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**Agro-industrial residues from Amazonian fruits: bioactive composition,
bioaccessibility and potential use as a prebiotic**

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Thesis presented for obtaining the title of Doctor in
Science. Area: Food Science and Technology

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Bacharel in Food Science

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RESUMO

Resíduos agroindustriais de frutos amazônicos: composição bioativa, bioacessibilidade e potencial de uso como prebiótico

A intensa geração global de resíduos e subprodutos resultantes das atividades agroindustriais sinaliza a necessidade de estudos para reaproveitamento destes materiais. Este trabalho teve como objetivo determinar a composição nutricional, bioativa e o potencial prebiótico de subprodutos (tortas) do processamento do açaí (*Euterpe oleracea* Mart) e do inajá (*Maximiliana maripa* Aubl Drude) como fontes naturais de nutrientes e compostos com atividades antioxidante e anti-inflamatória. As tortas de açaí e inajá foram provenientes do processamento dos frutos para extração de óleo. Inicialmente, foram determinados a composição centesimal e o perfil mineral dos materiais brutos, seguido do potencial prebiótico utilizando bactérias probióticas (*Lacticaseibacillus rhamnosus* e *Bifidobacterium animalis* subsp. *lactis*). Posteriormente, os extratos das tortas foram submetidos à digestão gastrointestinal *in vitro*, obtendo-se a fração bioacessível (BF) de cada um, as quais foram analisadas por LC-ESI-QTOF-MS/MS para identificação de moléculas bioativas. Os extratos não digeridos (NDE) e suas respectivas BF foram avaliados *in vitro* quanto ao teor de compostos fenólicos totais (TPC) e capacidade de sequestro dos radicais peroxila, ânion superóxido e ácido hipocloroso. A atividade anti-inflamatória dos NDE e BF também foi determinada por meio da inibição da ativação do NF-κB e TNF-α. Os NDE e BF de açaí e inajá ainda foram avaliados quanto à toxicidade aguda em *Galleria mellonella*. Na composição centesimal, o teor de fibras alimentares foi de $80,11 \pm 2,55$ e $62,16 \pm 3,39\%$ para as tortas de açaí e inajá, respectivamente). Ambos os subprodutos apresentaram um perfil mineral diverso e, de modo geral, com maiores teores em comparação às polpas dos respectivos frutos. Mg, K e S estão entre os elementos presentes nas maiores concentrações. Quanto ao potencial prebiótico, ambas as tortas aumentaram a viabilidade das bactérias após 24 h de fermentação em meios de cultura contendo os subprodutos, com destaque para o resultado da torta de inajá para *L. rhamnosus* ($8,3 \pm 0,1$ log CFU/mL). Ácidos fenólicos, isoorientina e vitexina foram identificados na BF de açaí, enquanto os ácidos caféico hexosídeo e cítrico, glabridina e eriodictiol 7-O-glicosídeo foram identificados na BF de inajá, evidenciando que esses compostos possam ser responsáveis pelas atividades biológicas determinadas. Para o TPC, não foi observada diferença estatística entre o NDE e a BF de um mesmo subproduto, porém houve diferença significativa NDE e BF de subprodutos diferentes. A capacidade de sequestro do radical peroxila do NDE de açaí diminuiu cerca de 2,3 vezes após a digestão, enquanto que para o inajá não houve diferença estatística, sendo a BF de inajá 1,42 vezes mais eficiente em comparação à BF de açaí. Os NDE e as BF dos subprodutos ainda demonstraram capacidade de sequestro do ácido hipocloroso. Para a atividade anti-inflamatória, macrófagos tratados com 100 µg/mL de NDE ou BF de açaí reduziram a ativação de NF-κB em 80 e 85%, contra 33 e 96% de NDE e BF de inajá, respectivamente. Os níveis de TNF-α foram reduzidos a 70 e 81% para NDE e BF de açaí, respectivamente, e em 93% para BF de inajá. Finalmente, a avaliação de toxicidade *in vivo* indicou toxicidade nas larvas de *G. mellonella* tratadas com NDE de açaí (0,1 g/kg) e de inajá (4 g/kg). Entretanto, as BF de ambos os subprodutos não apresentaram toxicidade. Portanto, os resultados deste trabalho indicam que as tortas de açaí e inajá possuem potencial para reaproveitamento industrial como possíveis fontes de fibras e minerais com potencial atividade prebiótica; bem como para matérias-primas para a extração de compostos naturais com ação antioxidant e anti-inflamatória.

Palavras-chave: Subprodutos de frutas; Compostos fenólicos; Digestão gastrointestinal *in vitro*; LC-MS/MS

ABSTRACT

Agro-industrial residues from Amazonian fruits: bioactive composition, bioaccessibility and potential use as a prebiotic

The intense global generation of waste and by-products resulting from agro-industrial activities indicates the need for studies to reuse these materials. This work aimed to determine the nutritional and bioactive composition, and prebiotic potential of by-products (meals) from the processing of açaí (*Euterpe oleracea* Mart) and inajá (*Maximiliana maripa* Aubl Drude) as natural sources of nutrients and compounds with antioxidant and anti-inflammatory activities. Açaí and inajá meals were prevent from the processing of the fruits for oil extraction. Initially, the proximate composition and the mineral profile of the raw materials were determined, followed by the assessment of the prebiotic potential using probiotic bacteria (*Lacticaseibacillus rhamnosus* and *Bifidobacterium animalis* subsp. *lactis*). Subsequently, the extracts of the meals were subjected to *in vitro* gastrointestinal digestion, thus obtaining the bioaccessible fraction (BF) of each one, which was analyzed by LC-ESI-QTOF-MS/MS for identifying bioactive molecules. The non-digested extracts (NDE) and their respective BF were evaluated *in vitro* for the total phenolic content (TPC) and scavenging capacity against the radicals peroxy, superoxide anion, and hypochlorous acid. The anti-inflammatory activity of NDE and BF was determined by the inhibition of NF-κB activation and TNF-α. The NDE and BF of açaí and inajá were also evaluated for acute toxicity in *Galleria mellonella*. In the proximate composition, the dietary fiber content was 80.11 ± 2.55 and $62.16 \pm 3.39\%$ for açaí and inajá, respectively). Both by-products showed a diversified mineral profile and, in general, with higher levels compared to the pulps of the respective fruits. Mg, K and S are among the elements present in higher concentrations. For the prebiotic potential, both meals increased the viability of the bacteria after 24 hours of fermentation in culture media containing the by-products, with emphasis on the result of the inajá meal for *L. rhamnosus* (8.3 ± 0.1 log CFU/mL). Phenolic acids, isoorientin, and vitexin, were identified in açaí BF, while caffeic acid hexoside, citric acid, glabridin, and eriodictyol 7-O-glucoside were identified in inajá BF, attesting that these compounds may be responsible for the assessed biological activities. For TPC, no statistical difference was observed between NDE and BF of the same by-product, but a significant difference was observed between NDE and BF of different by-products. The peroxy radical scavenging capacity of açaí NDE decreased about 2.3 times after digestion, while for inajá there was no statistical difference, with inajá BF being 1.42 times more efficient compared to açaí BF. The NDE and BF of the by-products also were capable of scavenging hypochlorous acid. For anti-inflammatory activity, macrophages treated with 100 µg/mL of NDE or BF from açaí reduced NF-κB activation by 80 and 85%, against 33 and 96% of NDE and BF from inajá, respectively. TNF-α levels were reduced by 70 and 81% for açaí NDE and BF, respectively, and by 93% for inajá BF. Finally, the evaluation of *in vivo* toxicity indicated toxic effects in *G. mellonella* larvae treated with NDE of açaí (0.1 g/kg) and inajá (4 g/kg). However, the BF of both by-products did not present toxicity. Therefore, the results of this work indicate that açaí and inajá meals have the potential for industrial reuse as possible sources of fibers and minerals with potential prebiotic activity; as well as raw materials for the extraction of natural compounds with antioxidant and anti-inflammatory action.

