University of São Paulo "Luiz de Queiroz" College of Agriculture

Semiochemicals involved in the sugarcane-*Diatraea saccharalis*-opportunistic fungi interaction

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Thesis presented to obtain the degree of Doctor in Science. Area: Entomology

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RESUMO

Semioquímicos envolvidos na interação cana-de-açúcar-*Diatraea saccharalis*-fungos oportunistas

Interações entre planta, inseto e microrganismos no ambiente revela uma miríade de simbioses estratégicas. Por exemplo, semioquímicos (i.e. feromônios e voláteis de plantas) têm mostrado um papel decisivo nessas interações associadas a metabólitos voláteis e não voláteis. As alterações dos semioquímicos induzidas por microrganismos resultam em uma maior atração de insetos vetores para a planta, ou mudanças no comportamento sexual dos insetos e, portanto, aumento na dispersão de patógenos, comumente relatado em interações envolvendo bactérias, vírus e arqueas. Diferentemente dos patógenos transmitidos por vetores, os patógenos oportunistas presumivelmente não têm uma associação íntima com o inseto. Fusarium verticillioides e Colletotrichum falcatum são a muito tempo reportados como fungos oportunistas, utilizando das aberturas deixadas pela herbivoria de lagartas de Diatraea saccharalis para infectar a planta. No entanto, os semioquímicos envolvidos nesta interação ainda são desconhecidos. Diante do exposto, a presente pesquisa de doutorado aborda investigações que visam melhor compreender os semioquímicos envolvidos na interação cana-de-açúcar-D. sacharalis-fungos oportunistas. Aqui estabelecemos um novo papel para a associação inseto-fungo-cana-de-açúcar. Inicialmente, abordamos a identificação química da infecção por fungos nas defesas das plantas e a escolha da preferência do hospedeiro e, consequentemente, os benefícios do hospedeiro para a prole. Avaliamos o papel de D. saccharalis como vetor de disseminação do fungo F. verticillioides, e os possíveis efeitos da alimentação do fungo na biologia e no sucesso reprodutivo de D. saccharalis. Abordamos especificamente se o fungo modifica o comportamento de D. saccharalis de forma a aumentar a probabilidade do fungo ser disseminado por insetos contaminados. Enquanto C. falcatum mostrou um efeito neutro na associação com D. saccharalis, curiosamente, nossos resultados mostraram que os voláteis de plantas infectadas com F. verticillioides atraem fêmeas de D. saccharalis para oviposição. Além disso, o fungo F. verticillioides é transmitido verticalmente à sua descendência, que continua o ciclo inoculando o fungo em plantas saudáveis. As fêmeas contaminadas pelo fungo preferem colocar seus ovos em plantas mock quando comparadas a plantas infectadas pelo fungo F. verticillioides. Em seguida, nos concentramos na interação envolvendo apenas D. saccharalis e F. verticillioides. Nossos dados demonstram que F. verticillioides manipula tanto a planta hospedeira quanto o inseto herbívoro ao longo do ciclo de vida, para promover sua infecção e disseminação, formando associações mutualísticas, mas inerentemente egoístas, com a planta e inseto.

Palavras-chave: Interação inseto-planta-microorganismo, *Fusarium verticillioides*, *Colletotrichum falcatum*, Voláteis de plantas, Feromônio

ABSTRACT

Semiochemicals involved in the sugarcane-*Diatraea saccharalis*-opportunistic fungi interaction

The interaction between plant, insect and microorganisms in environment reveal myriad strategies of symbioses. For instance, semiochemicals (i.e., pheromone and plant volatiles) have played a decisive role in these interactions associated with volatile and non-volatile metabolites. The changes in semiochemicals induced by microorganisms result in the increased attraction of insect vectors to the plant, or changes in sexual behavior of insects and, hence, to increased pathogen dispersal, commonly reported to bacteria, viruses, and archaea interactions. Different from vector-borne pathogens, opportunistic pathogens presumably do not have an intimate association with the insect. It has long been assumed that Fusarium verticillioides and Colletotrichum falcatum are opportunistic fungi, where it takes advantage of the openings left by Diatraea saccharalis attack to infect the plant. However, the semiochemicals involved in this interaction are still largely unknown. Given the above, the present doctoral research approaches investigations to better understand the semiochemicals involved in sugarcane-D. saccharalis-opportunistic fungi interaction. Here, we establish a new paradigm for the insect-fungi-sugarcane association. Initially, we addressed the chemical identification of fungi infection in plant defenses, host preference choice, and offspring benefits. We also evaluated the role of D. saccharalis as a dissemination vector for F. verticillioides and the possible effects of F. verticillioides feeding on D. saccharalis biology and reproductive success. Finally, we specifically tackled whether the fungus modifies D. saccharalis behavior in ways that increase the likelihood of the fungus being disseminated by contaminated insects. Interestingly, our results showed that volatile emissions from F. verticillioides-infected plants attract D. saccharalis adults to lay eggs while C. falcatum has a neutral effect against D. saccharalis. The F. verticillioides is transmitted vertically to their offspring, which continues the cycle by inoculating the fungus into healthy plants. Then females carrying the fungus prefer to lay their eggs on mock plants than fungus-infected plants. Lather, we focus on D. saccharalis-F. verticillioides interaction. Our data demonstrate that F. verticillioides manipulate both the host plant and insect herbivore across the life cycle to promote its infection and dissemination, forming mutualistic but inherently selfish alliances with plants and insects.

Keywords: Insect-plant-microorganism interaction, *Fusarium verticillioides*, *Colletotrichum falcatum*, Plant volatile, Pheromone

1. GENERAL INTRODUCTION

What drives insects for mating partners, food sources, host preference for oviposition sites, and avoid dangers in their environment have fascinated scientists for centuries. Semiochemicals¹ cues play a decisive role in these ecological contexts mediate insect perception of different types of odors, the volatile organic compounds (VOCs), presents in nature by other species or by conspecifics, presumably the driving force behind the evolution often complex olfactory system.

The interactions between insects and semiochemicals in the environment range from the specificity of sexual pheromones starting with the first VOC chemically recognized, bombykol, the *Bombyx mori* sex pheromone (Butenandt et al. 1959). However, the modulation of insect landscape results from dynamic interactions throughout life, including environment and multitrophic interactions. In fact, it is essential to point out that, within the evolutionary process, the plant factor plays an essential role in insect multitrophic interactions. The constitutive plant VOCs have been driving herbivores to avoid or chose the host to feed or oviposition site (Hanson and Dethier 1973; Visser 1986; Pare and Tumlinson 1999; Dicke 2000). Researchers lather found that the plants emit constitutively and *de noro* blends of volatiles in responses to herbivory, identified as herbivore-induced plant volatiles (HIPVs), constitute important cues for parasitoids and predators to find prey or hosts and plant-plant communication (Baldwin and Schultz 1983; Turlings et al. 1990; Dicke and Baldwin 2010; Dicke 2016; Naranjo-Guevara et al. 2021; Silva et al. 2021). Recently 'microbe-induced plant volatiles (MIPVs) have been considered in insect associations.

In general, relationships among microorganisms and insects are widespread, and this interaction's dynamics depend on their costs and benefits to hosts. If the benefits of interaction are consistently sturdy and mutualistic with beneficial effects on the hosts, selection should favor hosts and lead to fixation of the symbiosis; however, the antagonistic interactions, with adverse or no apparent effects on the hosts, can reduce the success of interaction and be lost (Batra 2016; Biedermann and Vega 2020; Hosokawa and Fukatsu 2020).

