University of São Paulo "Luiz de Queiroz" College of Agriculture

A comparative study of raspberry structural and biochemical responses to late leaf rust

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Thesis presented to obtain the degree of Doctor in Science. Area: Plant Physiology and Biochemistry

Piracicaba 2023 Márcia Gonçalves Dias BSc in Biology

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RESUMO

Estudo comparativo das respostas estruturais e bioquímicas de framboeseiras a ferrugem tardia da folha

Rosaceae é uma família composta por culturas importantes como maçã, pêssego, morango e framboesa. Entre essas espécies, as framboesas vermelhas e pretas pertencem ao gênero Rubus e são culturas de alto valor com crescente demanda mundial. No entanto, essas plantas são acometidas por diversas doenças que impactam sua qualidade e produtividade. A ferrugem tardia da folha é uma doenca fúngica em framboesas causada por Aculeastrum americanum (Farl.) M. Scholler & U. Braun (syn. Thekopsora americana (Farl.) Aime & McTaggart). Enquanto as framboesas vermelhas (Rubus idaeus L.) são suscetíveis, as framboesas pretas foram relatadas anteriormente como mais resistentes (R. occidentalis L.) e imunes (R. niveus Thunb.) a esse patógeno. Uma vez que a resistência genética é uma forma promissora de manejar esta doença, a hibridização de framboesas vermelhas e pretas pode fornecer características interessantes a novas cultivares. No entanto, é importante entender como as plantas respondem aos patógenos antes de desenvolver caros e longos programas de melhoramento. O objetivo desta tese foi investigar as respostas histopatológicas e bioquímicas de framboesas vermelhas e pretas e um híbrido entre elas à colonização por A. americanum. Observou-se que o patógeno germinou e colonizou todas as framboesas estudadas, entretanto, as framboesas pretas e o híbrido tiveram respostas distintas em comparação a R. idaeus. O mesofilo compacto, os compostos fenólicos pré e pós-formados, os compostos pécticos pós-formados e o colapso celular na área lesionada foram os principais mecanismos de defesa contra A. americanum. Adicionalmente aos estudos estruturais e bioquímicos das respostas de defesa da framboesa, foi realizada uma análise genômica comparativa para identificar as proteínas quinases dependentes de cálcio (CDPKs), com foco nos ortólogos CPK28 em framboesa preta (R. occidentalis) e morango (Fragaria vesca L.) como representante das Rosaceae. Ambos têm 26 CDPKs juntos. Os ortólogos FvCPK28 e RoCPK28 foram clonados e expressos em mutantes Nicotiana benthamiana e Arabidopsis cpk28. Foi encontrado que ambos os ortólogos CPK28 de Rosaceae são localizados na membrana plasmática e sua superexpressão amorteceu a explosão oxidativa após a elicitação. Esses resultados forneceram uma prova de conceito para investigar as semelhanças funcionais entre essas proteínas e Arabidopsis CPK28.

Palavras-chave: CPK28, Mecanismos de defesa, Compostos fenólicos, Rubus idaeus, Rubus occidentalis, Rubus niveus

ABSTRACT

A comparative study of raspberry structural and biochemical responses to late leaf rust

Rosaceae is a family composed of important crops such as apple, peach, strawberry, and raspberry. Among these species, red and black raspberries belong to the genus Rubus and are high-value crops with increasing worldwide demand. However, these plants are affected by several diseases that impact their quality and productivity. Late leaf rust is a fungal disease in raspberries caused by *Aculeastrum americanum* (Farl.) M. Scholler & U. Braun (syn. Thekopsora americana (Farl.) Aime & McTaggart). While red raspberries (*Rubus idaeus* L.) are susceptible, black raspberries were previously reported as more resistant (R. occidentalis L.) and immune (R. niveus Thunb.) to this pathogen. Since genetic resistance is a promising way to manage this disease, hybridizing red and black raspberries can provide new cultivars with interesting traits. However, it is important to understand how plants respond to pathogens before developing expensive and long breeding programs. The aim of this thesis was to investigate the histopathological and biochemical responses of red and black raspberries and a hybrid between them to A. americanum colonization. It was observed that the pathogen germinated and colonized all studied raspberries, however, black raspberries and the hybrid had distinctive responses compared to R. idaeus. The compact mesophyll, the pre- and post-formed phenolic compounds, post-formed pectic compounds, and cell collapse in the lesioned area were the main defense mechanisms against A. americanum. Additionally, to structural and biochemical studies of raspberries defense responses, it was performed a comparative genomics analysis to identify the calcium-dependent protein kinases (CDPKs), focusing on the CPK28 orthologs in black raspberry (R. occidentalis) and strawberry (Fragaria vesca L.) as representative of Rosaceae. Both have 26 CDPKs together. The orthologs FvCPK28 and RoCPK28 were cloned and expressed in *Nicotiana benthamiana* and *Arabidopsis* cpk28 mutants. It was found that both Rosaceae CPK28 orthologs are plasmamembrane localized, and their overexpression dampened the oxidative burst upon elicitation. These results provided a proof-of-concept to investigate the functional similarities between these proteins and Arabidopsis CPK28.

Keywords: CPK28, Defense mechanisms, Phenolic compounds, *Rubus idaeus*, *Rubus occidentalis*, *Rubus niveus*

1. GENERAL INTRODUCTION

Raspberries belong to the genus *Rubus* L. and occur naturally in all continents except Antarctica (Martin et al. 2018; Funt and Hall 2013). They are included within the family Rosaceae, along with blackberries, apples, pears, strawberries and several other species (Martin et al. 2018; Jung et al. 2019). Raspberries have been cultivated for centuries because of their nutritious fruit (druplets), as well as their herbal raw material rich in bioactive compounds (Oszmiański et al. 2015; Chwil and Kostryco 2021). Indeed, the global production of red and black raspberries has increased during the last couple of years to reach 822,000 tonnes in 2021 (Foster et al. 2019; Klewicka et al. 2020; FAO 2021). For this reason, local and global agricultural economies are highly dependent on maintaining production quality.

Rubus can be affected by a wide variety of diseases caused by viruses, fungi, and bacteria. More than 30 viruses and phytoplasmas affecting *Rubus* have been characterized in the last few years, and many others have not yet been identified (Funt 2013; Martin et al. 2018). Fungal diseases such as rusts are common in cane, fruits and leaves, and have the potential to cause severe yield loss (Hall et al. 2009; Funt 2013; Dolan et al. 2018).

Late leaf rust is a disease caused by *Aculeastrum americanum* (Farl.) M. Scholler & U. Braun (syn. *Thekopsora americana* (Farl.) Aime & McTaggart), an heteroecious and macrocyclic rust. Besides producing telia and uredinia in raspberries, white spruce (*Picea glauca* [Moench.] Voss) hosts the spermogonial and aecial stages of the *A. americanum* life cycle (Martin et al. 2018; Scholler et al. 2022). Although late leaf rust was once considered a minor concern, outbreaks have been reported in many locations of raspberry cultivation (Martin et al. 2018; Delisle-Houde et al. 2020; Oliveira 2021). The urediniospore stage is the only rust spore that causes reinfection, which may occur within a few days after sporulation and reach the same tissue or the neighbour plants (Aime et al. 2018; Duplessis et al. 2021). Due to this, they are persistent while finding optimal environmental conditions and are harder to control. Not surprisingly, late leaf rust has been found in raspberries in regions far from the white spruce occurrence, apparently overwintering on raspberries' aerial tissues (Martin et al. 2018).

Red raspberries (*Rubus idaeus* L.) are susceptible to late leaf rust, while black raspberries are considered immune (Luffman and Buszard 1989; Nelson 2011; Martin et al. 2018). It has been demonstrated that susceptibility varies among cultivars of *R*.

idaeus (Luffman and Buszard 1989; Hall et al. 2009). Even the black raspberry *R. niveus* is considered immune to *A. americanum* further studies are necessary to confirm this information. Moreover, based on the literature, the black raspberry *R. occidentalis* L. appears to have a contradictory host status. Even classified as black raspberry by Bushakra et al. (2012) and Foster et al. (2019) and immune to *A. americanum* by Dodge (1923), the occurrence of late leaf rust was previously observed (Martin et al. 2018).

Histopathological studies have helped clarify several mechanisms plants use during interaction with microbes (Braga et al. 2019; Navarro et al. 2019; Rasera et al. 2019; Rincón-Barón et al. 2020; Marques et al. 2022). In addition, the plant basal defense activates a cascade of responses performed by robust cellular machinery in response to the recognition of 'non-self' molecules (DeFalco and Zipfel 2021; Dias et al. 2022).

In order to control diseases, it is important to understand how plants respond to pathogens. In this context, the objective of this thesis was to uncover the structural and biochemical pre- and post-formed defense mechanisms of raspberries interacting with *A. americanum* (Chapters 2 and 3). In addition, calcium-dependent protein kinase CPK28 orthologs were identified and examined for their function in black raspberry and strawberry immunity (Chapter 4).