Keywords: Fruits by-products; Phenolic compounds; *In vitro* gastrointestinal digestion; LC-MS/MS

1. INTRODUCTION

The agribusiness activities represent important players in the Brazilian gross national product (GNP), and contributed with 26.6% (CNA, 2021), 27.5% and 25.5% in the years 2020, 2021 and 2022, respectively (CNA, 2022). Fruits in general are agricultural products with great importance in Brazilian exportations, which achieved 405.6 thousand tons and summed US\$ 381.2 million in 2022 (BRASIL, 2022). Brazil has a rich plant biodiversity representing 18% of the world (Biazotto et al., 2019). Regarding the fruitful species, 220 of the 500 varieties are native to the Amazonian region and represent 44% of the total diversity of fruits in Brazil (Neves et al., 2015).

However, agro-industrial activities also generate large amounts of solid residues or by-products which have become an issue of increasing concern worldwide due to the hard management mainly with respect to their disposal (Fierascul et al., 2020). Fruit processing, for example, can generate up to 65-80% of by-products by weight of the raw material (Melo et al., 2021, Santo et al., 2012), more than 50% of the fresh fruit (Torres-Leon et al., 2018), around 60% of harvested plants (Kodagoda and Marapana, 2017), and up to 35% of the raw mass in the form of pomace (Majerska et al., 2019). Peels, seeds, pomace, husks, stems, brans, oilseed cakes, and meals are examples of the diverse by-products generated in the processing of fruits (Carrillo et al., 2022). Most of these materials is discarded as residues in the environment, thus causing contamination and pollution of water and soils. Moreover, this low sustainable process also promotes logistic and transportation problems, thus leading to environmental and economic losses (Yi et al., 2009). Therefore, fruit by-products have been investigated for the recovery of components and for potential industrial applications (Carrillo et al., 2022), such as phenolic compounds.

Phenolic compounds are secondary metabolites of plant species with one or more hydroxyl groups attached to the aromatic rings that are broadly distributed in nature and, therefore, significantly present in the human diet (de Camargo and da Silva Lima, 2019, Heleno et al., 2015). These substances are produced by the secondary metabolism of plants throughout their development and in response to stress conditions and show a range of biological properties, such as antioxidant, anti-inflammatory, antimicrobial and cardioprotective effects, to name a few (Shahidi et al., 2019). Phenolic compounds can be found either in glycosylated or aglycone forms and to date, more than 8,000 polyphenols have been investigated in detail (Ramos, 2007, De Camargo et al., 2018).

Phenolic compounds can be grouped into flavonoids and non-flavonoids, based on the structure of the aglycone forms. The flavonoids comprise a wide range of compounds, classified as flavonols, flavones, isoflavones, flavanones, anthocyanidins, and flavan-3-ols, while non-flavonoids are classified as phenolic acids, hydroxycinnamates, hydrolyzable tannins and stilbenes. The structure of the phenolic compounds can influence their biological activities and bioavailability (Eseberri et al., 2022), which can be defined as the portion of the bioaccessible compound that is absorbed and reaches the systemic circulation, thus being distributed and metabolized in the organism (Reboreda-Rodriguez et al., 2021, Velderrain-Rodriguez et al., 2014). Besides the chemical structure, the bioavailability of a phenolic compound is highly dependent on its bioaccessibility, which can be defined as the amount of a phenolic compound that is released from the matrix

after the digestive process, reaching the gastrointestinal system, and that becomes available for absorption by the intestinal epithelial cells (Zahid et al., 2022, Arfaoui, 2021, Wojtunik-Kulesza et al., 2020).

The activity of phenolic compounds in scavenging reactive oxygen species (ROS), such as peroxyl radical, superoxide anion, and hypochlorous acid, in chelating metals and in reducing free radicals lead to the inhibition and/or mitigation of oxidative and inflammatory processes which are associated to several diseases (Circu and Aw, 2010, Wedick et al., 2012, Martinez et al., 2018, Romao et al., 2020, Li et al., 2020). Therefore, the ingestion of bioactive compounds from the diet is also necessary and of great relevance, because it consists of additional protection for maintaining the balance of the redox state (Pandey and Rizvi, 2009). Moreover, phenolic compounds can be industrially used as antioxidant agents, acting in food preservation, as well as in pharmacological and cosmetic products, acting as stabilizers, and inhibitors of lipid peroxidation (Luther et al., 2007, Melo et al., 2016). Brazilian native fruits are reported to present interesting antioxidant and anti-inflammatory properties mainly related to the presence of phenolic compounds (Infante et al., 2016, Lazarini et al., 2018, Lazarini et al., 2020). However, there are few studies investigating the biological activities of residues generated from Brazilian fruit processing for pulp and/or oil extraction.

Açaí (*Euterpe oleracea* Mart.), also known as açaí-do-Pará, and inajá (*Maximiliana maripa* Aubl. Drude) are species belonging to *Arecaceae* family found mainly in Pará, Amazonas, Acre, and Rondônia states, that are generally processed for obtaining pulp and oil, mainly for local and regional consumption. Nevertheless, the national and international demand for oleaginous fruits is growing due to the possibility of their use as ingredients, both in food products and in cosmetics and pharmaceuticals (Pacheco-Palencia et al., 2008, Rufino et al., 2011).

Açaí is a tropical fruit very popular in Brazil and worldwide, which has been reported as a superfruit due to its nutritional composition and bioactive compounds with valuable properties, such as antioxidant and anti-inflammatory (Kang et al., 2011, Yamaguchi et al., 2015). Açaí fruits are small and berry-like, clustered into bunches (Rufino et al., 2011) with higher fructification between July and September (Shanley et al., 2005). The market of this fruit shows a growing demand which is superior to the offer of the fruit, both locally in the producing regions, as national (Brazilian market), or internationally, which has impacted the prices and increased the exportation of this fruit (CONAB, 2022). Brazil is the biggest exporter of frozen açaí pulp, and Pará state is highlighted as the major producer of açaí in Brazil. In 2021, the Brazilian production of açaí reached nearly 1.5 million tons (Statista, 2022b, IBGE, 2021), with Pará state leading and representing 93.5% of the total amount of the fruits produced (Statista, 2022a). The main product extracted from the fruits is the pulp while the seeds, removed for pulp or oil extraction, represent 85-95% of the total volume of the fruit (Pompeu et al., 2009). For oil extraction, açaí fruits are generally macerated in hot water for removing the seeds. Afterward, the remaining fraction is dried and mechanically pressed for extracting the oil (de Souza Silva et al., 2023), thus yielding 50% of the oil depending on the processing conditions (Yamaguchi et al., 2015), and large amounts of a fiber-rich by-product are generated at the end of the process, named meal, which has been recently investigated, for the first time, for the phenolic composition as well as for antioxidant, anti-inflammatory and antimicrobial activities (de Souza Silva et al., 2021, de Souza Silva et al., 2023).