Since insects generally lack the capacity to synthesize sterols, which are the precursors for different hormones, they acquire either sterols or sterol precursors associated with microorganisms, include the cases of fungal cultivation by insects as ambrosia beetles (Morales-Ramos et al. 2000; Cardoza et al. 2003). Thus, the attraction of insects to infected-plant can be beneficial for insects and microorganisms, which that mothers choose the best oviposition sites for their offspring by superior nutritional quality for insect development through pathogens infection according to preference–performance hypothesis ("The mother know best") (Jaenike 1978, Thompson 1988).

On the other hand, phytopathogens take advantage of insect hosts to increased pathogen dispersal. This symbiotic mode of transmission can be horizontal, in which hosts contagiously acquire symbionts through an environmental source or, vertically, through parents inheritance to offspring; however, as there is a great diversity of organisms, this transmission also it can be mixed, that is, horizontally and vertically (Bright and Bulgheresi 2010).

The orchestration of these phytopathogens to be transmitted by insects is fascinating and involves myriad strategies to manipulate plant and insect vectors to their own benefit, including alterations of volatile and non-volatile host metabolites. This change in MIPVs release after host–plant infection leads to a strategy to attract insects, leading to increased pathogen dispersal by insect vectors according to the "host manipulation hypothesis" (Heil 2016; Mauck et al. 2018). The manipulation of insect behavior by microorganisms can change the insect host preference, making

¹ Semio= Greek $\sigma\eta\mu\epsilon$ iov (semeion), signal. Chemical signals (categorized as either pheromones or allelochemicals) that are recognized and modify the behavior of the recipient organism.

the vector move to healthy plants to pinpointing the location of pathogen dissemination, very well reported in viruses and bacteria interactions (Fereres and Moreno 2009; Ingwell et al. 2012; Mann et al. 2012; Keesey et al. 2017). For example, the *Barley yellow dwarf luteovirus* (BYDV) increase the quantity of volatiles, especially (Z)-3-hexenyl acetate in wheat plants, attracting virus-free aphids, in contrasting, virus-containing aphids prefer healthy plants, thereby resulting in higher virus uptake and spreading (Ingwell et al. 2012). Also, the *Cucumber mosaic virus* (CMV), a nonpersistent virus, elevates the overall emission of volatiles in Cucurbita pepo attracting *Myzus persicae* and *Aphis gossypii* virus vector. However, these infected plants are less palatable to virus vectors, and the vector dashes between plants spreading the virus (Mauck et al. 2010).

Different from vector-borne pathogens, opportunistic pathogens presumably do not have an intimate association with the insect. In sugarcane, infection by the fungi phytopathogens *Fusarium verticillioides* (pokkah boeng causal agent) and *Colletotrichum falcatum* (red rot causal agent) are widespread in areas of sugarcane. Even the disease can occur in the absence of borer-feeding insects (usually associated with crop damages or in endophytic association, entering it through a lesion in its structure), usually occurs associated with *Diatraea saccharalis*, the sugarcane-*D. saccharalis*-opportunistic fungi interaction (Ogunwolu et al. 1991; Franco et al. 2017). Colonization by these fungi promotes the inversion of sucrose into glucose and the disruption of water and nutrient transport, which are essential to produce sugars. Hitherto, it was assumed that the fungi take advantage of the openings made by the borer to penetrate the sugarcane stalk and infect the plant (Ogunwolu et al. 1991; Franco et al. 2017).

In previous studies, the sugarcane-D. saccharalis-opportunistic fungi interaction has been elucidated. Medeiros et al. (2012) and Franco and Santiago (2014) show that after the attack of D. saccharalis, herbivory strongly induced the genes encoding the plant expresses antifungal defense proteins (Sugarwin1 and Sugarwin2). Interestingly enough, contrary to the initial hypothesis, these proteins do not interfere in insects' development; however, they cause both fungal C. falcatum and F. verticillioides, death, but cannot affect Aspergillus nidulans, a non-sugarcane pathogenic fungus. Thus, the plant seems to predict pathogen infection as early as the first signs of insect herbivory protect the plant against opportunistic pathogens. Also, F. verticillioides-sugarcane infected release less attractive herbivore-induced plant VOCs to Cotesia flavipes, a parasitoid of D. saccharalis larvae (Peñaflor and Bento 2019), indicating an indirect benefit to the herbivore of co-occurring with the fungus as a strategy of feeding in a safe environment.

This close association between opportunistic fungi and the sugarcane borer led us to investigate the mechanisms of these interactions. In fact, in this dissertation, Franco (2017) shows that *C. falcatum* and *F. verticillioides* VOCs emission attract *D. saccharalis* larvae. Also, the fungi positively influencing the diet of *D. saccharalis* and larvae weight gain in diets supplemented with fungi. Chapter 2 focuses on *D. saccharalis* adult interaction among plants infected by opportunistic fungi, *C. falcatum*-infected plants, *F. verticillioides*-infected plants, and *C. falcatum+F. verticillioides*-infected plants. First, using chromatography tolls, we addressed the chemical identification of plant defenses induced by fungi infection. Also, based on the theory of preference–performance hypothesis, we presented insights of insect behavior into oviposition preferences and consequently offspring benefits in infected plants. In concomitance, Gallan (2019), studying the possible role of *D. saccharalis* as a vector of *C. falcatum* and *F. verticillioides*, showed that, in fact, at least *F. verticillioides* was vertically transmitted to *D. saccharalis* offspring. Then, still in Chapter 2, using a mutant *F. verticillioides* (*Fr:DsRed*) tracked by laser confocal microscopy, we also explicitly evaluated Koch's postulates, *in vivo*, to confirm the vertical transmission and role of *D. saccharalis* on *F. verticillioides* infection in the sugarcane plant. Also, we evaluated the possible effects of *F. verticillioides* feeding on the biology and reproductive success of *D. saccharalis*. Finally, using electroantennography tolls and olfactory behavior assays, we specifically tackled whether the fungus modifies *D. saccharalis* behavior to increase the likelihood of the fungus being disseminated by the contaminated insect,

corroborating with the "host manipulation hypothesis". The data of the three dissertations were combined and partially redrafted in Franco et al. (2021).

To make a four-year research story short, we showed the interaction in the following way (Figure 1): *D. saccharalis* moths orients towards volatiles emitted by infected plants, which is the preferred host for oviposition (1). When the larvae hatch, they penetrate the stems and are contaminated by the fungus (2). When they enter the pupal stage and develop into adult moths, they are already carriers of the fungus. Remarkably, the host preference rather than plastic changes whether insects themselves are carriers of the fungus. Instrumentalized by the VOCs, females not carrying *F. verticillioides* prefer to lay their eggs on fungus-infected plants, while females carrying the *F. verticillioides* prefer to lay their eggs on fungus-infected plants, while females by *D. saccharalis* vertical transmission through its offspring (4). In this scenario, our study indicates that *F. verticillioides* is not just an opportunistic fungus but also more intimately associated with an insect herbivore, manipulating the host plant and insect herbivore across the life cycle to promote its infection and dissemination.