Chapter 2: Investigating biochemical and histopathological responses between raspberries and *Aculeastrum americanum*

Chapter 3: A new hybrid between red and black raspberry and its response to late leaf rust from a histopathological view

Chapter 4: Initial characterization of the calcium-dependent protein kinase CPK28 in black raspberry and strawberry

- Aime MC, Bell CD, Wilson AW (2018) Deconstructing the evolutionary complexity of rust fungi (*Pucciniales*) and their plant hosts. **Studies in Mycology** 89:143-52.
- Braga ZV, Santos RF, Amorim L, Appezzato-da-Glória B (2019) Histopathology of infection and colonisation of *Elsinoë ampelina* on grapevine leaves. **European Journal of Plant Pathology** 154(4): 1009-1019.

- Bushakra JM, Stephens MJ, Atmadjaja AN, Lewers KS, Symonds VV, Udall JA, Chagné D, Buck EJ, Gardiner SE (2012) Construction of black (*Rubus occidentalis*) and red (*R. idaeus*) raspberry linkage maps and their comparison to the genomes of strawberry, apple, and peach. **Theoretical and Applied Genetics** 125(2):311-327.
- Chwil M, Kostryco M (2020) Histochemical assays of secretory trichomes and the structure and content of mineral nutrients in *Rubus idaeus* L. leaves. **Protoplasma** 257: 119-139.
- Dias MG, Soleimani F, Monaghan J (2022) Activation and turnover of the plant immune signaling kinase BIK1: a fine balance. **Essays in Biochemistry** 66(2):207-218.
- DeFalco TA, Zipfel C (2021) Molecular mechanisms of early plant pattern-triggered immune signaling. **Molecular Cell** 81(17):3449-67.
- Delisle-Houde M, Demers F, Tweddell R (2020) Evaluation of phytosanitary products for the management of raspberry late leaf rust [*Pucciniastrum americanum* (Farl.) Arthur]. Phytoprotection 100(1):16-21.
- Dodge O (1923) Morphology and host reactions of *Pucciniastrum americanum*. Journal Agricultural Research 24: 885-894.
- Dolan A, MacFarlane S, Jennings SN (2018) Pathogens in raspberry and other *Rubus* spp. In: Graham J, Brennan R. **Raspberry: breeding, challenges and advances**. Springer 41-61.
- Duplessis S, Lorrain C, Petre B, Figueroa M, Dodds PN, Aime MC (2021) Host adaptation and virulence in heteroecious rust fungi. **Annual Review of Phytopathology** 59:403-422.
- FAO (2021) FAOSTAT: agricultural data. http://www.fao.org/faostat/en/#data/QC [accessed 2022 Mar 28].
- Foster TM, Bassil NV, Dossett M, Worthington ML, Graham J (2019) Genetic and genomic resources for *Rubus* breeding: a roadmap for the future. **Horticulture Research** 6:116.
- Funt RC (2013) Pest and Disease Management, p. 133-155. In: Funt RC, Hall HK. ed. Raspberries. Crop Production Science in Horticulture series 23. CAB International 133-155.
- Funt RC, Hall HK (2013) **Raspberries**. Crop Production Science in Horticulture series 23. CAB International.
- Hall HK, Hummer K, Jamieson AJ, Jennings SN, Weber CA (2009) Raspberry breeding and genetics. Pages 39–353 in Janick J, ed. **Plant Breeding Reviews**. Wiley.
- Jung S, Lee T, Cheng CH, Buble K, Zheng P, Yu J, Humann J, Ficklin SP, Gasic K, Scott K, Frank M (2019) 15 years of GDR: New data and functionality in the Genome Database for Rosaceae. Nucleic Acids Research. 47(D1): D1137-1145.
- Klewicka E, Sójka M, Ścieszka S, Klewicki R, Milczarek A, Lipińska L, Kołodziejczyk K (2020) The antimycotic effect of ellagitannins from raspberry (*Rubus idaeus* L.) on *Alternaria alternata* ŁOCK 0409. **European Food Research and Technology** 246(7): 1341-1349.
- Luffman M, Buszard D (1989) A note on the susceptibility of six red raspberry cultivars and Tayberry to fruit infection by late yellow rust. **Phytoprotection** 71:93-95.

- Marques JPR, Cia MC, Granato, ABA, Muniz LF, Appezzato-da-Glória B, Camargo LEA (2022) Histopathology of the shoot apex of sugarcane colonized by *Leifsonia xyli* subsp. *xyli*. **Phytopathology** 112(10): 2062-2071.
- Martin RR, Ellis MA, Williamson B, Williams RN (2018) PART I: Diseases Caused by Biotic Factors. In: Martin et al. (Ed.). **Compendium of Raspberry and Blackberry Diseases and Pests**, Second Edition, APS Press.
- Navarro BL, Marques JP, Appezzato-da-Glória B, Spósito MB (2019) Histopathology of *Phakopsora euvitis* on *Vitis vinifera*. **European Journal of Plant Pathology** 154(4):1185-1893.
- Nelson S (2011) Raspberry late leaf rust in Hawaii caused by *Pucciniastrum americanum*. **Plant Disease** 5.
- Oszmiański J, Wojdyło A, Nowicka P, Teleszko M, Cebulak T, Wolanin M (2015) Determination of phenolic compounds and antioxidant activity in leaves from wild *Rubus* L. species. **Molecules** 20(3):4951-4966.
- Oliveira PB (2021) Manual de Boas Práticas de Fruticultura Framboesa. **Revista Frutas**, **Legumes e Flores** (INIAV) 215(7).
- Rasera JB, Amorim L, Marques JPR, Soares MK, Appezzato-da-Glória B (2019) Histopathological evidences of early grapevine leaf senescence caused by *Phakopsora euvitis* colonisation. **Physiological and Molecular Plant Pathology** 108:101434.
- Rincón-Barón EJ, Gutiérrez Rodríguez AM, Guerra BE, Espinosa Matías S (2020) Alteraciones histopatológicas causadas por la roya *Puccinia nakanishikii* (Pucciniales: Pucciniaceae) en plantas de *Cymbopogon citratus* (Poaceae). **Revista de Biología Tropical** 68(2):361-382.
- Scholler M, Braun U, Buchheit R, Schulte T, Bubner B (2022) Studies on European rust fungi, *Pucciniales*: molecular phylogeny, taxonomy, and nomenclature of miscellaneous genera and species in Pucciniastraceae and Coleosporiaceae. **Mycological Progress** 21(8):1-25.

2. INVESTIGATING BIOCHEMICAL AND HISTOPATHOLOGICAL RESPONSES BETWEEN RASPBERRIES AND ACULEASTRUM AMERICANUM

Abstract

Late leaf rust is a fungal disease in raspberries caused by Aculeastrum americanum (Farl.) M. Scholler & U. Braun (syn. Thekopsora americana (Farl.) Aime & McTaggart) leading to early defoliation and yield losses. Red raspberries (Rubus *idaeus* L.) are susceptible to this pathogen, even though this susceptibility varies among cultivars. In contrast, black raspberries were previously reported as more resistant (Rubus occidentalis L.) and immune (Rubus niveus Thunb.) to this pathogen, raising their importance in plant breeding programs. However, what features make them respond differently to the same pathogen? In this study, we characterized preand post-formed structural and biochemical defense mechanisms of R. idaeus 'Autumn Bliss', R. occidentalis and R. niveus. Ultrastructural and histopathological analyses were employed to uncover the interactions between these raspberries and A. americanum. The ultrastructural results indicated that the pathogen germinates on both leaf surfaces but can form appressoria only on stomata. Although the three raspberry species were infected and colonized by A. americanum, a clear difference in susceptibility was observed between them. A compact mesophyll, pre and post formed phenolic compounds, and post formed pectic compounds were the main plant defense mechanisms against fungal colonization. These findings provide new information about raspberries' defense mechanisms in response to A. americanum and elucidate the interactions occurring on these pathosystems.

Keywords: late leaf rust, *Pucciniastrum americanum*, *Rubus idaeus*, *Rubus occidentalis*, *Rubus niveus*.

Introduction

Late leaf rust, caused by *Aculeastrum americanum* (Farl.) M. Scholler & U. Braun (syn. *Thekopsora americana* (Farl.) Aime & McTaggart) affects red and purple raspberries (Martin et al. 2018; Scholler et al. 2022). The disease is hard to control and has caught attention after outbreaks in North American orchards (Martin et al. 2018; Delisle-Houde et al. 2020). It has also been reported as a concern in Argentina, Brazil, Chile, and Portugal (Figueiredo et al. 2003; Raseira et al. 2004; Lucero et al. 2008; Pio 2014; Oliveira 2021).