Inajá (*Maximiliana maripa* Aubl. Drude) is the fruit of inajazeiro palm tree, which fructifies between January and March in Pará state, and at the beginning of November in Acre state. Inajá pulp has around 23% (Shanley et al., 2012) to 35.50% oil, showing a potential to be used as a raw material in the cosmetic, soap, and food industries (Bezerra, 2011). For mechanical oil extraction, inajá fruits are vapor heated and mashed for removing the pulp. Then, the pulp is heated up to 90 °C and pressed for extracting the oil (Shanley et al., 2012), when large amounts of inajá meal are generated since the yield in oil is low. This fruit still has not an added value or a large market such as açaí, being mostly consumed in the producing regions. However, this species has received attention and becoming the target of research regarding its use as a biofuel (Stachiw et al., 2016), which can even benefit the communities in which the palm tree is present, with the generation of employment, incomes, and energy for isolated areas (EMBRAPA, 2014).

Since plant products are natural sources of phenolic compounds, new investigations have been developed with plant processing by-products aiming to discover new alternative sources of bioactive substances from these materials generated in large amounts (Melo et al., 2021, Melo et al., 2020, de Camargo et al., 2016, Oldoni et al., 2016, Melo et al., 2015). However, more studies of the properties of residues generated in food processing, such as meals, are needed considering the bioactive potential of these materials, as well as their great potential to be incorporated as ingredients in several applications, such as cosmetics, pharmaceutical, with a perspective for incorporation in the human feed once they present high nutritional value, mainly in minerals and fibers (Becker et al., 2018).

Agro-industrial by-products may generate fiber-rich ingredients with functional and technological properties (Selani et al., 2014). Besides nourishing by providing proteins, lipids, carbohydrates, and other non-nutritional compounds, such as antioxidants (Torres-Leon et al., 2018), the composition of açaí and inajá meals in macro and micronutrients can also indicate potential prebiotic effects, thus benefiting the development of probiotic bacteria and increase their population in the colon. Probiotics can be defined as live micro-organisms that confer health benefits to the host when administered in adequate amounts (Hill et al., 2014), while prebiotics can be defined as a substrate that is selectively utilized by host microorganisms conferring health benefits (Gibson et al., 2017).

Açaí and inajá meals are still underexplored by the scientific community. Recently, both by-products were primarily studied for an initial characterization of biological activities and phenolic profile of their respective hydroethanolic extracts (de Souza Silva et al., 2021, de Souza Silva et al., 2023). However, to the best of our knowledge, no studies have been performed for assessing the proximate composition and mineral profile of these materials to date, which can be related to a prebiotic potential. Also, the behavior of the *in vitro* activities, as well as the profile in bioactive compounds of the extracts subjected to *in vitro* gastrointestinal digestion, were unclear and non-investigated to date.

Therefore, the objective of this investigation was to characterize the by-products generated in the processing of açaí and inajá fruits to act as potential sources of dietary fibers and minerals, also investigating for the first time the prebiotic potential of those by-products on the counting of probiotic bacteria strains; and to assess the phenolic profile, antioxidant and anti-inflammatory capacities and *in vivo* toxic effects of the bioaccessible fractions of açaí and inajá meals.

For this, this work was divided into four work packages, which originated the two scientific manuscripts of this thesis (starting on page 27 and on page 46, respectively). The first manuscript assesses the proximate composition, mineral profile, and the prebiotic potential of açaí and inajá meals is presented. The second comprises the study investigating the phenolic profile of açaí and inajá meals after *in vitro* simulated gastrointestinal digestion, the antioxidant and anti-inflammatory activities of both non-digested and digested extracts, as well as the investigation for acute systemic toxicity of the two forms of the extracts using an *in vivo* model of *Galleria mellonella*. Hence, our findings contribute to presenting new possibilities for the use and valorization of agro-industrial by-products generated from Brazilian native fruits.

2. CONCLUSION

This study demonstrated that both by-products from the processing of açaí and inajá meals for oil extraction are rich sources of total dietary fibers, mainly the insoluble ones. Additionally, the meals presented a diverse profile of macro and microminerals, which are of great importance for the human diet, and that are present in higher concentrations compared to the pulps of the respective fruits. The prebiotic potential of both by-products was indicated by the increase in the viability of probiotic bacteria strains (*L. rhamnosus* and *B. animalis* subsp. *Lactis*) after 24 h of incubation. This finding may be mainly related to the total dietary fiber content of both by-products. Moreover, the samples did not exert antimicrobial effects at the established conditions. Therefore, açaí and inajá meals could be considered new natural sources of dietary fibers and minerals to be used as new food ingredients, with potential prebiotic activity. This study investigated the raw materials and opens new opportunities for further studies to explore the same parameters for the extracts produced from the by-products, also considering other probiotic strains grown individually or in co-cultures. Moreover, studies assessing the influence of the fermentation of the by-products by probiotic strains on the proximate composition, as well as determining the produced metabolites, are encouraged. Furthermore, the evaluation of different lots from the same and different regions would be important for developing an initial characterization standard of identity and quality of these materials for each region, which nutritional composition could be related to the response of the probiotic micro-organisms.

The results of this study indicate that antioxidant and anti-inflammatory activities are to some extent preserved after gastrointestinal digestion. Even gastrointestinal digestion affects the antioxidant capacity of the digested fractions somehow, the results are higher when compared to extracts obtained from other natural products. Furthermore, the bioaccessible fractions of both meals showed an important reduction of inflammation biomarkers by inhibiting the activation of NF-κB and decreasing TNF-α levels. This suggests that the remaining phenolic compounds in BF of both meals may still have an important role in the modulation of inflammatory processes when released from the matrix. Also, the bioaccessible fractions of each sample did not promote *in vivo* acute systemic toxicity, thus being safe at the same time while still conserve biological activities. Therefore, açaí and inajá meals can be considered important sources of natural bioactive compounds that may reach the small intestine for the development of new functional foods, or to be used as bioactive ingredients in food, cosmetic or pharmaceutical industries. Future studies may be developed employing dynamic digestion models for better simulating *in vivo* digestion process together the co-culture of cells for evaluating the permeability and the uptake of phenolic compounds, so that their bioavailability could be estimated closer to real metabolism conditions, also analyzing the effects of this new approach in the antioxidant, anti-inflammatory and toxicological responses.

