death. *Development*, *135*(15), 2573-2582.
- Nakajima, K., Furutani, I., Tachimoto, H., Matsubara, H., & Hashimoto, T. 2004. SPIRAL1 encodes a plant-specific microtubule-localized protein required for directional control of rapidly expanding *Arabidopsis* cells. *The Plant Cell*, *16*(5), 1178-1190.
- Nieminen, K., Immanen, J., Laxell, M., Kauppinen, L., Tarkowski, P., Dolezal, K., ... & Bhalerao, R. 2008. Cytokinin signaling regulates cambial development in poplar. *Proceedings of the National Academy of Sciences*, *105*(50), 20032-20037.
- Nilsson, J., Karlberg, A., Antti, H., Lopez-Vernaza, M., Mellerowicz, E., Perrot-Rechenmann, C., ... & Bhalerao, R. P. 2008. Dissecting the molecular basis of the regulation of wood formation by auxin in hybrid aspen. *The Plant Cell*, *20*(4), 843-855.
- Nuruzzaman, M., Manimekalai, R., Sharoni, A. M., Satoh, K., Kondoh, H., Ooka, H., & Kikuchi, S. 2010. Genome-wide analysis of NAC transcription factor family in rice. *Gene*, *465*(1-2), 30-44.
- Obaton, M. 1960. *Les Lianes ligneuses a structure anormale des forêts denses d'Afrique occidentale: thèse...* Masson & Cie.
- Ohashi-Ito, K., & Fukuda, H. 2003. HD-Zip III homeobox genes that include a novel member, ZeHB-13 (*Zinnia*)/ATHB-15 (*Arabidopsis*), are involved in procambium and xylem cell differentiation. *Plant and Cell Physiology*, *44*(12), 1350-1358.
- Olson, M. E., Anfodillo, T., Rosell, J. A., Petit, G., Crivellaro, A., Isnard, S., ... & Castorena, M. 2014. Universal hydraulics of the flowering plants: vessel diameter scales with stem length across angiosperm lineages, habits and climates. *Ecology Letters*, *17*(8), 988-997.
- Pace, M. R., Alcantara, S., Lohmann, L. G., & Angyalossy, V. 2015a. Secondary phloem diversity and evolution in Bignoniaceae (Bignoniaceae). *Annals of botany*, *116*(3), 333-358.
- Pace, M. R., & Angyalossy, V. 2013. Wood anatomy and evolution: a case study in the Bignoniaceae. *International Journal of Plant Sciences*, *174*(7), 1014-1048.

- Pace, M. R., Lohmann, L. G., Angyalossy, V. 2009. The rise & evolution of the cambial variant in Bignoniaceae. *Evolution & Development*, 11, 465-479.
- Pace, M. R., Lohmann, L. G., Angyalossy, V. 2011. Evolution of disparity between the regular & variant phloem in Bignoniaceae (Bignoniaceae). *American Journal of Botany*, 98, 602-618.
- Pace, M. R., Lohmann, L. G., Olmstead, R. G., & Angyalossy, V. 2015b. Wood anatomy of major Bignoniaceae clades. *Plant Systematics and Evolution*, 301(3), 967-995.
- Pace, M. R., Zuntini, A. R., Lohmann, L. G., & Angyalossy, V. 2016. Phylogenetic relationships of enigmatic Sphingiphila (Bignoniaceae) based on molecular and wood anatomical data. *Taxon*, 65(5), 1050-1063.
- Patro, R., Duggal, G., Love, M. I., Irizarry, R. A., & Kingsford, C. 2017. Salmon provides fast and bias-aware quantification of transcript expression. *Nature methods*, 14(4), 417.
- Petit, G., & Anfodillo, T. 2009. Plant physiology in theory and practice: an analysis of the WBE model for vascular plants. *Journal of Theoretical Biology*, 259(1), 1-4.
- Petit, G., & Anfodillo, T. 2011. Comment on “The blind men and the elephant: the impact of context and scale in evaluating conflicts between plant hydraulic safety and efficiency” by Meinzer et al.(2010). *Oecologia*, 165(2), 271-274.