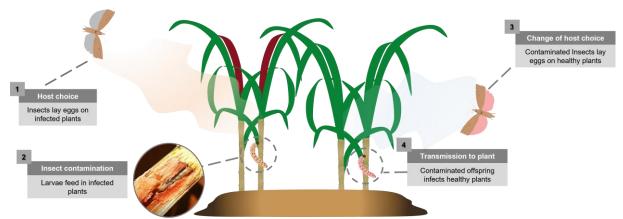


Figure 1. Semiochemicals involved in sugarcane-*Diatraea saccharallis*-opportunistic fungus interaction. Plants infected by *Fusarium verticillioides* emit VOCs (pink shadow) attractive to *D. saccharallis* moths to lay eggs (1). When the larvae hatch, they penetrate the stems, feed the contaminated plant, and are themselves contaminated by *F. verticillioides* (2). Females carrying the *F. verticillioides* (pink females) are attracted by VOCs emitted by healthy plants (light green shadow) to lay their eggs (3). The offspring from genitors fed the fungus (pink larvae) transmit the *F. verticillioides* to a new healthy plant (4).

- Baldwin IT, Schultz JC (1983) Rapid changes in tree leaf chemistry induced by damage: Evidence for communication between plants. Science (80-) 221:277–279. https://doi.org/10.1126/science.221.4607.277
- Batra LR (2016) Mycological Society of America Review Authors (s): Lekh R. Batra Review by: Lekh R. Batra Published by: Mycological Society of America Stable URL: http://www.jstor.org/stable/3757184 Accessed: 30-03-2016 14:57 UTC Your use of the JSTOR archive i. 60:466–469
- Biedermann PHW, Vega FE (2020) Annual Review of Entomology Ecology and Evolution of Insect Fungus Mutualisms. Annu Rev Entomol 65:22.1-22.25
- Bright M, Bulgheresi S (2010) A complex journey: Transmission of microbial symbionts. Nat Rev Microbiol 8:218–230. https://doi.org/10.1038/nrmicro2262
- Cardoza YJ, Teal PEA, Tumlinson JH (2003) Effect of peanut plant fungal infection on oviposition preference by *Spodoptera exigua* and on host-searching behavior by *Cotesia marginiventris*. Environ Entomol 32:970–976. https://doi.org/10.1603/0046-225X-32.5.970
- Dicke M (2000) Chemical ecology of host-plant selection by herbivorous arthropods: A multitrophic perspective. Biochem Syst Ecol 28:601–617. https://doi.org/10.1016/S0305-1978(99)00106-4
- Dicke M (2016) Plant phenotypic plasticity in the phytobiome: A volatile issue. Curr Opin Plant Biol 32:17-23. https://doi.org/10.1016/j.pbi.2016.05.004
- Dicke M, Baldwin IT (2010) The evolutionary context for herbivore-induced plant volatiles: beyond the "cry for help." Trends Plant Sci 15:167–175. https://doi.org/10.1016/j.tplants.2009.12.002

- Fereres A, Moreno A (2009) Behavioural aspects influencing plant virus transmission by homopteran insects. Virus Res 141:158–168. https://doi.org/10.1016/j.virusres.2008.10.020
- Flatau R, Segoli M, Hawlena H (2021) Wolbachia endosymbionts of fleas occur in all females but rarely in males and do not show evidence of obligatory relationships, fitness effects, or sex-distorting manipulations. Front Microbiol 12:. https://doi.org/10.3389/fmicb.2021.649248
- Flávia FP (2017) University of São Paulo " Luiz de Queiroz " College of Agriculture Unraveling sugarcane- *Diatraea saccharalis* -opportunistic fungi interaction in sugarcane.
- Franco FP, Moura DS, Vivanco JM, Silva-Filho MC (2017) Plant–insect–pathogen interactions: a naturally complex ménage à trois. Curr Opin Microbiol 37:54–60. https://doi.org/10.1016/j.mib.2017.04.007
- Franco FP, Santiago AC, Henrique-Silva F, et al (2014) The sugarcane defense protein SUGARWIN2 causes cell death in Collectrichum falcatum but not in non-pathogenic fungi. PLoS One 9:. https://doi.org/10.1371/journal.pone.0091159
- Franco FP, Túler AC, Gallan DZ, et al (2021) Fungal phytopathogen modulates plant and insect responses to promote its dissemination. ISME J. https://doi.org/10.1038/s41396-021-01010-z
- Franco P, Santiago AC (2014) The sugarcane defense protein Sugarwin2 causes cell death in *Colletotrichum falcatum* but not in non- pathogenic fungi. https://doi.org/10.1371/journal.pone.0091159
- Gallan DZ (2019) Universidade de São Paulo Escola Superior de Agricultura "Luiz de Queiroz " Estudo da interação entre a broca da cana-de-açúcar *Diatraea saccharalis* (Lepidoptera: Crambidae) e fungos oportunistas *Colletotrichum falcatum* e Fusarium verticillioides Dieg. 81
- Hanson FE, Dethier VG (1973) Rôle of gustation and olfaction in food plant discrimination in the tobacco hornworm, Manduca sexta. J Insect Physiol 19:1019–1031. https://doi.org/10.1016/0022-1910(73)90028-0
- Heil M (2016) Host manipulation by parasites: Cases, patterns, and remaining doubts. Front Ecol Evol 4:. https://doi.org/10.3389/fevo.2016.00080
- Hosokawa T, Fukatsu T (2020) Relevance of microbial symbiosis to insect behavior. Curr Opin Insect Sci 39:91–100. https://doi.org/10.1016/j.cois.2020.03.004
- Ingwell LL, Eigenbrode SD, Bosque-Pérez NA (2012) Plant viruses alter insect behavior to enhance their spread. Sci Rep 2:. https://doi.org/10.1038/srep00578
- Jaenike J (1978) On optimal oviposition behavior in phytophagous insects. Theor Popul Biol 14:350–356. https://doi.org/10.1016/0040-5809(78)90012-6
- Keesey IW, Koerte S, Khallaf MA, et al (2017) Pathogenic bacteria enhance dispersal through alteration of *Drosophila* social communication. Nat Commun 8:. https://doi.org/10.1038/s41467-017-00334-9
- Mann RS, Ali JG, Hermann SL, et al (2012) Induced release of a plant-defense volatile "deceptively" attracts insect vectors to plants infected with a bacterial pathogen. PLoS Pathog 8:. https://doi.org/10.1371/journal.ppat.1002610
- Mauck KE, Chesnais Q, Shapiro LR (2018) Evolutionary Determinants of Host and Vector Manipulation by Plant Viruses, 1st edn. Elsevier Inc.
- Mauck KE, De Moraes CM, Mescher MC (2010) Deceptive chemical signals induced by a plant virus attract insect vectors to inferior hosts. Proc Natl Acad Sci U S A 107:3600–3605. https://doi.org/10.1073/pnas.0907191107
- Medeiros AH, Franco FP, Matos JL, et al (2012) Sugarwin: A sugarcane insect-induced gene with antipathogenic activity. Mol Plant-Microbe Interact 25:613–624. https://doi.org/10.1094/MPMI-09-11-0254
- Morales-Ramos JA, Guadalupe Rojas M, Sittertz-Bhatkar H, Saldana G (2000) Symbiotic relationship between Hypothenemus hampei (Coleoptera: Scolytidae) and Fusarium solani (Moniliales: Tuberculariaceae. Ann Entomol Soc Am 93:541–547. https://doi.org/10.1603/0013-8746(2000)093[0541:SRBHHC]2.0.CO;2
- Naranjo-Guevara N, Peñaflor MFGV, Silva DB, Bento JMS (2021) A comparison of the direct and indirect defence abilities of cultivated maize versus perennial and annual teosintes. Chemoecology 31:63–74. https://doi.org/10.1007/s00049-020-00329-x
- Ogunwolu EO, Reagan TE, Flynn JL, Hensley SD (1991) Effects of *Diatraea saccharalis* (F.) (Lepidoptera: Pyralidae) damage and stalk rot fungi on sugarcane yield in Louisiana. Crop Prot 10:57–61. https://doi.org/10.1016/0261-2194(91)90027-O
- Pare PW, Tumlinson JH (1999) Update on plant-insect interactions plant volatiles as a defense against insect herbivores by releasing greater amounts of a variety. Plant Physiol 121:325–331
- Peñaflor MFGV, Bento JMS (2019) Red-rot infection in sugarcane attenuates the attractiveness of sugarcane borerinduced plant volatiles to parasitoid. Arthropod Plant Interact 13:117–125. https://doi.org/10.1007/s11829-018-9629-6
- Ren L, Yingao M, Xie M, et al (2020) Rectal bacteria produce sex pheromones in the male oriental fruit fly. bioRxiv. https://doi.org/10.1101/2020.12.17.423356
- Schneider DI, Ehrman L, Engl T, et al (2019) Symbiont-driven male mating success in the Neotropical Drosophila paulistorum superspecies. Behav Genet 49:83–98. https://doi.org/10.1007/s10519-018-9937-8
- Silva DB, Jiménez A, Urbaneja A, et al (2021) Changes in plant responses induced by an arthropod influence the colonization behavior of a subsequent herbivore. Pest Manag Sci. https://doi.org/10.1002/ps.6454
- Thompson JN (1988) Evolutionary ecology of the relationship between oviposition preference and performance of offspring in phytophagous insects. Entomol Exp Appl 47:3–14. https://doi.org/10.1111/j.1570-