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3. REFERENCES

- Abdel-Salam, O. M., Youness, E. R., Mohammed, N. A., Morsy, S. M., Omara, E. A., & Sleem, A. A. (2014). Citric acid effects on brain and liver oxidative stress in lipopolysaccharide-treated mice. *Journal of Medicinal Food*, 17(5), 588-598. <https://doi.org/10.1089/jmf.2013.0065>.
- Abdullah, Z., Hussain, K., Ismail, Z., & Ali, R. M. (2009). Anti-inflammatory activity of standardised extracts of leaves of three varieties of *Ficus deltoidea*. *International Journal of Pharmaceutical and Clinical Research*, 1(3), 100-105.
- Afifi, F. U., & Abu-Dahab, R. (2012). Phytochemical screening and biological activities of *Eminium spiculatum* (Blume) Kuntze (family Araceae). *Nat Prod Res*, 26(9), 878-882. <https://doi.org/10.1080/14786419.2011.565558>.
- Albuquerque, M.A.C., Bedani, R., Vieira, A.D.S., Leblanc, J.G., Saad S.M.I. (2016). Supplementation with fruit and okara soybean by-products and amaranth flour increases the folate production by starter and probiotic cultures. *International Journal of Food Microbiology*, 236, 26-31. <https://doi.org/10.1016/j.ijfoodmicro.2016.07.008>.
- Albuquerque, M.A.C., Yamacita, D.S., Bedani, R., Leblanc, J.G., Saad, S.M.I. (2019). Influence of passion fruit by-product and fructooligosaccharides on the viability of *Streptococcus thermophilus* TH-4 and *Lactobacillus rhamnosus* LGG in folate bio-enriched fermented soy products and their effect on probiotic survival and folate bio-accessibility under *in vitro* simulated gastrointestinal conditions. *International Journal of Food Microbiology*, 292, 126-136. <https://doi.org/10.1016/j.ijfoodmicro.2018.12.012>.
- Almeida, J.S.O., Dias, C.O., Arriola, N.D.A., Freitas, B.S.M., Francisco, A., Petkowicz, C.L.O., Araujo, L., Guerra, M.P., Nodari, R.O., Amboni, R.D.M.C. (2020). Feijoa (*Acca sellowiana*) peel flours: A source of dietary fibers and bioactive compounds. *Food Bioscience*, 38. <https://doi.org/10.1016/j.fbio.2020.100789>.
- Alqurashi R.M., Alarifi, S.N., Walton, G.E., Costabile, A.F., Rowland, I.R., Commane, D.M. (2017). In vitro approaches to assess the effects of açaí (*Euterpe oleracea*) digestion on polyphenol availability and the subsequent impact on the faecal microbiota. *Food Chemistry*, 234, 190-198. <https://doi.org/10.1016/j.foodchem.2017.04.164>.
- Alves, V.M., Asquieri, E.R., Araújo, E.S., Martins, G.A.S., Melo, A.A.M., Freitas, B.C.B., Damiani, C. (2022). Provenient residues from industrial processing of açaí berries (*Euterpe precatoria* Mart): nutritional and antinutritional contents, phenolic profile, and pigments. *Food Science and Technology*, 42. <https://doi.org/10.1590/fst.77521>.
- Alves-Santos, A.M., Sampaio, K.B., Lima, M.S., Coelho, A.S.G., Souza, E.L., Naves, M.M.V. (2022). Chemical composition and prebiotic activity of baru (*Dipteryx alata* Vog.) pulp on probiotic strains and human colonic microbiota. *Food Research International* 112366, ISSN 0963-9969. <https://doi.org/10.1016/j.foodres.2022.112366>.
- Anunciacao, P. C., Giuffrida, D., Murador, D. C., de Paula, G. X., Dugo, G., & Pinheiro-Sant'Ana, H. M. (2019). Identification and quantification of the native carotenoid composition in fruits from the Brazilian Amazon by HPLC-DAD-APCI/MS. *Journal of Food Composition and Analysis*, 83. <https://doi.org/10.1016/j.jfca.2019.103296>.
- Aoki, F., Nakagawa, K., Tanaka, A., Matsuzaki, K., Arai, N., & Mae, T. (2005). Determination of glabridin in human plasma by solid-phase extraction and LC-MS/MS. *J Chromatogr B Analyt Technol Biomed Life Sci*, 828(1-2), 70-74. <https://doi.org/10.1016/j.jchromb.2005.09.012>.
- Arfaoui, L. (2021). Dietary Plant Polyphenols: Effects of Food Processing on Their Content and Bioavailability. *Molecules*, 26(10). <https://doi.org/10.3390/molecules26102959>.

- Arruda, J.C.B., Da Fonseca, L.A.B., Pinto, L.C.P., Pinheiro, H.C.O., Monteiro, B.T.O., Manno, M.C., Lima, K.R.S., De Lima, A.R. (2018). Açaí seed bran in the feed of slow-growth broilers. *Acta Amazonica*, 48, 298–303. <https://doi.org/10.1590/1809-4392201703994>.
- Asp, N.G., Johansson, C.G. , Hallmer, H. (1983). Rapid enzymatic assay of insoluble and soluble dietary fiber. *Journal of Agricultural and Food Chemistry*, 31, 476-482. <https://doi.org/10.1021/jf00117a003>.
- Association Of Official Analytical Chemists – AOAC. (2006). Official methods of analysis of the Association of Official Analytical Chemists. 18th ed. Arlington. 1141p.
- Babbar, N., Oberoi, H. S., Uppal, D. S., & Patil, R. T. (2011). Total phenolic content and antioxidant capacity of extracts obtained from six important fruit residues. *Food Research International*, 44(1), 391-396. <https://doi.org/10.1016/j.foodres.2010.10.001>.
- Bamigbade, G.B., Subhash, A.J., Kamal-Eldin, A., Nyström, L., Ayyash, M. (2022). An Updated Review on Prebiotics: Insights on Potentials of Food Seeds Waste as Source of Potential Prebiotics. *Molecules*, 27. <https://doi.org/10.3390/molecules27185947>.
- Barbi, R. C. T., Hornung, P. S., Avila, S., Alves, F. E. D. B., Beta, T., & Ribani, R. H. (2020). Ripe and unripe inajá (*Maximilia maripa*) fruit: A new high source of added value bioactive compounds. *Food Chemistry*, 331. <https://doi.org/10.1016/j.foodchem.2020.127333>.
- Becker, M.M., Chagas, V.T., Marty, J.L., Mendes, T.M.F.F., Nunes, G.S. (2018). Chemical variability in Amazonian palm fruits: açaí (*Euterpe oleracea* Mart.), buriti (*Mauritia flexuosa* L. f.), and inajá [*Maximiliana maripa* (Aubl.) Drude] (Arecaceae). Boletim do Museu Paraense Emílio Goeldi, *Ciências Naturais*, 13, 67-77. <https://doi.org/10.46357/bcnaturais.v13i1.369>.
- Behera, B. C., Mishra, R., & Mohapatra, S. (2021). Microbial citric acid: Production, properties, application, and future perspectives. *Food Frontiers*, 2(1), 62-76.
- Beliek, V., Kolisek, M. (2021). Bioaccessibility and Bioavailability of Minerals in Relation to a Healthy Gut Microbiome. *International Journal of Molecular Sciences*, 22, 6803. <https://doi.org/10.3390/ijms22136803>.
- Ben Said, R., Hamed, A. I., Mahalel, U. A., Al-Ayed, A. S., Kowalczyk, M., Moldoch, J., . . . Stochmal, A. (2017). Tentative Characterization of Polyphenolic Compounds in the Male Flowers of *Phoenix dactylifera* by Liquid Chromatography Coupled with Mass Spectrometry and DFT. *International Journal of Molecular Sciences*, 18(3). <https://doi.org/10.3390/ijms18030512>.
- Bezerra, V. S. (2011). O Inajá (*Maximiliana maripa* (Aubl.) Drude) como fonte alimentar e oleaginosa. *Embrapa Amapá-Comunicado Técnico (INFOTECA-E)*. Available: <https://www.embrapa.br/en/busca-de-publicacoes/-/publicacao/917016/o-inaja-maximiliana-maripa-aubl-drude-como-fonte-alimentar-e-oleaginosa>. [Accessed 14/09/2022].
- Biazotto, K. R., Mesquita, L. M. D., Neves, B. V., Braga, A. R. C., Tangerina, M. M. P., Vilegas, W., . . . De Rosso, V. V. (2019). Brazilian Biodiversity Fruits: Discovering Bioactive Compounds from Underexplored Sources. *Journal of Agricultural and Food Chemistry*, 67(7), 1860-1876. <https://doi.org/10.1021/acs.jafc.8b05815>.
- Bojórquez-Quintal, E., Escalante-Magaña, C., Echevarría-Machado, I., Martínez-Estévez, M. (2017). Aluminum, a Friend or Foe of Higher Plants in Acid Soils. *Frontiers in Plant Science*, 8, 1767. <https://doi.org/10.3389/fpls.2017.01767>.
- Brodkorb, A., Egger, L., Alminger, M., Alvito, P., Assuncao, R., Ballance, S., . . . Recio, I. (2019). INFOGEST static *in vitro* simulation of gastrointestinal food digestion. *Nature Protocols*, 14(4), 991-1014. <https://doi.org/10.1038/s41596-018-0119-1>.