The main symptoms are powdery yellow spots, which correspond to reproductive structures called uredinia and are found in all aerial parts of infected plants (Dolan et al. 2018; Martin et al. 2018). Infected fruits become unfit for sale, and leaves may drop prematurely, causing severe yield loss (Hall et al. 2009; Funt 2013; Martin et al. 2018). Those are some outcomes of plant tissue colonization that lead to

structural and physiological responses during the plant-pathogen interaction. However, it is still unknown which defense mechanisms raspberries employ against the colonization process by *A. americanum*.

Red raspberries (*Rubus idaeus* L.) are classified as susceptible to late leaf rust, even though this susceptibility varies among cultivars (Luffman and Buszard 1990; Nelson 2011). In contrast, black raspberries such as *Rubus occidentalis* L. and *Rubus niveus* Thunb. are immune to this pathogen (Martin et al. 2018). Nevertheless, late leaf rust has been documented in *R. occidentalis* (Dodge 1923; Darker 1929; Nickerson 1991).

Because raspberries are economically relevant crops on almost all continents (Foster et al. 2019; FAO 2021) diseases affecting these plants can not be neglected. Histopathological and biochemical studies have allowed the understanding of important pathosystems (Braga et al. 2019; 2021; Primiano et al. 2019; Rasera et al. 2019; Alves et al. 2021, Marques et al. 2022) and shed light on disease cycle and epidemiology studies (Nogueira Júnior et al. 2017; Boufleur et al. 2022; Dias et al. 2022).

Since no histopathological investigations have been performed yet on the infection and colonization processes of raspberries by *A. americanum*, this work seeks to elucidate whether there are differences in leaf anatomical and biochemical traits among red and black raspberries which may hinder/delay the infection of *A. americanum* in black raspberries leaves. This study focuses on uncovering pre- and post-formed defense mechanisms in red and black raspberries.

Conclusion

In conclusion, a more compact mesophyll, pre and post formed phenolic compounds, and post formed pectic compounds are the main defense mechanisms found in raspberries that played a role against *A. americanum*. Although raspberries had both preformed and post formed defense mechanisms, they were not sufficient to totally contain an infection and colonization by A. *americanum*. Based on the results, we confirmed the susceptibility of *R. idaeus* 'Autumn Bliss' to *A. americanum* and showed the absence of immunity for *R. occidentalis* and *R. niveus*.

- Adendorff R, Rijkenberg FH (2000) Scanning electron microscopy of direct host leaf penetration by urediospore-derived infection structures of *Phakopsora apoda*. Mycological Research 104(3):317-324.
- Ainsworth EA, Gillespie KM (2007) Estimation of total phenolic content and other oxidation substrates in plant tissues using Folin–Ciocalteu reagent. **Nature Protocols** 2(4):875-877.
- Alves RF, Marques JPR, Appezzato-da-Glória B, Spósito MB (2021) Process of infection and colonization of *Pseudocercospora kaki* in persimmon leaves. Journal of Phytopathology 169(3):168-175.
- Avelino J, Willocquet L, Savary S (2004) Effects of crop management patterns on coffee rust epidemics. **Plant Pathology** 53(5):541-547.
- Babu AM, Phillip T, Kariappa BK, Kamble CK (2009) Scanning electron microscopy of the infection process of *Cercospora henningsii* on cassava leaves. **Journal of Phytopathology** 157:57-62.
- Bettgenhaeuser J, Gilbert B, Ayliffe M, and Moscou MJ (2014) Nonhost resistance to rust pathogens a continuation of continua. **Frontiers in Plant Science** 5:664.
- Boufleur T, Morales JVP, Martins TV, Gonçalves MP, Massola NS, Amorim L (2022) A diagnostic guide for myrtle rust. **Plant Health Progress**.
- Braga ZV, Santos RF, Amorim L, Appezzato-da-Glória B (2019) Histopathology of infection and colonisation of *Elsinoë ampelina* on grapevine leaves. **European Journal of Plant Pathology** 154(4): 1009-1019.
- Braga ZV, Muniz LF, Manarim GR, de Aguiar CL, Appezzato-da-Glória B (2021) Anatomical and biochemical changes in leaves of *Vitis labrusca* L. cv. Niagara Rosada in response to infection by *Elsinoë ampelina* Shear. **Brazilian Journal of Botany** 44(1):187-196.
- Broadhurst RB, Jones WT (1978) Analysis of condensed tannins using acidified vanillin. Journal of the Science of Food and Agriculture 29(9):788-794.
- Cheng Y, Zhang H, Yao J, Wang X, Xu J, Han Q, Wei G, Huang L, Kang Z (2012) Characterization of non-host resistance in broad bean to the wheat stripe rust pathogen. **BMC Plant Biology** 12(1):1-2.
- Chwil M, Kostryco M (2020) Histochemical assays of secretory trichomes and the structure and content of mineral nutrients in *Rubus idaeus* L. leaves. **Protoplasma** 257: 119-139.
- Costea T, Vlase L, Gostin IN, Olah NK, Predan GM (2016) Botanical characterization, phytochemical and antioxidant activity of indigenous red raspberry (*Rubus idaeus* L.) leaves. **Studia Universitatis Vasile Goldis. Seria Stiintele Vietii** 26:463-472.
- Darker G D (1929) Cultures of *Pucciniastrum Americanum* (Farlow) Arthur and *P. Arcticum* (Lagerheim) Tranzschel. Journal of the Arnold Arboretum 10(3):156-167.
- Delisle-Houde M, Demers F, Tweddell R (2020) Evaluation of phytosanitary products for the management of raspberry late leaf rust [*Pucciniastrum americanum* (Farl.) Arthur]. Phytoprotection 100(1):16-21.

- Dias MG, Ribeiro RR, Barbosa CMA, Jesus JMI, Sposito MB (2022) Diagrammatic scale for improved late leaf rust severity assessments in raspberry leaves. **Canadian Journal of Plant Pathology**.
- Dodge O (1923) Morphology and host reactions of *Pucciniastrum americanum*. Journal Agricultural Research 24: 885-894.
- Dolan A, MacFarlane S, Jennings SN (2018) Pathogens in raspberry and other *Rubus* spp. In: Graham J, Brennan R. **Raspberry: breeding, challenges and advances.** Springer 41-61.
- Duplessis S, Lorrain C, Petre B, Figueroa M, Dodds PN, Aime MC (2021) Host adaptation and virulence in heteroecious rust fungi. **Annual Review of Phytopathology** 59:403-422.
- FAO (2021) FAOSTAT: agricultural data. http://www.fao.org/faostat/en/#data/QC [accessed 2022 Mar 28].
- Fell KR, Rowson JM (1956) Anatomical studies in the genus *Rubus*: Part I. The Anatomy of the Leaf of *Rubus idaeus* L. Journal of Pharmacy and Pharmacology 8(1): 334-345.
- Fell KR, Rowson JM (1960) Anatomical studies in the genus *Rubus*: Part III. The Anatomy of the leaf of *Rubus loganobaccus* L.H. Bailey. Journal of Pharmacy and Pharmacology 12(1):473-487.
- Fell KR, Rowson JM (1961) Anatomical studies in the genus *Rubus*: Part IV. Anatomical variations in the leaves of cultivated varieties of *R. idaeus* L. and *R. loganobaccus* L.H. Bailey, and of certain species of Bramble. Journal of Pharmacy and Pharmacology 13(1): 83-92.
- Figueiredo MB, Nogueira EMC, Ferrari JT, Aparecido CC, Hennen JF (2003) Ocorrência de ferrugem em framboesa no Estado de São Paulo. Arquivo do Instituto Biológico, São Paulo 70:199-201.
- Foster TM, Bassil NV, Dossett M, Worthington ML, Graham J (2019) Genetic and genomic resources for *Rubus* breeding: a roadmap for the future. **Horticulture Research** 6:116.
- Funt, RC (2013) Pest and Disease Management, p. 133-155. In: In: Funt RC, Hall HK. ed. Raspberries. Crop Production Science in Horticulture series 23. CAB International 133-155.
- George BP, Parimelazhagan T, Chandran R, Saravanan S (2014) A comparative study on invitro and in-vivo antioxidant properties of *Rubus ellipticus* and *Rubus niveus*. **Pharmacology** (7):247-255.
- Goellner K, Loehrer M, Langenbach C, Conrath UW, Koch E, Schaffrath U (2010) *Phakopsora pachyrhizi*, the causal agent of Asian soybean rust. **Molecular Plant Pathology** 11(2):169-177.
- Hall HK, Hummer K, Jamieson AJ, Jennings SN, Weber CA (2009) Raspberry breeding and genetics. Pages 39-353 in Janick J, ed. **Plant Breeding Reviews**. Wiley.
- Hiscox JD, Israelstam GF (1979) A method for the extraction of chlorophyll from leaf tissue without maceration. **Canadian Journal of Botany** 57(12):1332-1334.
- Hoefle C, Loehrer M, Schaffrath U, Frank M, Schultheiss H, Hückelhoven R (2009) Transgenic suppression of cell death limits penetration success of the soybean rust fungus *Phakopsora pachyrhizi* into epidermal cells of barley. **Phytopathology** 99(3):220-226.