7458.1988.tb02275.x

- Turlings TCJ, Tumlinson JH, Lewis WJ (1990) Exploitation of herbivore-induced plant odors by host-seeking parasitic wasps. Science (80-) 250:1251–1253. https://doi.org/10.1126/science.250.4985.1251
- Visser J (1986) Host Odor Perception in Phytophagous Insects. Annu Rev Entomol 31:121–144. https://doi.org/10.1146/annurev.ento.31.1.121
- Zurek L, Wes Watson D, Krasnoff SB, Schal C (2002) Effect of the entomopathogenic fungus, Entomophthora muscae (Zygomycetes: Entomophthoraceae), on sex pheromone and other cuticular hydrocarbons of the house fly, Musca domestica. J Invertebr Pathol 80:171–176. https://doi.org/10.1016/S0022-2011(02)00109-X

2. PHYTOPATHOGENIC FUNGUS INDUCED CHANGES IN SUGARCANE MODULATES BEHAVIOR IN THE MOTH Diatraea saccharalis

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Fungal phytopathogen modulates plant and insect responses to promote its dissemination

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Abstract

Vector-borne plant pathogens often change host traits to manipulate vector behavior in a way that favors their spread. By contrast, infection by opportunistic fungi does not depend on vectors, although damage caused by an herbivore may facilitate infection. Manipulation of hosts and vectors, such as insect herbivores, has not been demonstrated in interactions with fungal pathogens. Herein, we establish a new paradigm for the plant-insect-fungus association in sugarcane. It has long been assumed that *Fusarium verticillioides* is an opportunistic fungus, where it takes advantage of the openings left by *Diatraea saccharalis* caterpillar attack to infect the plant. In this work, we show that volatile emissions from *F. verticillioides* attract *D. saccharalis* caterpillars. Once they become adults, the fungus is transmitted vertically to their offspring, which continues the cycle by inoculating the fungus into healthy plants. Females not carrying the fungus prefer to lay their eggs on fungus-infected plants. How may funguing the fungus impacts *D. saccharalis* sace plants than fungus ingete the fungus impacts *D. saccharalis* sex behavior, larval weight and reproduction rate, most individuals complete their development. Our data demonstrate that the fungus manipulates both the host plant and insect herbivore across life cycle to promote its infection and dissemination.

These authors contributed equally: Flávia P. Franco, Amanda C. Túler, Diego Z. Gallan

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Introduction

Plant-insect-fungus interactions involve plant defense responses that eventually influence insect behavior or pathogen infection [1-3]. Changes in plant metabolites and insect behavior can be explained by the "host manipulation hypothesis", which states that the pathogen can manipulate the insect vector and/or final host response to guarantee infection and dissemination [4-7]. Phytopathogens can change plant phenotypes, nutritional profiles, and the emission of volatile organic compounds (VOCs) [3], as a strategy to attract vectors and disseminate them [6]. An example is when the phytopathogen infection increases pre-existing host-location cues that are attractive to the vector, but also reduces host nutritional quality, making the vector move to a healthy plants [6]. Microorganisms can also alter insect behavior, for example, when they induce an increase in aggregation pheromone production resulting in the attraction of more individuals and, consequently, infecting more insects [8], when they affect insects' feeding behavior [9], perception of odors [10, 11] or tastes [12]. An extraordinary example of changes of insect behavior is "zombie-ants'

Abstract

Vector-borne plant pathogens often change host traits to manipulate vector behavior in a way that favors their spread. By contrast, infection by opportunistic fungi does not depend on vectors, although damage caused by an herbivore may facilitate infection. Manipulation of hosts and vectors, such as insect herbivores, has not been demonstrated in interactions with fungal pathogens. Herein we establish a new role for the insect-fungi-sugarcane association. It has long been assumed that *Fusarium verticillioides* and *Colletotrichum falcatum* are opportunistic fungi, where it takes advantage of the openings left by *Diatraea saccharalis* attack to infect the plant. In this work, we show that sugarcane infection attracts *D. saccharalis* adults to lay their eggs. However, only *F. verticillioides* seems to modulate the behavior of the *D. saccharalis* moth. More specifically, infected plants dramatically increased amounts of volatile organic compounds (VOCs) emission, attracting female moths for oviposition. Once they become adults, the fungus is transmitted vertically to their offspring, which continues the cycle by inoculating the fungus into healthy plants. Females not carrying the fungus prefer to lay their eggs on fungus-infected plants than mock plants, while females carrying the fungus prefer to lay their eggs on mock plants than fungus-infected plants. Most individuals complete their development even though the fungus impacts *D. saccharalis* sex behavior, larval weight, and reproduction rate. Our data demonstrate that the fungus manipulates both the host plant and insect herbivore across the life cycle to promote its infection and dissemination.

Keywords: Plant-fungi interaction, Fusarium verticillioides, Colletotrichum falcatum, vertical transmission, host manipulation hypothesis

2.1. Introduction

Plant-insect-fungus interactions involve plant defense responses that eventually influence insect behavior or pathogen infection (Ingwell et al. 2012; Franco et al. 2017; Sharifi and Ryu 2017). The "host manipulation hypothesis" can explain changes in plant defense pathways in primary and secondary metabolisms and insect behavior, which states that the pathogen can manipulate the insect vector and/or final host response to guarantee infection and dissemination (Fereres and Moreno 2009; Mauck et al. 2010, 2018; Ingwell et al. 2012; Heil 2016). Phytopathogens can change plant phenotypes, nutritional profiles, and the emission of volatile organic compounds (VOCs) (Franco et al. 2017) as a strategy to attract vectors and disseminate them (Mauck et al. 2010; Ingwell et al. 2012). An example is when the phytopathogen infection increases pre-existing host-location cues that are attractive to the vector, but also reduces host nutritional quality, making the vector move to a healthy plant (Mauck et al. 2010). Microorganisms can also alter insect behavior, for example, when they induce an increase in aggregation pheromone production resulting in the attraction of more individuals and, consequently, infecting more insects (Keesey et al. 2017) when they affect insects feeding behavior (Mauck et al. 2010; Stafford et al. 2011), perception of odors or tastes (Becher et al. 2010a; Stensmyr et al. 2012a). An extraordinary example of changes in insect behavior is ``zombie-ants`` which fungus manipulates the ants to behave like "zombies," to bite the underside of vegetation and spread the spores on nature (Hughes et al. 2011).