- Brunschwig, C., Leba, L. J., Saout, M., Martial, K., Bereau, D., & Robinson, J. C. (2017). Chemical Composition and Antioxidant Activity of *Euterpe oleracea* Roots and Leaflets. *International Journal of Molecular Sciences*, 18(1). <https://doi.org/10.3390/ijms18010061>.
- Campos , D.A., Gómez-García, R., Villas-Boas, A.A., Madureira, A.R., Pintado, M.M. (2020). Management of Fruit Industrial By-Products—A Case Study on Circular Economy Approach. *Molecules*, 25. <https://doi.org/10.3390/molecules25020320>.
- Carrillo, C., Nieto, G., Martínez-Zamora, L., Ros, G., Kamiloglu, S., Munekata, P.E.S., Pateir, M., Lorenzo, J.M., Fernandez-López, J., Viuda-Martos, M., Pérez-Álvarez, J.A., Barba, F.J. (2022). Novel Approaches for the Recovery of Natural Pigments with Potential Health Effects. *Journal of Agricultural and Food Chemistry*, 70, 6864-6883. <https://doi.org/10.1021/acs.jafc.1c07208>.
- Cavalcante, P.B. (2010). Frutas comestíveis na Amazônia. 7 ed. Belém: CNPq/Museu Paraense Emílio Goeldi, 282 p.
- Chae, S.A., Ramakrishnan, S.R., Kim, T., Kim, S.R., Bang, W.Y., Jeong, C.R., Yang, J., Kim, S.J. (2022). Anti-inflammatory and anti-pathogenic potential of *Lacticaseibacillus rhamnosus* IDCC 3201 isolated from feces of breast-fed infants. *Microbial Pathogenesis*, 173, 105857. <https://doi.org/10.1016/j.micpath.2022.105857>.
- Chaplin, A., Parra, P., Laraichi, S., Serra, F., Palou, A. (2016). Calcium supplementation modulates gut microbiota in a prebiotic manner in dietary obese mice. *Molecular Nutrition & Food Research*, 60, 468-480. <https://doi.org.ez67.periodicos.capes.gov.br/10.1002/mnfr.201500480>.
- Chiste, R. C., Freitas, M., Mercadante, A. Z., & Fernandes, E. (2015). Superoxide Anion Radical: Generation and Detection in Cellular and Non-Cellular Systems. *Curr Med Chem*, 22(37), 4234-4256. <https://doi.org/10.2174/092986732266151029104311>.
- Cory, H., Passarelli, S., Szeto, J., Tamez, M., Mattei, J. (2018). The Role of Polyphenols in Human Health and Food Systems: A Mini-Review. *Frontiers in Nutrition*, 5, 87. <https://doi.org/10.3389/fnut.2018.00087>.
- Dai, C. H., Ma, H. L., He, R. H., Huang, L. R., Zhu, S. Y., Ding, Q. Z., & Luo, L. (2017). Improvement of nutritional value and bioactivity of soybean meal by solid-state fermentation with *Bacillus subtilis*. *Lwt-Food Science and Technology*, 86, 1-7. <https://doi.org/10.1016/j.lwt.2017.07.041>.
- D'Archivio, M., Filesi, C., Vari, R., Scazzocchio, B., & Masella, R. (2010). Bioavailability of the polyphenols: status and controversies. *International Journal of Molecular Sciences*, 11(4), 1321-1342. <https://doi.org/10.3390/ijms11041321>.
- Davani-Davari, D., Negahdaripour, M., Karimzadeh, I., Seifan, M., Mohkam, M., Masoumi, S.J., Berenjian, A., Ghasemi, Y. (2019). Prebiotics: Definition, Types, Sources, Mechanisms, and Clinical Applications. *Foods*, 8. <https://doi.org/10.3390/foods8030092>.
- de Francisco, L., Pinto, D., Rosseto, H., Toledo, L., Santos, R., Tobaldini-Valerio, F., . . . Rodrigues, F. (2018). Evaluation of radical scavenging activity, intestinal cell viability and antifungal activity of Brazilian propolis by-product. *Food Research International*, 105, 537-547. <https://doi.org/10.1016/j.foodres.2017.11.046>.
- de Oliveira, P. D., da Silva, D. A., Pires, W. P., Bezerra, C. V., da Silva, L. H. M., & da Cruz Rodrigues, A. M. (2021). Enzymatic interesterification effect on the physicochemical and technological properties of cupuassu seed fat and inajá pulp oil blends. *Food Research International*, 145, 110384. <https://doi.org/10.1016/j.foodres.2021.110384>.
- de Souza Silva, A. P., de Camargo, A. C., Lazarini, J. G., Franchin, M., Sardi, J. d. C. O., Rosalen, P. L., & de Alencar, S. M. (2023). Phenolic Profile and the Antioxidant, Anti-Inflammatory, and Antimicrobial Properties of Açaí (*Euterpe oleracea*) Meal: A Prospective Study. *Foods*, 12(1), 86.
- de Souza Silva, A. P., Rosalen, P. L., de Camargo, A. C., Lazarini, J. G., Rocha, G., Shahidi, F., . . . de Alencar, S. M. (2021). Inajá oil processing by-product: A novel source of bioactive catechins and procyanidins from a