- Horridge GA, Tamm SL (1969) Critical point drying for scanning electron microscopic study of ciliary motion. **Science** 163(3869):817-818.
- Hunt P (1968) Cuticular penetration by germinating uredospores. **Transactions of the British Mycological Society** 51: 103-112.

Jensen WA (1962) Botanical histochemistry: principles and practice. Freeman, San Francisco.

Johansen DA (1940) Plant microtechnique. McGraw-Hill, New York.

- Karnovsky JM (1965) A formaldehyde-glutaraldehyde fixative of high osmolality for use in electron microscopy. **Journal of Cell Biology** 27:137-138.
- Karley AJ, Mitchell C, Brookes C, McNicol J, O'Neill T, Roberts H, Graham J, Johnson SN (2016) Exploiting physical defence traits for crop protection: Leaf trichomes of *Rubus idaeus* have deterrent effects on spider mites but not aphids. **Annals of Applied Biology** 168(2): 159-172.
- Kolmer JA, Ordonez ME, Groth JV (2009) The rust fungi. In: **Encyclopedia of Life Sciences** (ELS), pp. 1-8. Chichester, UK: Wiley & Sons.
- Luque R, Sousa HC, Kraus JE (1996) Métodos de coloração de Roeser (1972): modificado-e Kropp (1972) visando a substituição do azul de astra por azul de alcião 8GS ou 8GX. Acta Botanica Brasilica 10:199-212.
- Lygin AV, Li S, Vittal R, Widholm JM, Hartman GL, Lozovaya VV (2009) The importance of phenolic metabolism to limit the growth of *Phakopsora pachyrhizi*. **Phytopathology** 99(12):1412-1420.
- Li M, Li W, Sun Y, Mao P, Qi X, Wang Y (2018) Analysis of leaf tissue structures between rustresistant and rust-susceptible *Zoysia grass* (*Zoysia japonica*). Acta Physiologiae Plantarum 40(4):1-9.
- Lu Y, Chen Q, Bu Y, Luo R, Hao S, Zhang J, Tian J and Yao Y (2017) Flavonoid accumulation plays an important role in the rust resistance of malus plant leaves. **Frontiers in Plant Science** 8:1286.
- Lucero X, Wright ER, Pérez BA (2008) Occurrence of late leaf rust caused by *Pucciniastrum americanum* in red raspberry (*Rubus idaeus*) in Buenos Aires, Córdoba, and Entre Ríos, Argentina. **Plant Disease** 92:653.
- Luffman M, Buszard D (1989) A note on the susceptibility of six red raspberry cultivars and Tayberry to fruit infection by late yellow rust. **Phytoprotection** 71:93-95.
- Mabry TJ, Markham KR, Thomas MB (1970) Reagents and procedures for the ultraviolet spectral analysis of flavonoids. In: Mabry TJ, Markham KR, Thomas MB (eds). **The systematic identification of flavonoids.** Springer, Berlin.
- Marques JP, Hoy JW, Appezzato-da-Glória B, Viveros AF, Vieira ML, Baisakh N (2018) Sugarcane cell wall-associated defense responses to infection by *Sporisorium scitamineum*. Frontiers in Plant Science 9:698.

- Marques JPR, Cia MC, Granato, ABA, Muniz LF, Appezzato-da-Glória B, Camargo LEA (2022) Histopathology of the shoot apex of sugarcane colonized by *Leifsonia xyli* subsp. *xyli*. **Phytopathology** 112(10): 2062-2071.
- Martin RR, Ellis MA, Williamson B, Williams RN (2018) PART I: Diseases Caused by Biotic Factors. In: Martin et al. (Ed.). **Compendium of Raspberry and Blackberry Diseases and Pests**, Second Edition, APS Press.
- Mendgen K, Deising H (1993) Infection structures of fungal plant pathogens–a cytological and physiological evaluation. **New Phytologist** 124(2):193-213.
- Minchio CA, Fantin LH, de Oliveira KB, Rocha JA, Canteri MG (2017) Morphological changes of the urediniospore of *Puccinia kuehnii* germ tube in function of temperature. **Agronomy Science and Biotechnology** 3(1):19-24.
- Molyneux P (2004) The use of the stable free radical diphenylpicryl-hydrazyl (DPPH) for estimating antioxidant activity. **Songklanakarin Journal of Science and Technology** 26(2):211-219.
- Moss EH (1926) The uredo stage of the Pucciniastreae. Annals of Botany 40(160):813-847.
- Muir CD (2020) A stomatal model of anatomical tradeoffs between gas exchange and pathogen colonization. Frontiers in Plant Science 11:1631.
- Nakamura Y, Tsuji S, Tonogai Y (2003) Analysis of proanthocyanidins in grape seed extracts, health foods and grape seed oils. **Journal of Health Science** 49(1):45-54.
- Navarro BL, Marques JP, Appezzato-da-Glória B, Spósito MB (2019) Histopathology of *Phakopsora euvitis* on *Vitis vinifera*. **European Journal of Plant Pathology** 154(4):1185-1893.
- Nelson S (2011) Raspberry late leaf rust in Hawaii caused by *Pucciniastrum americanum*. **Plant Disease** 5.
- Nickerson NL (1991) Late leaf rust. In: Ellis MA, Converse RH, Williams R.N, Williamson B (Ed.). **Compendium of Raspberry and Blackberry Diseases and Insects.** Saint Paul, APS Press 30-32.
- Nogueira Júnior AF, Ribeiro RV, Appezzato-da-Glória B, Soares MK, Rasera JB, Amorim L (2017) *Phakopsora euvitis* causes unusual damage to leaves and modifies carbohydrate metabolism in grapevine. **Frontiers in Plant Science** 8:1675.
- Oszmiański J, Wojdyło A, Nowicka P, Teleszko M, Cebulak T, Wolanin M (2015) Determination of phenolic compounds and antioxidant activity in leaves from wild *Rubus* L. species. **Molecules** 20(3):4951-4966.
- Oliveira PB (2021) Manual de Boas Práticas de Fruticultura Framboesa. **Revista Frutas**, **Legumes e Flores** (INIAV) 215(7).
- Paul V, Sharma L, Pandey R, Meena RC (2017) Measurements of stomatal density and stomatal index on leaf/plant surfaces. In: Manual of ICAR Sponsored Training Programme for Technical Staff of ICAR Institutes on "Physiological Techniques to Analyze the Impact of Climate Change on Crop Plants". Bombay, India.
- Perera FM, Bertani PR, Arias EM, Hechavarria MD, Navarro ZMD, Debes AM, Luque AC, Cuenya IM, Rojas R, Castagnaro PA (2020) Morphological and molecular characterization

of *Puccinia kuehnii*, the causal agent of sugarcane orange rust, in Cuba. **Scientia Agricola** 77(2): e20180038.

- Pio R (2014) Cultivo de fruteiras de clima temperado em regiões subtropicais e tropicais, 1st ed. Lavras, UFLA (4):223-248.
- Primiano IV, Loehrer M, Schaffrath U, Amorim L (2019) Formation of satellite uredinia as an important trait related to grapevine colonization by *Phakopsora meliosmae-myrianthae*. **Plant Pathology** 68(9): 1732-1740.
- Rasband WS (2018) Image J. United States National Institute of Health, Bethesda, Maryland.
- Rasera JB, Amorim L, Marques JPR, Soares MK, Appezzato-da-Glória B (2019) Histopathological evidences of early grapevine leaf senescence caused by *Phakopsora euvitis* colonisation. **Physiological and Molecular Plant Pathology** 108:101434.
- Raseira MCB, Gonçalves ED, Trevisan R, Antunes LEC (2004) Aspectos técnicos da cultura da framboeseira. Pelotas: Embrapa Clima Temperado (Documento, 120) 22.
- Ribeiro RR, Spósito MB (2022) Interference of late rust associated with water deficit in the primary metabolism of raspberries. **European Journal of Plant Pathology** 163(2):279-292.
- Rincón-Barón EJ, Gutiérrez Rodríguez AM, Guerra BE, Espinosa Matías S (2020) Alteraciones histopatológicas causadas por la roya *Puccinia nakanishikii* (Pucciniales: Pucciniaceae) en plantas de *Cymbopogon citratus* (Poaceae). **Revista de Biología Tropical** 68(2):361-382.
- RStudio Team (2018) RStudio: integrated development for R. Rstudio, Inc., Boston, MA http://www.rstudio.com/
- Sakai WS (1973) Simple method for differential staining of paraffin embedded plant material using toluidine blue O. **Stain Technology** 48:247-324.
- Scholler M, Braun U, Buchheit R, Schulte T, Bubner B (2022) Studies on European rust fungi, *Pucciniales*: molecular phylogeny, taxonomy, and nomenclature of miscellaneous genera and species in Pucciniastraceae and Coleosporiaceae. **Mycological Progress** 21(8):1-25.
- Sharma OP, Bhat TK (2009) DPPH antioxidant assay revisited. Food Chemistry 113(4):1202-1205.
- Shibu Prasanth SC, Chandran P (2017) Phytochemical and antimicrobial analysis of leaf samples of different *Rubus* species. **International Journal of ChemTech Research** 10(4):359-368.
- Silva MD, Várzea V, Guerra-Guimarães L, Azinheira HG, Fernandez D, Petitot AS, Bertrand B, Lashermes P, Nicole M (2006) Coffee resistance to the main diseases: leaf rust and coffee berry disease. **Brazilian Journal of Plant Physiology** 18:119-147.
- Singleton VL, Rossi JA (1965) Colorimetry of total phenolics with phosphomolybdicphosphotungstic acid reagents. American Journal of Enology and Viticulture 16(3):144-158.
- Solanki S, Gazala A, Borowicz P, Brueggeman RS (2019) Shedding light on penetration of cereal host stomata by wheat stem rust using improved methodology. **Nature** 9:7939.