On the other hand, insects can participate in fungal dispersion by spreading spores over long distances (Phoku et al. 2016) and/or facilitating the entry of opportunistic fungi through feeding, as can occur in Ambrosia beetles (Zhao et al. 2019). As a vector, insects can transmit pathogens horizontally in which hosts contagiously acquire symbionts through an environmental source or, vertically, through parents inheritance to offspring; however, as there is a great diversity of organisms, this transmission also it can be mixed, that is, horizontally and vertically (Bright and Bulgheresi 2010).

Opportunistic pathogens presumably do not have an intimate association with the insect; moreover, the association between *Fusarium* spp. and borer-feeding insects have been reported, including the role of insects as fungal vectors, such as *Eldana saccharina* (Ako et al. 2003; McFarlane et al. 2009). In fact, "Bt crops" have been associated with lower mycotoxin levels because of the absence of moth larvae in the field, as *Ostrinia nubilalis*, the European corn borer (Bakan et al. 2002, Gatch and Munkvold 2002), *E. saccharina*, the African corn borer (Schulthess et al. 2002) and

Helicoverpa zea, the corn earworm (Gatch and Munkvold 2002; Bowers et al. 2013). Furthermore, the interaction between *F. veticillioides* and plants from the family Poaceae is a global problem for human and animal food due to the fumonisin production by the fungus of *Fusarium* spp. (Theumer et al. 2010).

The same fungi, F. *verticillioides*, usually occur in association with *Diatraea saccharalis* (F.) (Lepidoptera: Crambidae) and the fungus *Colletotrichum falcatum*, a prevalent pest in sugarcane fields (Ogunwolu et al. 1991). Hitherto, it was assumed that the fungus takes advantage of the openings made by the borer to penetrate the sugarcane stalk and infect the plant, resulting in additional crop damage due to pokkah boeng and red rot disease, respectively.

Nevertheless, recent research shows that *D. saccharalis* herbivory strongly induces genes encoding in the sugarcane plant to expresses defense proteins (Sugarwin1 and Sugarwin2). Interesting, these proteins affect specific *C. falcatum* and *F. verticillioides* morphology, causing fungal death, but are not able to affect *Aspergillus nidulans*, a non-sugarcane pathogenic fungus, even the *D. saccharalis*, the insect that induces the plant to produce these proteins (Medeiros et al. 2012, 2016; Franco et al. 2014). Thus, the sugarcane plants seem to protect the plant against opportunistic pathogens associating the *D. saccharalis* herbivory to *C. falcatum* and *F. verticillioides* pathogenicity. Then, the sugarcane-*D. saccharalis*-opportunistic fungi interaction appears to be more elaborate than the pre-established concept of the insect mediating fungal infection. Therefore, the sugarcane-*D. saccharalis*-opportunistic fungi were considered. Based on the previous association between *Fusarium* spp. and borer-feeding insects and the strong interaction between *F. verticillioides* and *D. saccharalis*, we investigated the role of *D. saccharalis* as a dissemination vector *F. verticillioides* possibly corroborating the "host manipulation hypothesis". We specifically addressed whether the fungus modifies *D. saccharalis* behavior in ways that increase the likelihood of the fungus being disseminated by the contaminated insect. Additionally, we evaluated whether this interaction benefits both organisms or only *F. verticillioides*.

2.2. Conclusion

Our results showed at the list for *F. verticillioides*, strong interaction with *D. saccharalis* in sugarcane-infected plants and support a paradigm change in which the fungus is not merely an opportunistic player manipulates both the host plant and insect throughout its life cycle to promote infection and dissemination. The *D. saccharalis* moths orient towards volatiles emitted by infected plants, which is the preferred host for oviposition. Interestingly, the host preference rather than plastic changes whether insects themselves are carriers of the fungus. Females not carrying *F. verticillioides* prefer to lay their eggs on fungus-infected plants, while females carrying the *F. verticillioides* prefer to lay their eggs on mock plants. In addition, *D. saccharalis* vertically transmits the fungus *F. verticillioides* to healthy plants by its offspring. In this scenario, our study indicates that *F. verticillioides* is not just an opportunistic fungus but also more intimately associated with an insect herbivore, manipulating the plant and the insect to increase its potential for dissemination.

- Ako M, Schulthess F, Gumedzoe MYD, Cardwell KF (2003) The effect of *Fusarium verticillioides* on oviposition behaviour and bionomics of lepidopteran and coleopteran pests attacking the stem and cobs of maize in West Africa. Entomol Exp Appl 106:201–210. https://doi.org/10.1046/j.1570-7458.2003.00026.x
- Bakan B, Melcion D, Richard-Molard D, Cahagnier B (2002) Fungal growth and *Fusarium* mycotoxin content in isogenic traditional maize and genetically modified maize grown in France and Spain. J Agric Food Chem 50:728–731. https://doi.org/10.1021/jf0108258