- Brazilian native fruit. *Food Research International*, 144. <https://doi.org/10.1016/j.foodres.2021.110353>.
- De Souza Silva, A.P., Rosalen, P.L., De Camargo, A.C., Lazarini, J.G., Rocha, G., Shahidi, F., Franchin, M., de Alencar, S.M. (2021). Inajá oil processing by-product: A novel source of bioactive catechins and procyanidins from a Brazilian native fruit. *Food Research International*, 144. <https://doi.org/10.1016/j.foodres.2021.110353>.
- Dhingra, D., Michael, M., Rajput, H., Patil, R.T. (2012). Dietary fibre in foods: a review. *Journal of Food Science and Technology*, 49, 255-266. <https://doi.org/10.1007/s13197-011-0365-5>.
- di Gesso, J. L., Kerr, J. S., Zhang, Q., Raheem, S., Yalamanchili, S. K., O'Hagan, D., . . . O'Connell, M. A. (2015). Flavonoid metabolites reduce tumor necrosis factor-alpha secretion to a greater extent than their precursor compounds in human THP-1 monocytes. *Mol Nutr Food Res*, 59(6), 1143-1154. <https://doi.org/10.1002/mnfr.201400799>.
- Di Lorenzo, C., Colombo, F., Biella, S., Stockley, C., & Restani, P. (2021). Polyphenols and Human Health: The Role of Bioavailability. *Nutrients*, 13(1). <https://doi.org/10.3390/nu13010273>.
- Dias, A. L. S., Rozet, E., Larondelle, Y., Hubert, P., Rogez, H., & Quetin-Leclercq, J. (2013). Development and validation of an UHPLC-LTQ-Orbitrap MS method for non-anthocyanin flavonoids quantification in *Euterpe oleracea* juice. *Analytical and Bioanalytical Chemistry*, 405(28), 9235-9249. <https://doi.org/10.1007/s00216-013-7325-z>.
- EMBRAPA. (2014). Prosa Rural - Palmeira inajá: matéria-prima para o biodiesel e fonte de renda para o pequeno produtor. Retrieved from: <https://www.embrapa.br/web/portal/busca-de-noticias/-/noticia/1806420/prosa-rural---palmeira-inaja-materia-prima-para-o-biodiesel-e-fonte-de-renda-para-o-pequeno-produtor> [Accessed 12/10/2021].
- Empresa Brasileira De Pesquisa Agropecuária – EMBRAPA (2014) Prosa Rural - Palmeira inajá: matéria-prima para o biodiesel e fonte de renda para o pequeno produtor. Available at:<<https://www.embrapa.br/web/portal/busca-de-noticias/-/noticia/1806420/prosa-rural---palmeira-inaja-materia-prima-para-o-biodiesel-e-fonte-de-renda-para-o-pequeno-produtor>>. Accessed on october 2022.
- Falcone, M., Marques, A.B. (1965). Estudo sobre as condições de hidrólise pelo ácido clorídrico na dosagem de açúcares redutores totais. *Tecnologia de Alimentos e Bebidas*, 4, 24-29.
- Farsi, E., Shafaei, A., Hor, S. Y., Ahamed, M. B. K., Yam, M. F., Attitalla, I. H., . . . Ismail, Z. (2011). Correlation between enzymes inhibitory effects and antioxidant activities of standardized fractions of methanolic extract obtained from *Ficus deltoidea* leaves. *African journal of Biotechnology*, 10(67), 15184-15194.
- Ferreira, D. S. (2016). Antioxidant capacity and chemical characterization of açaí (*Euterpe oleracea* Mart.) fruit fractions. *Food Science and Technology*, 4 (5), 95-102. <https://doi.org/10.13189/fst.2016.040502>.
- Ferreres, F., Silva, B. M., Andrade, P. B., Seabra, R. M., & Ferreira, M. A. (2003). Approach to the study of C-glycosyl flavones by ion trap HPLC-PAD-ESI/MS/MS: Application to seeds of quince (*Cydonia oblonga*). *Phytochemical Analysis*, 14(6), 352-359. <https://doi.org/10.1002/pca.727>.
- Fierascu, R.C., Sieniawska, E., Ortan, A., Fierascu, I., Xiao, J. (2020). Fruits By-Products – A Source of Valuable Active Principles. A Short Review. *Frontiers in Bioengineering and Biotechnology*, 8. <https://doi.org/10.3389/fbioe.2020.00319>.
- Fritsch, C., Heinrich, V., Vogel, R.F., Toelstede, S. (2021). Phenolic acid degradation potential and growth behavior of lactic acid bacteria in sunflower substrates. *Food Microbiology*, 57, 178–186. <https://doi.org/10.1016/j.fm.2016.03.003>.
- Garzon, G. A., Narvaez-Cuenca, C. E., Vincken, J. P., & Gruppen, H. (2017). Polyphenolic composition and antioxidant activity of açaí (*Euterpe oleracea* Mart.) from Colombia. *Food Chemistry*, 217, 364-372. <https://doi.org/10.1016/j.foodchem.2016.08.107>.
- Gibson, G., Hutkins, R., Sanders, M.E., Prescott, S.L., Reimer, R.A., Salminen, S.J., Scott, K., Stanton, C., Swanson, K.S., Cani, P.D., Verbeke, K., Reid, G. (2017). Expert consensus document: The International Scientific

- Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. *Nature Reviews Gastroenterology & Hepatology*, 14, 491-502. <https://doi.org/10.1038/nrgastro.2017.75>.
- Gordon, A., Cruz, A. P. G., Cabral, L. M. C., de Freitas, S. C., Taxi, C. M. A. D., Donangelo, C. M., . . . Marx, F. (2012). Chemical characterization and evaluation of antioxidant properties of Acai fruits (*Euterpe oleracea* Mart.) during ripening. *Food Chemistry*, 133(2), 256-263. <https://doi.org/10.1016/j.foodchem.2011.11.150>.
- Harris, G.K., Marshall, M.R. (2017). Ash Analysis. In: Nielsen, S.S. (eds) Food Analysis. Food Science Text Series. Springer, Cham. https://doi.org/10.1007/978-3-319-45776-5_16.
- He, C., Sampers, I., Raes, K. (2021). Dietary fiber concentrates recovered from agro-industrial by-products: Functional properties and application as physical carriers for probiotics. *Food Hydrocolloids*, 111. <https://doi.org/10.1016/j.foodhyd.2020.106175>.
- He, M., Min, J. W., Kong, W. L., He, X. H., Li, J. X., & Peng, B. W. (2016). A review on the pharmacological effects of vitexin and isovitexin. *Fitoterapia*, 115, 74-85. <https://doi.org/10.1016/j.fitote.2016.09.011>.
- Hill, C., Guarner, F., Reid, G., Gibson, G.R., Merenstein, D.J., Pot, B., Morelli, L., Canani, R.B., Flint, H.J., Salminen, S., Calder, P.C., Sanders, M.E. (2014), The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nature Reviews Gastroenterology & Hepatology*, 11, 506-514. <https://doi.org/10.1038/nrgastro.2014.66>.
- IBGE. The Brazilian Institute of Geography and Statistics (2021) Production of Plant Extraction and Forestry. Vegetable extraction: Table 1 – Amount produced and the production value of Brazil, great regions and federal units, according to the extractive products. Available at: <<https://www.ibge.gov.br/estatisticas/economicas/agricultura-e-pecaaria/9105-producao-da-extracao-vegetal-e-da-silvicultura.html?=&t=resultados>>. Accessed on october 2022.
- Jing, X., Ren, D. M., Wei, X. B., Shi, H. Y., Zhang, X. M., Perez, R. G., . . . Lou, H. X. (2013). Eriodictyol-7-O-glucoside activates Nrf2 and protects against cerebral ischemic injury. *Toxicology and Applied Pharmacology*, 273(3), 672-679. <https://doi.org/10.1016/j.taap.2013.10.018>.
- Jones, J.M. (2013). Dietary Fiber Future Directions: Integrating New Definitions and Findings to Inform Nutrition Research and Communication. *Advances in Nutrition*, 4, 8-15. <https://doi.org/10.3945/an.112.002907>.
- Jones, J.M. (2014). CODEX-aligned dietary fiber definitions help to bridge the 'fiber gap'. *Nutrition Journal*, 13. <https://doi.org/10.1186/1475-2891-13-34>.
- Jørgensen, B.P., Winther, G., Kihl, P., Nielsen, D.S., Wegener, G., Hansen, A.K., Sørensen, D.B. (2015). Dietary magnesium deficiency affects gut microbiota and anxiety-like behaviour in C57BL/6N mice. *Acta Neuropsychiatrica*, 27, 307-311. <https://doi.org/10.1017/neu.2015.10>.
- Jorjao, A. L., Oliveira, L. D., Scorzoni, L., Figueiredo-Godoi, L. M. A., Prata, M. C. A., Jorge, A. O. C., & Junqueira, J. C. (2018). From moths to caterpillars: Ideal conditions for *Galleria mellonella* rearing for *in vivo* microbiological studies. *Virulence*, 9(1), 383-389. <https://doi.org/10.1080/21505594.2017.1397871>.
- Júnior, A.F.S., Matos, R.A., Andrade, E.M.J., Santos, W.N.L., Magalhães, H.I.F., Costa, F.N., Korn, M.G. (2017). Multielement Determination of Macro and Micro Contents in Medicinal Plants and Phyto-medicines from Brazil by ICP OES. *Journal of the Brazilian Chemical Society*, 28, 376-384. <https://doi.org/10.5935/0103-5053.20160187>.
- Kang, J., Xie, C. H., Li, Z. M., Nagarajan, S., Schauss, A. G., Wu, T., & Wu, X. L. (2011). Flavonoids from acai (*Euterpe oleracea* Mart.) pulp and their antioxidant and anti-inflammatory activities. *Food Chemistry*, 128(1), 152-157. <https://doi.org/10.1016/j.foodchem.2011.03.011>.