- Tomaszewski D, Zieliński J, Gawlak M (2014) Foliar indumentum in central-European *Rubus* species (Rosaceae) and its contribution to the systematics of the group. **Nordic Journal of Botany** 32(1):1-10.
- Unger S, Büche C, Boso S, Kassemeyer HH (2007) The course of colonization of two different *Vitis* genotypes by *Plasmopara viticola* indicates compatible and incompatible host-pathogen interactions. **Phytopathology** 97(7):780-786.
- Upadhyaya MK, Furness NH (1998) Primocane morphology and leaf surface characteristics of greenhouse-grown red raspberry cultivars. **HortScience** 33:330-332.
- Wang Y, Zeng J, Xia X, Xu Y, Sun J, Gu J, Sun H, Lei H, Chen F, Jiang J, Fang W (2020) Comparative analysis of leaf trichomes, epidermal wax and defense enzymes activities in response to *Puccinia horiana* in *Chrysanthemum* and *Ajania* species. **Horticultural Plant Journal** 6(3):191-198.
- Wellburn AR (1994) The spectral determination of chlorophylls a and b, as well as total carotenoids, using various solvents with spectrophotometers of different resolution. **Journal** of Plant Physiology 144(3): 307-313.
- Yang H, Han S, He D, Jiang S, Cao G, Wan X, Chen L, Xiao J, Zhu P (2021) Resistance evaluation of walnut (*Juglans* spp.) against *Xanthomonas arboricola* and the correlation between leaf structure and resistance. **Forest Pathology** 51(1): e12659.
- Yong WT, Ades PK, Tibbits JF, Bossinger G, Runa FA, Sandhu KS, Taylor PW (2019) Disease cycle of *Austropuccinia psidii* on *Eucalyptus globulus* and *Eucalyptus obliqua* leaves of different rust response phenotypes. **Plant Pathology** 68(3):547-555.

3. A NEW HYBRID BETWEEN RED AND BLACK RASPBERRY AND ITS RESPONSE TO LATE LEAF RUST FROM A HISTOPATHOLOGICAL VIEW

Abstract

Rubus idaeus 'Heritage' is a red raspberry cultivated worldwide for fresh or processed consumption. However, 'Heritage' is susceptible de late leaf rust, a disease caused by Aculeastrum americanum (Farl.) M. Scholler & U. Braun (syn. Thekopsora americana (Farl.) Aime & McTaggart). Genetic resistance is a promising way to manage this disease. Hybridization between red and black raspberries is commonly used for their genetic improvement. The wild black raspberry R. niveus is a reported as hight-value progenitor resistant to diseases. In fact, R. niveus exhibited remarkable histochemical responses to A. americanum. In this study, we characterized the leaf anatomy of a hybrid between R. niveus and R. idaeus 'Heritage' and its biochemical and histopathological responses to A. americanum. Our findings showed that the hybrid leaves inherited the morphology phenotype from *R. idaeus* and leaf anatomical traits from R. niveus. Only small pustules per leaf were observed 20 days after inoculation with A. americanum. In the lesioned area, the epidermal and mesophyll cells underwent collapses and exhibited phenol accumulation. This work sheds light on the histopathological interactions in the pathosystem A. americanum-raspberry and highlights traits that can help plant breeders target late leaf rust resistance in future cultivars.

Keywords: Aculeastrum americanum, defense mechanisms, phenolic compounds, *Pucciniastrum americanum, Rubus idaeus, Rubus niveus*

Introduction

Raspberries belong to the genus *Rubus*, a member of the globally-important Rosaceae family (Graham et al. 2018; Chwil and Kostryco 2020). As a result of consumer demand and research into their bioactive compounds, global production of red and black raspberries has increased during the last couple of years to reach 822,000 tonnes in 2021 (Bushakra et al. 2015; Foster et al. 2019; Klewicka et al. 2020; FAO 2021).

Plant breeding programs around the world have developed new cultivars in order to meet this demand and ensure fruit quality. Raspberry breeding focus on several agronomic traits, including pest and disease resistance (Hall et al. 2009; Zasada and Moore 2014; Foster et al. 2019). Because red and black raspberries are in the same subgenus *Idaeobatus* they have been crossed to improve traits (Hall et al. 2009; Martin et al. 2018; Foster et al. 2019). Successful hybridization has been reported between *Rubus idaeus* L. cultivars and the black raspberry *R. occidentalis* L. (Bushakra et al. 2012; Foster et al. 2019). Despite this, further research using other species of black raspberries remains to be done.

R. niveus Thunb. was previously classified as a high-value progenitor in North American breeding programs due to its cane and leaf disease resistance, among other agronomic and physiological characters (Finn et al. 2002; Zasada and Moore 2014). Indeed, *R. niveus* has displayed structural and biochemical distinctive responses to *Aculeastrum americanum*, the causal agent of late leaf rust in raspberries (Chapter 2). Although *R. idaeus* 'Heritage' is susceptible to late leaf rust, it is cultivated worldwide, performing high-yielding and good fruit quality (Luffman and Buszard 1990; Volk et al. 2013; Chapter 2). Combining these traits could result in new cultivars that are more productive and resistance to diseases.

Current work in an associated research group has performed classical breeding between different raspberry cultivars (Barbosa 2022) and species, including one between *R. niveus* as female parent and *R. idaeus* 'Heritage'. In our study, we aim to characterize the leaf morphoanatomy of the F1 hybrid and to investigate its histopathological and biochemical responses to *A. americanum*.

Conclusion

NH, as a new raspberry cultivar, is still in its early stages since other agronomic traits need to be explored. Nevertheless, considering that resistance to disease is one target in plant breeding, our results documented novelties about ultrastructural, histopathological, and biochemical responses of a hybrid offspring between *Rubus niveus* x *Rubus idaeus* 'Heritage' with potential resistance to *A. americanum*.

- Alves RF, Marques JPR., Appezzato-da-Glória B, Spósito MB (2021) Process of infection and colonization of *Pseudocercospora kaki* in persimmon leaves. Journal of Phytopathology 169(3):168-175.
- Azinheira HG, Silva MC, Talhinhas P, Medeira C, Maia I, Petitot A, Fernandez D (2010) Non-host resistance responses of *Arabidopsis thaliana* to the coffee leaf rust fungus (*Hemileia vastatrix*). **Botany** 88:621-629.
- Barbosa CMA (2022) Seleção de híbridos de framboeseiras desenvolvidos por melhoramento convencional Tese (Doutorado em Agronomia) - Escola Superior de Agricultura "Luiz de Queiroz".
- Braga ZV, Santos RF, Amorim L, Appezzato-da-Glória B (2019) Histopathology of infection and colonisation of *Elsinoë ampelina* on grapevine leaves. **European Journal of Plant Pathology** 154(4):1009-1019.