- Pinheiro, J., Bates, D., DebRoy, S., Sarkar, D., Heisterkamp, S., Van Willigen, B., & Maintainer, R. 2017. Package ‘nlme’. *Linear and Nonlinear Mixed Effects Models, version*, 3-1.
- Poorter, L., McDonald, I., Alarcón, A., Fichtler, E., Licona, J. C., Peña-Claros, M., ... & Sass-Klaassen, U. 2010. The importance of wood traits and hydraulic conductance for the performance and life history strategies of 42 rainforest tree species. *New phytologist*, 185(2), 481-492.
- Putz, F. E. 1984. The natural history of lianas on Barro Colorado Island, Panama. *Ecology*, 65(6), 1713-1724.
- Quast, C., Pruesse, E., Yilmaz, P., Gerken, J., Schweer, T., Yarza, P., ... & Glöckner, F. O. 2012. The **SILVA** ribosomal RNA gene database project: improved data processing and web-based tools. *Nucleic acids research*, 41(1), 590-596.
- R Core Team 2013. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.
- Rajput, K. S., Raole, V. M., & Gandhi, D. 2008. Radial secondary growth and formation of successive cambia and their products in *Ipomoea hederifolia* L.(Convolvulaceae). *Botanical journal of the Linnean Society*, 158(1), 30-40.
- Robinson, M., McCarthy, D., Chen, Y., & Smyth, G. K. 2010. edgeR: differential expression analysis of digital gene expression data. *J. Hosp. Palliat. Nurs*, 4, 206-207.
- Robinson, M. D., & Oshlack, A. 2010. A scaling normalization method for differential expression analysis of RNA-seq data. *Genome biology*, 11(3), 25.

- Robinson, M. D. & Smyth, G. K. 2008. Small-sample estimation of negative binomial dispersion, with applications to SAGE data. *Biostatistics*, 9, 21–332.
- Robischon, M., Du, J., Miura, E., & Groover, A. 2011. The Populus class III HD ZIP, popREVOLUTA, influences cambium initiation and patterning of woody stems. *Plant physiology*, 155(3), 1214-1225.
- Rowe, N. P. & Speck, T. 1996. Biomechanical characteristics of the ontogeny & growth habit of the tropical liana *Condylocarpon guianense* (Apocynaceae). *International Journal of Plant Sciences*, 157,406-417.
- Sachs, T. 1981. The control of the patterned differentiation of vascular tissues. In *Advances in botanical research*. Academic Press, 151-262.
- Saez, A., Apostolova, N., Gonzalez-Guzman, M., Gonzalez-Garcia, M. P., Nicolas, C., Lorenzo, O., & Rodriguez, P. L. 2004. Gain-of-function and loss-of-function phenotypes of the protein phosphatase 2C HAB1 reveal its role as a negative regulator of abscisic acid signalling. *The Plant Journal*, 37(3), 354-369.
- Sahni, S., Prasad, B. D., Liu, Q., Grbic, V., Sharpe, A., Singh, S. P., & Krishna, P. 2016. Overexpression of the brassinosteroid biosynthetic gene DWF4 in Brassica napus simultaneously increases seed yield and stress tolerance. *Scientific reports*, 6, 28298.
- Sanchez-Moran, E., Osman, K., Higgins, J. D., Pradillo, M., Cunado, N., Jones, G. H., & Franklin, F. C. H. 2008. ASY1 coordinates early events in the plant meiotic recombination pathway. *Cytogenetic and genome research*, 120(3-4), 302-312.
- Scarpella, E., Marcos, D., Friml, J., & Berleth, T. 2006. Control of leaf vascular patterning by polar auxin transport. *Genes & development*, 20(8), 1015-1027.
- Schenck, H. 1893. Beiträge zur Biologie und Anatomie der Lianen im Besonderen der in Brasilien einheimischen Arten. In Schimper, A. F. W. (ed.). Beiträge zur Anatomie der Lianen: Botanische Mittheilungen aus den Tropen. G Fischer, Jena.