- Khan, J., Deb, P.K., Priya, S., Medina, K.D., Devi, R., Walode, S.G., Rudrapal, M. (2021). Dietary Flavonoids: Cardioprotective Potential with Antioxidant Effects and Their Pharmacokinetic, Toxicological and Therapeutic Concerns. *Molecules*, 26. <https://doi.org/10.3390/molecules26134021>.
- Kodagoda, K.H.G.K., Marapana, R.A.U.J. (2017). Utilization of fruit processing by-products for industrial applications: A review. *International Journal of Food Science and Nutrition*, 2, 24-30.
- Kramberger, K., Barlic-Maganja, D., Bandelj, D., Baruca Arbeiter, A., Peeters, K., Miklavcic Visnjevec, A., & Jenko Praznikar, Z. (2020). HPLC-DAD-ESI-QTOF-MS Determination of Bioactive Compounds and Antioxidant Activity Comparison of the Hydroalcoholic and Water Extracts from Two Helichrysum italicum Species. *Metabolites*, 10(10). <https://doi.org/10.3390/metabo10100403>.
- Kumar, K., Yadav, A.N., Kumar, V., Vyas, P., Dhaliwal, H.S. (2017). Food waste: a potential bioresource for extraction of nutraceuticals and bioactive compounds. *Bioresources and Bioprocessing*, 4. <https://doi.org/10.1186/s40643-017-0148-6>.
- Lazarini, J. G., Sardi, J. D. O., Franchin, M., Nani, B. D., Freires, I. A., Infante, J., . . . Rosalen, P. L. (2018). Bioprospection of Eugenia brasiliensis, a Brazilian native fruit, as a source of anti-inflammatory and antibiofilm compounds. *Biomedicine & Pharmacotherapy*, 102, 132-139. <https://doi.org/10.1016/j.biopha.2018.03.034>.
- Lende, A. B., Kshirsagar, A. D., Deshpande, A. D., Muley, M. M., Patil, R. R., Bafna, P. A., & Naik, S. R. (2011). Anti-inflammatory and analgesic activity of protocatechuic acid in rats and mice. *Inflammopharmacology*, 19(5), 255-263. <https://doi.org/10.1007/s10787-011-0086-4>.
- Li, D., Rui, Y. X., Guo, S. D., Luan, F., Liu, R., & Zeng, N. (2021). Ferulic acid: A review of its pharmacology, pharmacokinetics and derivatives. *Life Sciences*, 284. <https://doi.org/10.1016/j.lfs.2021.119921>.
- Li, Z. H., Guo, H., Xu, W. B., Ge, J., Li, X., Alimu, M., & He, D. J. (2016). Rapid Identification of Flavonoid Constituents Directly from PTP1B Inhibitive Extract of Raspberry (*Rubus idaeus* L.) Leaves by HPLC-ESI-QTOF-MS-MS. *Journal of Chromatographic Science*, 54(5), 805-810. <https://doi.org/10.1093/chromsci/bmw016>.
- Lim, C.C., Ferguson, L.R., Tannock, G.W. (2005). Dietary fibres as “prebiotics”: Implications for colorectal cancer. *Molecular Nutrition & Food Research*, 49, 609-619. <https://doi.org/10.1002/mnfr.200500015>.
- Lim, J. S., Yang, J. H., Chun, B. Y., Kam, S., Jacobs, D. R., Jr., & Lee, D. H. (2004). Is serum gamma-glutamyltransferase inversely associated with serum antioxidants as a marker of oxidative stress? *Free Radic Biol Med*, 37(7), 1018-1023. <https://doi.org/10.1016/j.freeradbiomed.2004.06.032>.
- Liu, C. L., Wang, J. M., Chu, C. Y., Cheng, M. T., & Tseng, T. H. (2002). *In vivo* protective effect of protocatechuic acid on tert-butyl hydroperoxide-induced rat hepatotoxicity. *Food and Chemical Toxicology*, 40(5), 635-641. [https://doi.org/10.1016/s0278-6915\(02\)00002-9](https://doi.org/10.1016/s0278-6915(02)00002-9).
- Long, G.L., Winefordner, J.D. (1983). Limit of detection: a closer look at IUPAC definition. *Analytical Chemistry*, 55, 712-724. <https://doi.org/10.1021/ac00258a724>.
- Lu, C. C., Xu, Y. Q., Wu, J. C., Hang, P. Z., Wang, Y., Wang, C., . . . Du, Z. M. (2013). Vitexin protects against cardiac hypertrophy via inhibiting calcineurin and CaMKII signaling pathways. *Naunyn Schmiedebergs Arch Pharmacol*, 386(8), 747-755. <https://doi.org/10.1007/s00210-013-0873-0>.
- Macfarlane, S., Macfarlane, G.T., Cummings, J.H. (2005). Review article: prebiotics in the gastrointestinal tract. *Alimentary Pharmacology & Therapeutics*, 24, 701-714. <https://doi.org/10.1111/j.1365-2036.2006.03042.x>.
- Majerska, J., Michalska, A., Figiel, A. (2019). A review of new directions in managing fruit and vegetable processing by-products. *Trends in Food Science & Technology*, 88, 207-219. <https://doi.org/10.1016/j.tifs.2019.03.021>.
- Martins, G. R., do Amaral, F. R. L., Brum, F. L., Mohana-Borges, R., de Moura, S. S. T., Ferreira, F. A., . . . da Silva, A. S. (2020). Chemical characterization, antioxidant and antimicrobial activities of acai seed (*Euterpe*