- Braga ZV, Muniz LF, Manarim GR, de Aguiar CL, Appezzato-da-Glória B (2021) Anatomical and biochemical changes in leaves of *Vitis labrusca* L. cv. Niagara Rosada in response to infection by *Elsinoë ampelina* Shear. **Brazilian Journal of Botany** 44(1):187-196.
- Bushakra JM, Stephens MJ, Atmadjaja AN, Lewers KS, Symonds VV, Udall JA, Chagné D, Buck EJ, Gardiner SE (2012) Construction of black (*Rubus occidentalis*) and red (*R. idaeus*) raspberry linkage maps and their comparison to the genomes of strawberry, apple, and peach. **Theoretical and Applied Genetics** 125(2):311-327.
- Bushakra JM, Bryant DW, Dossett M, Vining KJ, VanBuren R, Gilmore BS, Lee J, Mockler TC, Finn CE, Bassil NV (2015) A genetic linkage map of black raspberry (*Rubus occidentalis*) and the mapping of Ag 4 conferring resistance to the aphid *Amphorophora agathonica*. **Theoretical and Applied Genetics** 128(8):1631-1646.
- Bettgenhaeuser J, Gilbert B, Ayliffe M, and Moscou MJ (2014) Nonhost resistance to rust pathogens a continuation of continua. **Frontiers in Plant Science** 5:664.
- Castro MC, Demarco D (2008) Phenolic compounds produced by secretory structures in plants: a brief review. **Natural Product Communications** 3(8): 1273-1284.
- Cheng Y, Zhang H, Yao J, Wang X, Xu J, Han Q, Wei G, Huang L, Kang Z. (2012) Characterization of non-host resistance in broad bean to the wheat stripe rust pathogen. **BMC Plant Biology** 12(1):1-2.
- Chwil M, Kostryco M (2020) Histochemical assays of secretory trichomes and the structure and content of mineral nutrients in *Rubus idaeus* L. leaves. **Protoplasma** 257: 119-139.
- Costea T, Vlase L, Gostin IN, Olah NK, Predan GM (2016) Botanical characterization, phytochemical and antioxidant activity of indigenous red raspberry (*Rubus idaeus* L.) leaves. **Studia Universitatis Vasile Goldis. Seria Stiintele Vietii** 26:463-472.
- Darker G D (1929) Cultures of *Pucciniastrum Americanum* (Farlow) Arthur and *P. Arcticum* (Lagerheim) Tranzschel. Journal of the Arnold Arboretum 10(3): 156-167.
- Duplessis S, Lorrain C, Petre B, Figueroa M, Dodds PN, Aime MC (2021) Host adaptation and virulence in heteroecious rust fungi. Annual Review of Phytopathology 59:403-422.
- FAO (2021) FAOSTAT: agricultural data. http://www.fao.org/faostat/en/#data/QC [accessed 2022 Mar 28].
- Fell KR, Rowson JM (1956) Anatomical studies in the genus *Rubus*: Part I. The Anatomy of the Leaf of *Rubus idaeus* L. Journal of Pharmacy and Pharmacology 8(1): 334-345.
- Fell KR, Rowson JM (1961) Anatomical studies in the genus *Rubus*: Part IV. Anatomical variations in the leaves of cultivated varieties of *R. idaeus* L. and *R. loganobaccus* L.H. Bailey, and of certain species of Bramble. Journal of Pharmacy and Pharmacology 13(1): 83-92.
- Finn C, Swartz H, Moore PP, Ballington JR, Kempler C (2002) Use of 58 Rubus species in five North American breeding programmes-Breeders notes. Acta Horticulturae 1:113-120.
- Foster TM, Bassil NV, Dossett M, Worthington ML, Graham J (2019) Genetic and genomic resources for Rubus breeding: a roadmap for the future. **Horticulture Research** 6:116.

Graham J, Brennan R (2018) Raspberry: breeding, challenges and advances. Springer.

- Hall HK, Hummer K, Jamieson AJ, Jennings SN, Weber CA (2009) Raspberry breeding and genetics. **Plant Breeding Reviews** 32:1-290.
- Luffman M, Buszard D (1989) A note on the susceptibility of six red raspberry cultivars and Tayberry to fruit infection by late yellow rust. **Phytoprotection** 71:93-95.
- Hiscox JD, Israelstam GF (1979) A method for the extraction of chlorophyll from leaf tissue without maceration. **Canadian Journal of Botany** 57(12):1332-1334.
- Hoch HC, Staples RC (1987) Structural and chemical changes among the rust fungi during appressorium development. **Annual Review of Phytopathology** 25(1):231-247.
- Horridge GA, Tamm SL (1969) Critical point drying for scanning electron microscopic study of ciliary motion. **Science** 163(3869):817-818.
- Jensen WA (1962) Botanical histochemistry: principles and practice. Freeman, San Francisco.
- Johansen DA (1940) Plant microtechnique. McGraw-Hill, New York.
- Karnovsky JM (1965) A formaldehyde-glutaraldehyde fixative of high osmolality for use in electron microscopy. **Journal of Cell Biology** 27:137-138.
- Klewicka E, Sójka M, Ścieszka S, Klewicki R, Milczarek A, Lipińska L, Kołodziejczyk K (2020) The antimycotic effect of ellagitannins from raspberry (*Rubus idaeus* L.) on Alternaria *alternata* ŁOCK 0409. **European Food Research and Technology** 246(7): 1341-1349.
- Kretschmer M, Damoo D, Djamei A, Kronstad J (2019) Chloroplasts and plant immunity: where are the fungal effectors? **Pathogens** 9(1):19.
- Luffman M, Buszard D (1989) A note on the susceptibility of six red raspberry cultivars and Tayberry to fruit infection by late yellow rust. **Phytoprotection** 71:93-95.
- Martin RR, Ellis MA, Williamson B, Williams RN (2018) PART I: Diseases Caused by Biotic Factors. In: Martin et al. (Ed.). **Compendium of Raspberry and Blackberry Diseases and Pests**, Second Edition, APS Press.
- Marques JP, Kitajima EW, Freitas-Astúa J, Appezzato-da-Glória B (2010) Comparative morpho-anatomical studies of the lesions caused by citrus leprosis virus on sweet orange. Anais da Academia Brasileira de Ciências 82:501-511.
- Marques JPR, Cia MC, Granato, ABA, Muniz LF, Appezzato-da-Glória B, Camargo LEA (2022) Histopathology of the shoot apex of sugarcane colonized by *Leifsonia xyli* subsp. *xyli*. **Phytopathology** 112(10): 2062-2071.
- Molyneux P (2004) The use of the stable free radical diphenylpicryl-hydrazyl (DPPH) for estimating antioxidant activity. **Songklanakarin Journal of Science and Technology** 26(2):211-219.
- Murria S, Kaur N, Arora NK, Mahal AK (2018) Field reaction and metabolic alterations in grape (*Vitis vinifera* L.) varieties infested with anthracnose. **Scientia Horticulturae** 235:286-293.

- Navarro BL, Marques JP, Appezzato-da-Glória B, Spósito MB (2019) Histopathology of Phakopsora euvitis on Vitis vinifera. **European Journal of Plant Pathology** 154(4):1185-1193.
- Nogueira Júnior AF, Ribeiro RV, Appezzato-da-Glória B, Soares MK, Rasera JB, Amorim L (2017) *Phakopsora euvitis* causes unusual damage to leaves and modifies carbohydrate metabolism in grapevine. **Frontiers in Plant Science** 8:1675.
- Rasera JB, Amorim L, Marques JPR, Soares MK, Appezzato-da-Glória B (2019) Histopathological evidences of early grapevine leaf senescence caused by *Phakopsora euvitis* colonisation. **Physiological and Molecular Plant Pathology** 108:101434.
- RStudio Team (2018) RStudio: integrated development for R. Rstudio, Inc., Boston, MA http://www.rstudio.com/
- Sakai WS (1973) Simple method for differential staining of paraffin embedded plant material using toluidine blue O. **Stain Technology** 48:247-324.
- Sharma OP, Bhat TK (2009) DPPH antioxidant assay revisited. Food Chemistry 113(4):1202-1205.
- Scholler M, Braun U, Buchheit R, Schulte T, Bubner B (2022) Studies on European rust fungi, Pucciniales: molecular phylogeny, taxonomy, and nomenclature of miscellaneous genera and species in Pucciniastraceae and Coleosporiaceae. Mycological Progress 21(8):1-25.
- Singleton VL, Rossi JA (1965) Colorimetry of total phenolics with phosphomolybdicphosphotungstic acid reagents. **American Journal of Enology and Viticulture** 16(3):144-158.
- Volk GM, Olmstead JW, Finn CE, Janick J (2013) The ASHS outstanding fruit cultivar award: a 25-year retrospective. **HortScience** 48(1):4-12.
- Yu ZD, Peng SB, Ren ZZ, Wang DM, Cao ZM (2011) Infection behaviour of *Melampsora larici-populina* on the leaf surface of *Populus purdomii*. Agricultural Sciences in China 10:1562-1569.
- Wan Z, Li Y, Liu M, Chen Y, Yin T (2015) Natural infectious behavior of the urediniospores of *Melampsora larici-populina* on poplar leaves. Journal of Forestry Research 26(1):225-231.
- Zasada IA, Moore PP (2014) Host status of Rubus species and hybrids for the root lesion nematode, *Pratylenchus penetrans*. HortScience 49(9):1128-1131.Wellburn AR (1994) The spectral determination of chlorophylls a and b, as well as total carotenoids, using various solvents with spectrophotometers of different resolution. Journal of Plant Physiology 144(3): 307-313.