- Schneider, C. A., Rasband, W. S., & Eliceiri, K. W. 2012. NIH Image to ImageJ: 25 years of image analysis. *Nature methods*, 9(7), 671.
- Scholz, A., Klepsch, M., Karimi, Z., & Jansen, S. 2013. How to quantify conduits in wood?. *Frontiers in Plant Science*, 4, 56.
- Smetana, O., Mäkilä, R., Lyu, M., Amiryousefi, A., Rodriguez, F. S., Wu, M. F., ... & Roszak, P. 2019. High levels of auxin signalling define the stem-cell organizer of the vascular cambium. *Nature*, 565(7740), 485.
- Smyth, D. R. 2016. Helical growth in plant organs: mechanisms and significance. *Development*, 143(18), 3272-3282.
- Snow, R. 1935. Activation of cambial growth by pure hormones. *New Phytologist*, 34(5), 347-360.

- Soler, M., Camargo, E. L. O., Carocha, V., Cassan-Wang, H., San Clemente, H., Savelli, B., ... & Grima-Pettenati, J. 2015. The Eucalyptus grandis R2R3-MYB transcription factor family: evidence for woody growth-related evolution and function. *New Phytologist*, 206(4), 1364-1377.
- Sousa-Baena, M. S., Lohmann, L. G., Rossi, M., & Sinha, N. R. 2014a. Acquisition and diversification of tendrilled leaves in Bignoniaceae (Bignoniaceae) involved changes in expression patterns of SHOOTMERISTEMLESS (STM), LEAFY/FLORICAULA (LFY/FLO), and PHANTASTICA (PHAN). *New Phytologist*, 201(3), 993-1008.
- Sousa-Baena, M. S., Sinha, N. R., & Lohmann, L. G. 2014b. Evolution and Development of Tendrils in Bignoniaceae (Lamiales, Bignoniaceae) 1. *Annals of the Missouri Botanical Garden*, 99(3), 323-348.
- Sperry, J. S., Meinzer, F. C., & McCULLOH, K. A. 2008. Safety and efficiency conflicts in hydraulic architecture: scaling from tissues to trees. *Plant, Cell & Environment*, 31(5), 632-645.
- Sperry, J. S., Smith, D. D., Savage, V. M., Enquist, B. J., McCulloh, K. A., Reich, P. B., ... & von Allmen, E. I. 2012. A species-level model for metabolic scaling in trees I. Exploring boundaries to scaling space within and across species. *Functional Ecology*, 26(5), 1054-1065.
- Steppe, K., & Lemeur, R. 2007. Effects of ring-porous and diffuse-porous stem wood anatomy on the hydraulic parameters used in a water flow and storage model. *Tree physiology*, 27(1), 43-52.
- Sundell, D., Street, N. R., Kumar, M., Mellerowicz, E. J., Kucukoglu, M., Johnsson, C., ... & Tuominen, H. 2017. AspWood: high-spatial-resolution transcriptome profiles reveal uncharacterized modularity of wood formation in *Populus tremula*. *The Plant Cell*, 29(7), 1585-1604.
- Tamaio, N., Joffily, A., Braga, J. M. A., & Rajput, K. S. 2010. Stem anatomy and pattern of secondary growth in some herbaceous vine species of Menispermaceae. *The Journal of the Torrey Botanical Society*, 137(2), 157-166.
- Taylor, N. G., Howells, R. M., Huttly, A. K., Vickers, K., & Turner, S. R. 2003. Interactions among three distinct CesA proteins essential for cellulose synthesis. *Proceedings of the National Academy of Sciences*, 100(3), 1450-1455.
- Thode, V. A., Sanmartín, I., & Lohmann, L. G. 2019. Contrasting patterns of diversification between Amazonian and Atlantic forest clades of Neotropical lianas (Amphilophium, Bignoniaceae) inferred from plastid genomic data. *Molecular phylogenetics and evolution*, 133, 92-106.

- Torres, C. A., Zamora, C. M. P., Nuñez, M. B., & Gonzalez, A. M. 2018. In vitro antioxidant, antilipoxygenase and antimicrobial activities of extracts from seven climbing plants belonging to the Bignoniaceae. *Journal of integrative medicine*, 16(4), 255-262.