- Becher PG, Bengtsson M, Hansson BS (2010) Flying the fly: Long-range flight behavior of *Drosophila melanogaster* to attractive odors. 599–607. https://doi.org/10.1007/s10886-010-9794-2
- Bowers E, Hellmich R, Munkvold G (2013) Vip3Aa and Cry1Ab proteins in maize reduce *Fusarium* ear rot and fumonisins by deterring kernel injury from multiple Lepidopteran pests. World Mycotoxin J 6:127–135. https://doi.org/10.3920/WMJ2012.1510
- Bright M, Bulgheresi S (2010) A complex journey: Transmission of microbial symbionts. Nat Rev Microbiol 8:218–230. https://doi.org/10.1038/nrmicro2262
- Cardoza YJ, Teal PEA, Tumlinson JH (2003) Effect of peanut plant fungal infection on oviposition preference by *Spodoptera exigua* and on host-searching behavior by *Cotesia marginiventris*. Environ Entomol 32:970–976. https://doi.org/10.1603/0046-225X-32.5.970
- Fereres A, Moreno A (2009) Behavioural aspects influencing plant virus transmission by homopteran insects. Virus Res 141:158–168. https://doi.org/10.1016/j.virusres.2008.10.020
- Franco FP, Moura DS, Vivanco JM, Silva-Filho MC (2017) Plant–insect–pathogen interactions: a naturally complex ménage à trois. Curr Opin Microbiol 37:54–60. https://doi.org/10.1016/j.mib.2017.04.007
- Franco FP, Santiago AC, Henrique-Silva F, et al (2014) The sugarcane defense protein SUGARWIN2 causes cell death in *Collectotrichum falcatum* but not in non-pathogenic fungi. PLoS One 9:. https://doi.org/10.1371/journal.pone.0091159
- Franco FP, Túler AC, Gallan DZ, et al (2021) Fungal phytopathogen modulates plant and insect responses to promote its dissemination. ISME J. https://doi.org/10.1038/s41396-021-01010-z
- Gallan DZ (2019) Universidade de São Paulo Escola Superior de Agricultura "Luiz de Queiroz " Estudo da interação entre a broca da cana-de-açúcar Diatraea saccharalis (Lepidoptera: Crambidae) e fungos oportunistas Colletotrichum falcatum e Fusarium verticillioides Dieg. 81
- Gatch EW, Munkvold GP (2002) Fungal species composition in maize stalks in relation to European corn borer injury and transgenic insect protection. Plant Dis 86:1156–1162. https://doi.org/10.1094/PDIS.2002.86.10.1156
- Heil M (2016) Host manipulation by parasites: Cases, patterns, and remaining doubts. Front Ecol Evol 4:. https://doi.org/10.3389/fevo.2016.00080
- Hughes DP, Andersen SB, Hywel-Jones NL, et al (2011) Behavioral mechanisms and morphological symptoms of zombie ants dying from fungal infection. BMC Ecol 11:. https://doi.org/10.1186/1472-6785-11-13
- Ingwell LL, Eigenbrode SD, Bosque-Pérez NA (2012) Plant viruses alter insect behavior to enhance their spread. Sci Rep 2:. https://doi.org/10.1038/srep00578
- Jaenike J (1978) On optimal oviposition behavior in phytophagous insects. Theor Popul Biol 14:350-356. https://doi.org/10.1016/0040-5809(78)90012-6
- Keesey IW, Koerte S, Khallaf MA, et al (2017) Pathogenic bacteria enhance dispersal through alteration of *Drosophila* social communication. Nat Commun 8:. https://doi.org/10.1038/s41467-017-00334-9
- Mauck KE, Chesnais Q, Shapiro LR (2018) Evolutionary determinants of host and vector manipulation by plant viruses, 1st edn. Elsevier Inc.
- Mauck KE, De Moraes CM, Mescher MC (2010) Deceptive chemical signals induced by a plant virus attract insect vectors to inferior hosts. Proc Natl Acad Sci U S A 107:3600–3605. https://doi.org/10.1073/pnas.0907191107
- McFarlane SA, Govender P, Rutherford RS (2009) Interactions between *Fusarium* species from sugarcane and the stalk borer, *Eldana saccharina* (Lepidoptera: Pyralidae). Ann Appl Biol 155:349–359. https://doi.org/10.1111/j.1744-7348.2009.00345.x
- Medeiros AH, Franco FP, Matos JL, et al (2012) Sugarwin: A sugarcane insect-induced gene with antipathogenic activity. Mol Plant-Microbe Interact 25:613–624. https://doi.org/10.1094/MPMI-09-11-0254
- Medeiros AH, Mingossi FB, Dias RO, et al (2016) Sugarcane serine peptidase inhibitors, serine peptidases, and clp protease system subunits associated with sugarcane borer (*Diatraea saccharalis*) herbivory and wounding. Int J Mol Sci 17:. https://doi.org/10.3390/ijms17091444
- Meyer JS, Ingersoll CG, McDonald LL, Boyce MS (1986) Estimating uncertainty in population growth rates: jackknife vs. bootstrap techniques. Ecology 67:1156–1166. https://doi.org/10.2307/1938671
- Morales-Ramos JA, Guadalupe Rojas M, Sittertz-Bhatkar H, Saldana G (2000) Symbiotic relationship between Hypothenemus hampei (Coleoptera: Scolytidae) and Fusarium solani (Moniliales: Tuberculariaceae. Ann Entomol Soc Am 93:541–547. https://doi.org/10.1603/0013-8746(2000)093[0541:SRBHHC]2.0.CO;2
- Ogunwolu EO, Reagan TE, Flynn JL, Hensley SD (1991) Effects of *Diatraea saccharalis* (F.) (Lepidoptera: Pyralidae) damage and stalk rot fungi on sugarcane yield in Louisiana. Crop Prot 10:57–61. https://doi.org/10.1016/0261-2194(91)90027-O
- Peñaflor MFGV, Bento JMS (2019) Red-rot infection in sugarcane attenuates the attractiveness of sugarcane borerinduced plant volatiles to parasitoid. Arthropod Plant Interact 13:117–125. https://doi.org/10.1007/s11829-018-9629-6

- Phoku JZ, Barnard TG, Potgieter N, Dutton MF (2016) International Journal of Food Microbiology Fungal dissemination by house fly (*Musca domestica* L.) and contamination of food commodities in rural areas of South Africa. Int J Food Microbiol 217:177–181. https://doi.org/10.1016/j.ijfoodmicro.2015.10.028
- Schmelz EA, Engelberth J, Alborn HT, et al (2003) Simultaneous analysis of phytohormones, phytotoxins, and volatile organic compounds in plants. Proc Natl Acad Sci U S A 100:10552–10557. https://doi.org/10.1073/pnas.1633615100
- Schmidt-Büsser D, von Arx M, Guerin PM (2009) Host plant volatiles serve to increase the response of male European grape berry moths, *Eupoecilia ambiguella*, to their sex pheromone. J Comp Physiol A Neuroethol Sensory, Neural, Behav Physiol 195:853–864. https://doi.org/10.1007/s00359-009-0464-1
- Schulthess F, Cardwell KF, Gounou S (2002) The effect of endophytic *Fusarium verticillioides* on infestation of two maize varieties by lepidopterous stemborers and coleopteran grain feeders. Phytopathology 92:120–128. https://doi.org/10.1094/PHYTO.2002.92.2.120Seidenspinners ÜDS (2013) \ S. 283–284
- Stafford CA, Walker GP, Ullman DE (2011) Infection with a plant virus modifies vector feeding behavior. https://doi.org/10.1073/pnas.1100773108/-

/DCSupplemental.www.pnas.org/cgi/doi/10.1073/pnas.1100773108

- Stensmyr MC, Dweck HKM, Farhan A, et al (2012) A conserved dedicated olfactory circuit for detecting harmful microbes in *Drosophila*. Cell 151:1345–1357. https://doi.org/10.1016/j.cell.2012.09.046
- Theumer MG, Cánepa MC, López AG, et al (2010) Subchronic mycotoxicoses in Wistar rats: Assessment of the in vivo and in vitro genotoxicity induced by fumonisins and aflatoxin B1, and oxidative stress biomarkers status. Toxicology 268:104–110. https://doi.org/10.1016/j.tox.2009.12.007
- Thompson JN (1988) Evolutionary ecology of the relationship between oviposition preference and performance of offspring in phytophagous insects. Entomol Exp Appl 47:3–14. https://doi.org/10.1111/j.1570-7458.1988.tb02275.x
- Van Den Dool H, Kratz PD (1963) A generalization of the retention index system including linear temperature programmed gas-liquid partition chromatography. J Chromatogr A 11:463–471. https://doi.org/10.1016/S0021-9673(01)80947-X
- Von Arx M, Schmidt-Büsser D, Guerin PM (2011) Host plant volatiles induce oriented flight behaviour in male European grapevine moths, *Lobesia botrana*. J Insect Physiol 57:1323–1331. https://doi.org/10.1016/j.jinsphys.2011.06.010
- Zhao T, Ganji S, Schiebe C, et al (2019) Convergent evolution of semiochemicals across Kingdoms : bark beetles and their fungal symbionts. ISME J 1535–1545. https://doi.org/10.1038/s41396-019-0370-7

3. PHYTOPATHOGENIC FUNGUS PROMOTES CHANGES IN SEXUAL BEHAVIOR OF MOTH Diatraea saccharalis

Manuscript in preparation for submission

Abstract

A remarkably complex and fascinating network among *Diatraea saccharalis-Fusarium verticillioides*-sugarcane interaction elucidates the fungus manipulation to attract the moth *D. saccharalis* to lay eggs and feed on contaminated plants, promoting its infection and dissemination. The fact that these insects feed on these fungi without causing any consequences for their biology we argue whether the acquisition of the fungus *F. verticillioides* would alter the sexual behavior of *D. saccharalis*. Few examples of entomopathogenic microorganisms associated with insects resulting from pheromone modification and the attraction of more individuals than, consequently, infecting more insects or change the insect preference of mating. There is a paucity of information on microorganisms not associated with insects such as phytopathogens. Using chromatography and electroantennography tolls, we were able to evaluate the *F. verticillioides* effects in *D. saccharalis*.