4. INITIAL CHARACTERIZATION OF THE CALCIUM-DEPENDENT PROTEIN KINASE CPK28 IN BLACK RASPBERRY AND STRAWBERRY

Abstract

Plant diseases are a threat to the maintenance of biodiversity and the production of food. Throughout their evolutionary history, plants have evolved several mechanisms for dealing with environmental and biotic stressors. When plants recognize microbes, it triggers signals and responses by different pathways, including those with Ca2+-dependent protein kinases (CDPKs). These kinases are evolutionarily conserved, however, only a few have been described in the Rosaceae family, which includes important species such as raspberry, and strawberry. In this work, we use comparative genomics to identify CDPK orthologs in black raspberry (Rubus occidentalis) and strawberry (Fragaria vesca L.) as representative of the globally important Rosaceae family. We found that R. occidentalis and F. vesca have together 26 CDPKs. We focused subsequent analysis on those in subgroup group IV, due to their roles in plant immunity already described in other species such as rice, cotton and tobacco. To determine orthology with well-studied Arabidopsis subgroup IV CDPK, CPK28, we cloned and expressed FvCPK28 and RoCPK28 in Nicotiana benthamiana and Arabidopsis cpk28 mutants to assess genetic complementation. We found that both Rosaceae CPK28 orthologs are plasma-membrane localized and their overexpression dampened the oxidative burst upon elicitation. Further investigations must be conducted to ascertain the functional role of Rosaceae CPK28 orthologs. Nevertheless, this study provided a proof-of-concept to investigate the functional similarities between these proteins and Arabidopsis CPK28.

Keywords: Arabidopsis, CDPK, Fragaria vesca, PAMP-triggered immunity, oxidative burst, Rubus occidentalis

Introduction

Raspberries and strawberries, both members of the Rosaceae family, are widely cultivated horticultural crops (Foster et al. 2019; Chebotar et al. 2022). Due to their relevance, healthy plants are essential to local and global agricultural economies. However, a variety of diseases can affect these plants, including viruses, fungi and bacteria (Silva et al. 2017; Martin et al. 2018). The black raspberry *Rubus occidentalis* L., for example, is seriously impacted by anthracnose (*Elsinoë necator*) and is reported as not immune to *Aculeastrum americanum*, the causal agent of late leaf rust (Martin et al. 2018; Chapter 2). Likewise, the woodland strawberry *Fragaria vesca* L. is vulnerable to diseases such as angular leaf spot and grey mold, caused by the pathogens *Xanthomonas fragariae* and *Botrytis cinerea*, respectively (Silva et al. 2017; Badmi et al. 2022).

Although plants can succumb to disease, resistance is the rule rather than the exception (Staskawicz 2001; Ávila-Méndez and Romero 2017). Plants have evolved several pre and pos-formed structural, biochemical and molecular mechanisms for dealing with environmental and biotic stressors (Jones and Dangl 2006; DeFalco and Zipfel 2021; Chapter 2; Chapter 3). If these pre-formed defense mechanisms are not enough to stop a pathogen invasion, the plant-pathogen interaction is raised to another level (Jones and Dangl 2006; Kaur et al. 2022). When plants recognize a pathogen, a cascade of responses takes place, resulting in changes in biological processes that can confer resistance to the pathogen (Couto and Zipfel 2016; Bentham et al. 2020).

The plant basal defense is activated by the recognition of pathogen-associated molecular patterns (PAMPs), which are perceived by pattern recognition receptors (PRRs), located on the plasma membrane surface (Hogenhout et al. 2009; Bentham et al. 2020; Defalco and Zipfel 2021). Upon recognition, PRRs form complexes with other proteins and/or receptor-like cytoplasmic kinases (RLCKs; Saijo et al. 2018, Bentham et al. 2020; Defalco and Zipfel 2021). The BOTRYTIS-INDUCED KINASE 1 (BIK1) is an RLCK considered a key plant immune signalling protein since it mediates immune responses triggered by several PAMPs (Monaghan et al. 2014; Dias et al. 2022). For this, BIK1 activity is fine-tuned through post-translational modifications such as phosphorylation (Dias et al. 2022).

The Ca²⁺-dependent protein kinases (CDPKs or CPKs) are signal transducers that play a crucial role as regulators in biological processes in plants, such as innate immune responses (Bredow and Monaghan 2019; Crizel et al. 2020). Although these proteins were identified for several species, information for Rosaceae remains scarce (Cheng et al. 2002; Wang et al. 2018; Crizel et al. 2020).

The CDPKs are divided into four main groups that are evolutionarily conserved in plants but not found in animals and fungi (Boudsocq et al. 2010; Monaghan et al. 2014; Valmonte et al. 2014; Crizel et al. 2020). In group IV CDPKs, the CPK28 has been described as a negative regulator of PAMP-induced signalling and indicated as a hub in plant immunity responses (Bredow and Monaghan 2019; Delormel and Boudsocq 2019). Indeed, *Arabidopsis* plants CPK28 loss-of-function enhance the stress and immune response. Furthermore, previous studies have shown the possibility of dynamic regulation between CPK28 and BIK1, demonstrating that cellular homeostasis requires tight regulation (Monaghan et al. 2014; 2015; Wang et al. 2018; Wu et al. 2021). Genome-wide comparisons between *Rubus* and *Fragaria* have supported high collinearity between these species (Bushakra et al. 2012; Foster et al. 2019; Wight et al. 2019). Since none of these mechanisms mentioned above are well understood in *R. occidentalis* and *F. vesca*, we aim to identify and analyze the CDPK family across these two Rosaceae species. Additionally, we seek to assess the conservation of group IV CDPKs using a transgenic approach to characterize and ascertain the functional role of the black raspberry and strawberry CPK28 orthologs.

Concluding remarks and future directions

This study has identified for the first time the CDPKs across *R. occidentalis* and *F. vesca* and encountered the CPK28 orthologs for these species.

The subcellular localization of FvCPK28 and RoCPK28 in *N. benthamiana* was an efficient tool to indicate their presence at the plasma membrane. Additionally, the elicitor-triggered oxidative burst assay performed in tobacco tissues expressing CPK28 orthologs was a proof of concept indicating that these proteins play the same biological role in different plants. Nevertheless, determining whether CPK28 orthologs can complement the immune phenotype of *cpk28-1* mutants requires more investigation.

Further studies must be conducted to ascertain the functional role of the Rosaceae CPK28 orthologs because the function of a protein can only be accurately explored by biochemical and structural investigations (Fang et al. 2010). For example, the orthology inferences can be integrated with gene expression patterns analysis and the CPK28 interactions with well-known partners in the same protein complex.

In conclusion, this work shed light on the significance of CPK28 role in immune signaling for the studied Rosaceae species.

- Amil-Ruiz F, Blanco-Portales R, Munoz-Blanco J, Caballero JL (2011) The strawberry plant defense mechanism: a molecular review. Plant and Cell Physiology 52(11): 1873-1903.
- Ávila-Méndez K, Romero HM (2017) Plant responses to pathogen attack: molecular basis of qualitative resistance. **Revista Facultad Nacional de Agronomía Medellín** 70(2):8225-35.
- Badmi R, Tengs T, Brurberg MB, Elameen A, Haugland LK, Fossdal CG, Hytonen T, Krokene P, Thorstensen T (2022) Transcriptional profiling of defence responses to Botrytis cinerea infection in leaves of *Fragaria vesca* plants soil-drenched with β-aminobutyric acid (BABA). **Frontiers in Plant Science** 8:5015.

Benchling (2022) [Biology Software] Retrieved from https://benchling.com.