- Torrey, J. G., & Loomis, R. S. 1967. Auxin-cytokinin control of secondary vascular tissue formation in isolated roots of *Raphanus*. *American Journal of Botany*, 54(9), 1098-1106.
- Tuominen, H., Puech, L., Fink, S., & Sundberg, B. 1997. A radial concentration gradient of indole-3-acetic acid is related to secondary xylem development in hybrid aspen. *Plant Physiology*, 115(2), 577-585.
- Tyree, M. T., Davis, S. D., & Cochard, H. 1994. Biophysical perspectives of xylem evolution: is there a tradeoff of hydraulic efficiency for vulnerability to dysfunction?. *IAWA journal*, 15(4), 335-360.
- Tyree, M. T., & Ewers, F. W. 1991. The hydraulic architecture of trees and other woody plants. *New Phytologist*, 119(3), 345-360.
- Tyree, M. T., & Zimmermann, M. H. 2002. Hydraulic architecture of whole plants and plant performance. In *Xylem structure and the ascent of sap*. Springer, Berlin, Heidelberg. 175-214.
- Uggla, C., Moritz, T., Sandberg, G., & Sundberg, B. 1996. Auxin as a positional signal in pattern formation in plants. *Proceedings of the national academy of sciences*, 93(17), 9282-9286.
- Valdivia, E. R., Herrera, M. T., Gianzo, C., Fidalgo, J., Revilla, G., Zarra, I., & Sampedro, J. 2013. Regulation of secondary wall synthesis and cell death by NAC transcription factors in the monocot *Brachypodium distachyon*. *Journal of experimental botany*, 64(5), 1333-1343.
- Vanstraelen, M., Inzé, D., & Geelen, D. 2006. Mitosis-specific kinesins in *Arabidopsis*. *Trends in plant science*, 11(4), 167-175.
- Victorio MP. 2016. Roots and stems anatomy of Bignoniaceae: lianescent syndrome and secondary xylem. MSc dissertation, University of São Paulo, São Paulo, Brazil.
- Vieten, A., Vanneste, S., Wiśniewska, J., Benková, E., Benjamins, R., Beeckman, T., ... & Friml, J. 2005. Functional redundancy of PIN proteins is accompanied by auxin-dependent cross-regulation of PIN expression. *Development*, 132(20), 4521-4531.
- West, G. B., Brown, J. H., & Enquist, B. J. 1999. A general model for the structure and allometry of plant vascular systems. *Nature*, 400(6745), 664.
- Wang, Z., Gerstein, M., & Snyder, M. 2009. RNA-Seq: a revolutionary tool for transcriptomics. *Nature reviews genetics*, 10(1), 57.
- Wyka, T. P., Zadworny, M., Mucha, J., Żytkowiak, R., Nowak, K., & Oleksyn, J. 2019. Species-specific responses of growth and biomass distribution to trellis availability in three temperate lianas. *Trees*, 33(3), 921-932.

- Xu, P., Kong, Y., Song, D., Huang, C., Li, X., & Li, L. 2014. Conservation and functional influence of alternative splicing in wood formation of *Populus* and *Eucalyptus*. *BMC genomics*, 15(1), 780.
- Yamamoto, R., Demura, T., & Fukuda, H. 1997. Brassinosteroids induce entry into the final stage of tracheary element differentiation in cultured *Zinnia* cells. *Plant and Cell Physiology*, 38(8), 980-983.
- Yamamoto, R., Fujioka, S., Demura, T., Takatsuto, S., Yoshida, S., & Fukuda, H. 2001. Brassinosteroid levels increase drastically prior to morphogenesis of tracheary elements. *Plant Physiology*, 125(2), 556-563.
- Yang, C., Song, J., Ferguson, A. C., Klisch, D., Simpson, K., Mo, R., ... & Wilson, Z. A. 2017. Transcription factor MYB26 is key to spatial specificity in anther secondary thickening formation. *Plant physiology*, 175(1), 333-350.