Keywords Sugarcane-borer, Insect-microorganism interaction, Fusarium verticillioides, Pheromone

3.1. Introduction

Insect-microorganism interactions have been reported in several cases as oviposition stimulants and host location, and attraction to food resources (Becher et al. 2010; Stafford et al. 2011; Stensmyr et al. 2012; Soldano et al. 2016; Engl and Kaltenpoth 2018; Weisskopf et al. 2021). In addition, the most common microbial symbionts as *Wolbachia* and *Rickettsia* can also impact their host reproductive and sexual behavior to facilitate their spread since they can be sexually transmitted (Schneider et al. 2019; Flatau et al. 2021). Although, a shred of insufficient literature evidences the impacts of chemical communication between individuals by microorganisms, including symbiont and entomopathogenic microorganisms (Zurek et al. 2002; Keesey et al. 2017; Ren et al. 2020), there is a paucity of information on microorganisms not associated with insects such as phytopathogens.

A recent study showed that infection in sugarcane modulates behavior in the moth *Diatraea saccharalis*, guiding the moths to acquire *Fusarium verticillioides* and further enhance pathogen dispersal vertically transmitted to the offspring. Furthermore, even though the fungus no impacts the insect performance, it was demonstrated that the presence of the fungus influences *D. saccharalis* sexual behavior, decreasing the latency period and unsettle the copulation success when the adults were contaminated by the fungus (Franco et al. 2021). We argue whether the acquisition of the fungus *F. verticillioides* would alter the sexual behavior of *D. saccharalis*. Such discovery would be beneficial since *D. saccharalis* is a significant pest in Poaceous crops and a potential vector of *F. verticillioides*, the causal agent of pokkah boeng disease in sugarcane crop, and also, a global problem for human and animal food due to the fumonisin production by the fungi of *Fusarium* spp. (Theumer et al. 2010; Blacutt et al. 2018).

3.2. Conclusion

The advancement of a holistic view of insect-microbe interaction can increase insect pest and disease management efficacy. For instance, the knowledge of ecological interaction among plants, invertebrates, and fungi contributes to reduces pathogenic problems through insect pest control before transmitting the inoculum. Thus, such an approach would be ideal for controlling mycotoxins, a global human and animal health problem. However, the results were not conclusive.

- Becher PG, Bengtsson M, Hansson BS (2010) Flying the fly: Long-range flight behavior of *Drosophila melanogaster* to attractive odors. 599-607. https://doi.org/10.1007/s10886-010-9794-2
- Blacutt AA, Gold SE, Voss KA, et al (2018) Fusarium verticillioides: Advancements in understanding the toxicity, virulence, and niche adaptations of a model mycotoxigenic pathogen of maize. Phytopathology 108:312–326. https://doi.org/10.1094/PHYTO-06-17-0203-RVW
- Engl T, Kaltenpoth M (2018) Influence of microbial symbionts on insect pheromones. Nat Prod Rep 35:386–397. https://doi.org/10.1039/C7NP00068E
- Flatau R, Segoli M, Hawlena H (2021) Wolbachia endosymbionts of fleas occur in all females but rarely in males and do not show evidence of obligatory relationships, fitness effects, or sex-distorting manipulations. Front Microbiol 12:. https://doi.org/10.3389/fmicb.2021.649248
- Franco FP, Túler AC, Gallan DZ, et al (2021) Fungal phytopathogen modulates plant and insect responses to promote its dissemination. ISME J. https://doi.org/10.1038/s41396-021-01010-z
- Keesey IW, Koerte S, Khallaf MA, et al (2017) Pathogenic bacteria enhance dispersal through alteration of *Drosophila* social communication. Nat Commun 8:. https://doi.org/10.1038/s41467-017-00334-9
- Ren L, Yingao M, Xie M, et al (2020) Rectal bacteria produce sex pheromones in the male oriental fruit fly. bioRxiv. https://doi.org/10.1101/2020.12.17.423356
- Schneider DI, Ehrman L, Engl T, et al (2019) Symbiont-driven male mating success in the Neotropical Drosophila paulistorum superspecies. Behav Genet 49:83–98. https://doi.org/10.1007/s10519-018-9937-8
- Stafford CA, Walker GP, Ullman DE (2011) Infection with a plant virus modifies vector feeding behavior. https://doi.org/10.1073/pnas.1100773108/-
 - /DCSupplemental.www.pnas.org/cgi/doi/10.1073/pnas.1100773108
- Stensmyr MC, Dweck HKM, Farhan A, et al (2012) A conserved dedicated olfactory circuit for detecting harmful microbes in *Drosophila*. Cell 151:1345–1357. https://doi.org/10.1016/j.cell.2012.09.046
- Soldano A, Alpizar YA, Boonen B, Franco L, Lopez-Requena A, Liu GD, et al. Gustatory-mediated avoidance of bacterial lipopolysaccharides via TRPA1 activation in Drosophila. Elife. 2016;5. https://doi.org/10.7554/eLife.13133
- Theumer MG, Cánepa MC, López AG, et al (2010) Subchronic mycotoxicoses in Wistar rats: Assessment of the in vivo and in vitro genotoxicity induced by fumonisins and aflatoxin B1, and oxidative stress biomarkers status. Toxicology 268:104–110. https://doi.org/10.1016/j.tox.2009.12.007
- Zurek L, Wes Watson D, Krasnoff SB, Schal C (2002) Effect of the entomopathogenic fungus, *Entomophthora muscae* (Zygomycetes: Entomophthoraceae), on sex pheromone and other cuticular hydrocarbons of the house fly, *Musca domestica*. J Invertebr Pathol 80:171–176. https://doi.org/10.1016/S0022-2011(02)00109-X
- Weisskopf, L., Schulz, S. & Garbeva, P. (2021) Microbial volatile organic compounds in intra-kingdom and interkingdom interactions. Nat Rev Microbiol 19, 391–404. https://doi.org/10.1038/s41579-020-00508-1

4. CONCLUDING REMAKERS AND FUTURE PERSPECTIVES

The present thesis was conducted to obtain a comprehensive picture of the semiochemicals involved in sugarcane-*Diatraea saccharalis*-opportunistic fungi interaction, the fungi *Fusarium verticillioides*, and *Colletotrichum falcatum*. Briefly, the mains goals of the work were to address the chemical identification of plant defenses induced by both fungi infection, separated, or concomitant in the plant. Consequently, we evaluated the effects of plant defenses on insect behavior and fitness. Also, we evaluated the role of *D. saccharalis* as a vector of *F. verticillioides* infection in the sugarcane plant. Finally, we combined behavioral, electrophysiological, microscopy, and chromatography approach to testing our hypotheses specifically.