- Bentham AR, De la Concepcion JC, Mukhi N, Zdrzałek R, Draeger M, Gorenkin D, Hughes RK, Banfield MJ (2020) A molecular roadmap to the plant immune system. Journal of Biological Chemistry 295(44):14916-14935.
- Boudsocq M, Sheen J (2013) CDPKs in immune and stress signaling. **Trends in Plant Science** 18(1):30-40.
- Bredow M, Monaghan J (2019) Regulation of plant immune signaling by calcium-dependent protein kinases. **Molecular Plant-Microbe Interactions** 32(1):6-19.
- Bredow M, Sementchoukova I, Siegel K, Monaghan J (2019) Pattern-triggered oxidative burst and seedling growth inhibition assays in *Arabidopsis thaliana*. **JoVE (Journal of Visualized Experiments)** (147):e59437.
- Bushakra JM, Stephens MJ, Atmadjaja AN, Lewers KS, Symonds VV, Udall JA, Chagné D, Buck EJ, Gardiner SE (2012) Construction of black (*Rubus occidentalis*) and red (*R. idaeus*) raspberry linkage maps and their comparison to the genomes of strawberry, apple, and peach. **Theoretical and Applied Genetics** 125(2):311-27.
- Chebotar VK, Chizhevskaya EP, Vorobyov NI, Bobkova VV, Pomyaksheva LV, Khomyakov YV, Konovalov SN (2022) The Quality and Productivity of Strawberry (*Fragaria* x *ananassa* Duch.) Improved by the Inoculation of PGPR *Bacillus velezensis* BS89 in Field Experiments. **Agronomy** 12(11):2600.
- Cheng SH, Willmann MR, Chen HC, Sheen J (2002) Calcium signaling through protein kinases. The *Arabidopsis* calcium-dependent protein kinase gene family. **Plant Physiology** 129(2): 469-485.
- Clough SJ, Bent AF (1998) Floral dip: a simplified method for Agrobacterium-mediated transformation of *Arabidopsis thaliana*. **The Plant Journal** 16(6): 735-743.
- Crizel RL, Perin EC, Vighi IL, Woloski R, Seixas A, da Silva Pinto L, Rombaldi CV, Galli V (2020) Genome-wide identification, and characterization of the CDPK gene family reveal their involvement in abiotic stress response in *Fragaria* x *ananassa*. **Scientific Reports** 10(1): 1-17.
- Couto D, Zipfel C (2016) Regulation of pattern recognition receptor signalling in plants. **Nature Reviews Immunology** 16(9):537-552.
- David R, Menezes RJ, De Klerk J, Castleden IR, Hooper CM, Carneiro G, Gilliham M (2021) Identifying protein subcellular localisation in scientific literature using bidirectional deep recurrent neural network. Scientific Reports 11(1):1-11.
- DeFalco TA, Zipfel C (2021) Molecular mechanisms of early plant pattern-triggered immune signaling. **Molecular Cell** 81(17):3449-67.
- Delormel TY, Boudsocq M (2019) Properties and functions of calcium-dependent protein kinases and their relatives in *Arabidopsis thaliana*. **New Phytologist** 224(2):585-604.
- Dias MG, Soleimani F, Monaghan J (2022) Activation and turnover of the plant immune signaling kinase BIK1: a fine balance. **Essays in Biochemistry** 66(2):207-218.
- Fang G, Bhardwaj N, Robilotto R, Gerstein MB (2010) Getting started in gene orthology and functional analysis. **PLoS Computational Biology** 6(3):e1000703.

- Foster TM, Bassil NV, Dossett M, Worthington ML, Graham J (2019) Genetic and genomic resources for *Rubus* breeding: a roadmap for the future. **Horticulture Research** 6:116.
- Glover N, Dessimoz C, Ebersberger I, Forslund SK, Gabaldón T, Huerta-Cepas J, Thomas PD (2019) Advances and applications in the quest for orthologs. **Molecular Biology and Evolution** 36(10): 2157-2164.
- Gong Z, Shao ZQ, Chen JQ, Han GZ (2022) Plant immune receptors evolved hand in hand. **Nature Plants** 8(10):1138-1139.
- Hogenhout SA, Van der Hoorn RA, Terauchi R, Kamoun S (2009) Emerging concepts in effector biology of plant-associated organisms. **Molecular Plant-Microbe Interactions** 22(2):115-122.
- Hu Z, Li J, Ding S, Cheng F, Li X, Jiang Y, Yu J, Foyer CH, Shi K (2021) The protein kinase CPK28 phosphorylates ascorbate peroxidase and enhances thermotolerance in tomato. **Plant Physiology** 186:1302-1317.

Jones J, Dangl J (2006) The plant immune system. Nature 444: 323-329.

- Jung S, Lee T, Cheng CH, Buble K, Zheng P, Yu J, Humann J, Ficklin SP, Gasic K, Scott K, Frank M (2018) 15 years of GDR: New data and functionality in the Genome Database for Rosaceae. **Nucleic Acids Research** 47(D1):D1137-1145.
- Karimi M, Inzé D, Depicker A (2002) GATEWAY[™] vectors for Agrobacterium-mediated plant transformation. **Trends in Plant Science** 7(5):193-195.
- Kaur S, Samota MK, Choudhary M, Choudhary M, Pandey AK, Sharma A, Thakur J (2022) How do plants defend themselves against pathogens-Biochemical mechanisms and genetic interventions. **Physiology and Molecular Biology of Plants** 28(2): 485-504.
- Martin RR, Ellis MA, Williamson B, Williams RN (2018) PART I: Diseases Caused by Biotic Factors. In: Martin et al. (Ed.). **Compendium of Raspberry and Blackberry Diseases and Pests**, Second Edition, APS Press.
- Matschi S, Werner S, Schulze WX, Legen J, Hilger HH, Romeis T (2013) Function of calcium-dependent protein kinase CPK 28 of *Arabidopsis thaliana* in plant stem elongation and vascular development. **The Plant Journal** 73(6):883-896.
- Monaghan J, Matschi S, Shorinola O, Rovenich H, Matei A, Segonzac C, Malinovsky FG, Rathjen JP, MacLean D, Romeis T, Zipfel C (2014) The calcium-dependent protein kinase CPK28 buffers plant immunity and regulates BIK1 turnover. Cell Host Microbe 16(5):605-615.
- Monaghan J, Matschi S, Romeis T, Zipfel C (2015) The calcium-dependent protein kinase CPK28 negatively regulates the BIK1-mediated PAMP-induced calcium burst. **Plant Signaling and Behavior** 10(5):e1018497.
- Monaghan J (2018) Conserved degradation of orthologous RLCKs regulates immune homeostasis. **Trends in Plant Science** 23(7):554-557.
- Mueller K, Chinchilla D, Albert M, Jehle AK, Kalbacher H, Boller T, Felix G (2012) Inadvertent cross-contamination as a risk for work with synthetic peptides: flg22 as an example of a pirate peptide in commercial peptide preparations. **Plant Cell** 24:2213-24.

- Saijo Y, Loo EP, Yasuda S (2018) Pattern recognition receptors and signaling in plantmicrobe interactions. The Plant Journal 93(4):592-613.
- Saitou N, Nei M (1987) The neighbor-joining method: A new method for reconstructing phylogenetic trees. **Molecular Biology and Evolution** 4:406-425.
- Silva KJ, Brunings AM, Pereira JA, Peres NA, Folta KM, Mou (2017) The Arabidopsis *ELP3/ELO3* and *ELP4/ELO1* genes enhance disease resistance in *Fragaria vesca* L. **BMC Plant Biology** 17(1):1-2.
- Staskawicz BJ (2001) Genetics of plant-pathogen interactions specifying plant disease resistance. **Plant Physiology** 125(1):73-6.
- Stecher G, Tamura K, Kumar S (2020) Molecular evolutionary genetics analysis (MEGA) for macOS. **Molecular Biology and Evolution** 37(4):1237-1239.
- TheArabidopsisInformationResource(TAIR),[https://www.arabidopsis.org/servlets/TairObject?id=132313&type=locus],onwww.arabidopsis.org, [Dec 2021].on
- Valmonte GR, Arthur K, Higgins CM, MacDiarmid RM (2014) Calcium-dependent protein kinases in plants: evolution, expression and function. Plant and Cell Physiology 55(3):551-569.
- Voinnet O, Rivas S, Mestre P, Baulcombe D (2003) Retracted: An enhanced transient expression system in plants based on suppression of gene silencing by the p19 protein of tomato bushy stunt virus. **The Plant Journal** 33(5):949-956.
- Wang J, Wang S, Hu K, Yang J, Xin X, Zhou W, Fan J, Cui F, Mou B, Zhang S, Wang G, Sun W (2018) The kinase OsCPK4 regulates a buffering mechanism that fine-tunes innate immunity. **Plant Physiology** 176(2):1835-1849.
- Wight H, Zhou J, Li M, Hannenhalli S, Mount S, Liu Z (2019) Draft genome assembly and annotation of red raspberry *Rubus Idaeus*. **bioRxiv** 1-22.
- Wu Y, Zhang L, Zhou J, Zhang X, Feng Z, Wei F, Zhao L, Zhang Y, Feng H, Zhu H (2021) Calcium-dependent protein kinase GhCDPK28 was identified and involved in verticillium wilt resistance in cotton. Frontiers in Plant Science 12:772649.
- Yamada K, Yamaguchi K, Shirakawa T, Nakagami H, Mine A, Ishikawa K, Fujiwara M, Narusaka M, Narusaka Y, Ichimura K, Kobayashi Y (2016) The *Arabidopsis* CERK1associated kinase PBL27 connects chitin perception to MAPK activation. The EMBO Journal 35(22):2468-2483.