- Yang, C., Xu, Z., Song, J., Conner, K., Barrena, G. V., & Wilson, Z. A. 2007. Arabidopsis MYB26/MALE STERILE35 regulates secondary thickening in the endothecium and is essential for anther dehiscence. *The Plant Cell*, 19(2), 534-548.
- Ye, Z. H., & Zhong, R. 2015. Molecular control of wood formation in trees. *Journal of experimental botany*, 66(14), 4119-4131.
- Yoshida, T., Nishimura, N., Kitahata, N., Kuromori, T., Ito, T., Asami, T., ... & Hirayama, T. 2006. ABA-hypersensitive germination3 encodes a protein phosphatase 2C (AtPP2CA) that strongly regulates abscisic acid signaling during germination among Arabidopsis protein phosphatase 2Cs. *Plant physiology*, 140(1), 115-126.
- Zamariola, L., De Storme, N., Vannerum, K., Vandepoele, K., Armstrong, S. J., Franklin, F. C. H., & Geelen, D. 2014. SHUGOSHIN s and PATRONUS protect meiotic centromere cohesion in *Arabidopsis thaliana*. *The Plant Journal*, 77(5), 782-794.
- Zhang, J., Eswaran, G., Alonso-Serra, J., Kucukoglu, M., Xiang, J., Yang, W., ... & Yun, J. Y. 2019. Transcriptional regulatory framework for vascular cambium development in Arabidopsis roots. *Nature plants*, 5(10), 1033-1042.
- Zhang, Q., Luo, F., Zhong, Y., He, J., & Li, L. 2019. Modulation of NST1 activity by XND1 regulates secondary cell wall formation in *Arabidopsis thaliana*. *Journal of experimental botany*.
- Zhang, J., Nieminen, K., Serra, J. A. A., & Helariutta, Y. 2014. The formation of wood and its control. *Current opinion in plant biology*, 17, 56-63.
- Zhao, C., Avci, U., Grant, E. H., Haigler, C. H., & Beers, E. P. 2008. XND1, a member of the NAC domain family in *Arabidopsis thaliana*, negatively regulates lignocellulose synthesis and programmed cell death in xylem. *The Plant Journal*, 53(3), 425-436.
- Zhbannikov, I. Y., Hunter, S. S., Foster, J. A., & Settles, M. L. 2017. SeqyClean: a pipeline for high-throughput sequence data preprocessing. In *Proceedings of the 8th ACM International*

- Conference on Bioinformatics, Computational Biology, and Health Informatics* (pp. 407-416). ACM.
- Zhong, R., Demura, T., & Ye, Z. H. 2006. SND1, a NAC domain transcription factor, is a key regulator of secondary wall synthesis in fibers of Arabidopsis. *The Plant Cell*, 18(11), 3158-3170.
- Zhong, R., Lee, C., McCarthy, R. L., Reeves, C. K., Jones, E. G., & Ye, Z. H. 2011. Transcriptional activation of secondary wall biosynthesis by rice and maize NAC and MYB transcription factors. *Plant and Cell Physiology*, 52(10), 1856-1871.
- Zhong, R., Lee, C., & Ye, Z. H. 2010. Functional characterization of poplar wood-associated NAC domain transcription factors. *Plant Physiology*, 152(2), 1044-1055.
- Ziemińska, K., Butler, D. W., Gleason, S. M., Wright, I. J., & Westoby, M. 2013. Fibre wall and lumen fractions drive wood density variation across 24 Australian angiosperms. *AoB Plants*, 5.
- Ziemińska, K., Westoby, M., & Wright, I. J. 2015. Broad anatomical variation within a narrow wood density range—a study of twig wood across 69 Australian angiosperms. *PLoS One*, 10(4), e0124892.
- Zimmermann, M. H. 1983. Xylem structure and the ascent of sap. Springer, Berlin.
- Zinkgraf, M., Gerttula, S., & Groover, A. 2017. Transcript profiling of a novel plant meristem, the monocot cambium. *Journal of integrative plant biology*, 59(6), 436-449.