In fact, the sugarcane infection by the fungus separated or associated with the plant induces volatiles and nonvolatile chemical defenses. Even though *C. falcatum* has a neutral effect against *D. saccharalis*; the *F. verticillioides*-infected plants release VOCs, attract *D. saccharalis* adults to lay their eggs. The *F. verticillioides* is transmitted vertically to their offspring, which continues the cycle by inoculating the fungus into healthy plants. However, in contrast to other interactions, in which pathogen infection provides superior nutritional quality for the vector's offspring, *F. verticillioides* infection in sugarcane does not directly benefit the insect. Then, females carrying the fungus prefer to lay their eggs on mock plants than fungus-infected plants. Altogether, our results support a paradigm change in which the fungus is not merely an opportunistic player but also manipulates both the host plant and the insect throughout its life cycle to promote infection and dissemination, forming a mutualistic but inherently selfish association with sugarcane and *D. saccharalis*.

Even the sugarcane-*D. saccharalis*-opportunistic fungi interaction is the prevalent pest in sugarcane fields; a shred of insufficient literature evidences the chemical identification of plant volatiles in this interaction. Essentially, this disease control has been associated with managing the *D. saccharalis* larvae using the biological-control agent *Cotesia flavipes* on a large scale (Parra 2014). However, the absence of injury of the insect vector can increase insect pest and disease management efficacy; mainly allied to the fact that plants infected with *F. verticillioides* release less attractive herbivore-induced plant VOCs to *C. flavipes* (Peñaflor and Bento 2019), which can reduce the efficiency of parasitism in the field of co-occurring of *D. saccharalis* and *F. verticillioides*.

Because of the close sugarcane-*D. saccharalis*- *F. verticillioides* association and the specificity of *D. saccharalis* feeding on plants of the family Poaceae, the background of plant volatiles could influence pheromone perception in *D. saccharalis* previously reported to Noctuidae moths (Kromann et al. 2015; Badeke et al. 2016; Hoffmann et al. 2020). Also, microbe-induced plant volatiles (MIPVs) have been considered in developing new emerging techniques for sustainable plant disease management and confer immunity to neighboring susceptible cultivars (Lee 2017).

The co-occurrence of the pathogen, herbivore, and plants implicates complex interactions between them and is a complex task for many researchers; because this has been overlooked. However, this study changes our previous knowledge regarding sugarcane-*D. saccharalis*-opportunistic fungi interaction and enhances our understanding of plant-insect-fungus interactions, which can be extended to other systems.

In fact, the association between *Fusarium* spp. and borer-feeding insects have been reported, including the role of insects as fungal vectors, such as *Eldana saccharina*, the African corn borer (Ako et al. 2003; McFarlane et al. 2009), and *Ostrinia nubilalis*, the European corn borer (Gatch and Munkvold 2002; Bowers et al. 2013). Furthermore, "Bt crops" have been associated with lower mycotoxin levels because of deterring kernel injury from *Ostrinia nubilalis*, the European corn borer (Bakan et al. 2002, Gatch and Munkvold 2002), *E. saccharina*, (Schulthess et al. 2002), and *Helicoverpa zea*, the corn earworm (Gatch and Munkvold 2002; Bowers et al. 2013). The interactions among *Fusarium*

spp. and these Lepidopteran pests are a global problem for human and animal food due to the fumonisin, which exposes a toxic effect on farm animals and humans (Theumer et al. 2010; Drakulic et al. 2017; Blacutt et al. 2018).

On top of that, social problems associated with this interaction, the knowledge of ecological interaction among plants, herbivores, and fungi might contribute to reducing pathogenic problems through insect pest control before transmitting the inoculum. Thus, such an approach would be ideal for controlling mycotoxins, a global human and animal health problem.

- Ako M, Schulthess F, Gumedzoe MYD, Cardwell KF (2003) The effect of *Fusarium verticillioides* on oviposition behaviour and bionomics of lepidopteran and coleopteran pests attacking the stem and cobs of maize in West Africa. Entomol Exp Appl 106:201–210. https://doi.org/10.1046/j.1570-7458.2003.00026.x
- Badeke E, Haverkamp A, Hansson BS, Sachse S (2016) A challenge for a male noctuid moth? Discerning the female sex pheromone against the background of plant volatiles. Front Physiol 7:1–12. https://doi.org/10.3389/fphys.2016.00143
- Bakan B, Melcion D, Richard-Molard D, Cahagnier B (2002) Fungal growth and *Fusarium* mycotoxin content in isogenic traditional maize and genetically modified maize grown in France and Spain. J Agric Food Chem 50:728–731. https://doi.org/10.1021/jf0108258
- Blacutt AA, Gold SE, Voss KA, et al (2018) Fusarium verticillioides: Advancements in understanding the toxicity, virulence, and niche adaptations of a model mycotoxigenic pathogen of maize. Phytopathology 108:312–326. https://doi.org/10.1094/PHYTO-06-17-0203-RVW
- Bowers E, Hellmich R, Munkvold G (2013) Vip3Aa and Cry1Ab proteins in maize reduce *Fusarium* ear rot and fumonisins by deterring kernel injury from multiple Lepidopteran pests. World Mycotoxin J 6:127–135. https://doi.org/10.3920/WMJ2012.1510
- Drakulic J, Bruce TJA, Ray R V. (2017) Direct and host-mediated interactions between *Fusarium* pathogens and herbivorous arthropods in cereals. Plant Pathol 66:3–13. https://doi.org/10.1111/ppa.12546
- Gatch EW, Munkvold GP (2002) Fungal species composition in maize stalks in relation to European corn borer injury and transgenic insect protection. Plant Dis 86:1156–1162. https://doi.org/10.1094/PDIS.2002.86.10.1156
- Hoffmann A, Bourgeois T, Munoz A, et al (2020) A plant volatile alters the perception of sex pheromone blend ratios in a moth. J Comp Physiol A Neuroethol Sensory, Neural, Behav Physiol 206:553–570. https://doi.org/10.1007/s00359-020-01420-y
- Hosokawa T, Fukatsu T (2020) Relevance of microbial symbiosis to insect behavior. Curr Opin Insect Sci 39:91–100. https://doi.org/10.1016/j.cois.2020.03.004
- Kromann SH, Saveer AM, Binyameen M, et al (2015) Concurrent modulation of neuronal and behavioural olfactory responses to sex and host plant cues in a male moth. Proc R Soc B Biol Sci 282:. https://doi.org/10.1098/rspb.2014.1884
- Lee S (2017) Research review Microbe-induced plant volatiles. https://doi.org/10.1111/nph.14955
- McFarlane SA, Govender P, Rutherford RS (2009) Interactions between *Fusarium* species from sugarcane and the stalk borer, Eldana saccharina (Lepidoptera: Pyralidae). Ann Appl Biol 155:349–359. https://doi.org/10.1111/j.1744-7348.2009.00345.x
- Parra JRP (2014) Biological control in Brazil: An overview. Sci Agric 71:420–429. https://doi.org/10.1590/0103-9016-2014-0167
- Peñaflor MFGV, Bento JMS (2019) Red-rot infection in sugarcane attenuates the attractiveness of sugarcane borerinduced plant volatiles to parasitoid. Arthropod Plant Interact 13:117–125. https://doi.org/10.1007/s11829-018-9629-6
- Schulthess F, Cardwell KF, Gounou S (2002) The effect of endophytic *Fusarium verticillioides* on infestation of two maize varieties by lepidopterous stemborers and coleopteran grain feeders. Phytopathology 92:120–128. https://doi.org/10.1094/PHYTO.2002.92.2.120
- Theumer MG, Cánepa MC, López AG, et al (2010) Subchronic mycotoxicoses in Wistar rats: Assessment of the in vivo and in vitro genotoxicity induced by fumonisins and aflatoxin B1, and oxidative stress biomarkers status. Toxicology 268:104–110. https://doi.org/10.1016/j.tox.2009.12.007