- oleracea Mart.) extracts containing A- and B-type procyanidins. Lwt-Food Science and Technology, 132.*
<https://doi.org/10.1016/j.lwt.2020.109830>.
- Melo, P. S., Arrivetti, L. D. R., de Alencar, S. M., & Skibsted, L. H. (2016). Antioxidative and prooxidative effects in food lipids and synergism with alpha-tocopherol of acai seed extracts and grape rachis extracts. *Food Chemistry, 213*, 440-449. <https://doi.org/10.1016/j.foodchem.2016.06.101>.
- Melo, P. S., Massarioli, A. P., Denny, C., dos Santos, L. F., Franchin, M., Pereira, G. E., . . . de Alencar, S. M. (2015). Winery by-products: Extraction optimization, phenolic composition and cytotoxic evaluation to act as a new source of scavenging of reactive oxygen species. *Food Chemistry, 181*, 160-169. <https://doi.org/10.1016/j.foodchem.2015.02.087>.
- Melo, P. S., Massarioli, A. P., Lazarini, J. G., Soares, J. C., Franchin, M., Rosalen, P. L., & de Alencar, S. M. (2020). Simulated gastrointestinal digestion of Brazilian açaí seeds affects the content of flavan-3-ol derivatives, and their antioxidant and anti-inflammatory activities. *Helijon, 6(10)*. <https://doi.org/10.1016/j.heliyon.2020.e05214>.
- Melo, P. S., Selani, M. M., Goncalves, R. H., Paulino, J. D., Massarioli, A. P., & de Alencar, S. M. (2021). Açaí seeds: An unexplored agro-industrial residue as a potential source of lipids, fibers, and antioxidant phenolic compounds. *Industrial Crops and Products, 161*. <https://doi.org/10.1016/j.indcrop.2020.113204>.
- Moorthy, M., Chaiyakunapruk, N., Jacob, S.A., Palanisamy, U.D. (2020). Prebiotic potential of polyphenols, its effect on gut microbiota and anthropometric/clinical markers: A systematic review of randomized controlled trials. *Trends in Food Science & Technology, 99*, 634-649. <https://doi.org/10.1016/j.tifs.2020.03.036>.
- Mutua, J.K., Imathi, S., Owino, W. (2017). Evaluation of the proximate composition, antioxidant potential, and antimicrobial activity of mango seed kernel extracts. *Food Science & Nutrition, 5*, 349-357. <https://doi.org/10.1002/fsn3.399>.
- Neves, L. C., Tosin, J. M., Benedette, R. M., & Cisneros-Zevallos, L. (2015). Post-harvest nutraceutical behaviour during ripening and senescence of 8 highly perishable fruit species from the Northern Brazilian Amazon region. *Food Chemistry, 174*, 188-196. <https://doi.org/10.1016/j.foodchem.2014.10.111>.
- Oliveira, A.L. (2006). Elemental contents in exotic Brazilian tropical fruits evaluated by energy dispersive X-ray fluorescence. *Scientia Agricola, 63*, 82-84. <https://doi.org/10.1590/S0103-90162006000100013>.
- Oliveira, S.R., Chacón-Madrid, K., Arruda, M.A.Z., Júnior, F.B. (2019). *In vitro* gastrointestinal digestion to evaluate the total, bioaccessible and bioavailable concentrations of iron and manganese in açaí (*Euterpe oleracea Mart.*) pulps. *Journal of Trace Elements in Medicine and Biology, 53*, 27-33. <https://doi.org/10.1016/j.jtemb.2019.01.016>.
- Ozdal, T., Sela, D.A., Xiao, J., Boyacioglu, D., Chen, F., Capanoglu, E. (2016). The Reciprocal Interactions between Polyphenols and Gut Microbiota and Effects on Bioaccessibility. *Nutrients, 8*, 78. <https://doi.org/10.3390/nu8020078>.
- Pacheco-Palencia, L. A., Mertens-Talcott, S., & Talcott, S. T. (2008). Chemical composition, antioxidant properties, and thermal stability of a phytochemical enriched oil from acai (*Euterpe oleracea Mart.*). *Journal of Agricultural and Food Chemistry, 56(12)*, 4631-4636. <https://doi.org/10.1021/jf800161u>.
- Pandey, K.B., Rizvi, S.I. (2009). Plant polyphenols as dietary antioxidants in human health and disease. *Oxidative Medicine and Cellular Longevity, 2*, 270-278. <https://doi.org/10.4161/oxim.2.5.9498>.
- Pessoa, J. D. C., Arduin, M., Martins, M. A., & de Carvalho, J. E. U. (2010). Characterization of Açaí (*E. oleracea*) Fruits and its Processing Residues. *Brazilian Archives of Biology and Technology, 53(6)*, 1451-1460. <https://doi.org/10.1590/S1516-89132010000600022>.

- Pompeu, D. R., Silva, E. M., & Rogez, H. (2009). Optimisation of the solvent extraction of phenolic antioxidants from fruits of *Euterpe oleracea* using Response Surface Methodology. *Bioresource Technology*, 100(23), 6076-6082. <https://doi.org/10.1016/j.biortech.2009.03.083>.
- Pop, C., Suharoschi, R., Pop, O.L. (2021). Dietary Fiber and Prebiotic Compounds in Fruits and Vegetables Food Waste. *Sustainability*, 13, 7219. <https://doi.org/10.3390/su13137219>.
- Qayyum, M.M.N., Butt, M.S., Anjum, F.M., Nawaz, H. (2012). Composition analysis of some selected legumes for protein isolates recovery. *The Journal of Animal & Plant Sciences*, 22, 1156-1162.
- Radzik, P.L., Klewicka, E. (2021). Mutual influence of polyphenols and *Lacticaseibacillus* spp. bacteria in food: a review. *European Food Research and Technology*, 247, 9-24. <https://doi.org/10.1007/s00217-020-03603-y>.
- Ramos, A. S., Souza, R. O. S., Boleti, A. P. D., Bruginski, E. R. D., Lima, E. S., Campos, F. R., & Machado, M. B. (2015). Chemical characterization and antioxidant capacity of the araca-pera (*Psidium acutangulum*): An exotic Amazon fruit. *Food Research International*, 75, 315-327. <https://doi.org/10.1016/j.foodres.2015.06.026>.
- Reboreda-Rodriguez, P., Gonzalez-Barreiro, C., Martinez-Carballo, E., Cambeiro-Perez, N., Rial-Otero, R., Figueiredo-Gonzalez, M., & Cancho-Grande, B. (2021). Applicability of an *In-Vitro* Digestion Model to Assess the Bioaccessibility of Phenolic Compounds from Olive-Related Products. *Molecules*, 26(21). <https://doi.org/10.3390/molecules26216667>.
- Requena, T., Monagas, M., Pozo-Bayon, M. A., Martin-Alvarez, P. J., Bartolome, B., del Campo, R., . . . Moreno-Arribas, M. V. (2010). Perspectives of the potential implications of wine polyphenols on human oral and gut microbiota. *Trends in Food Science & Technology*, 21(7), 332-344. <https://doi.org/10.1016/j.tifs.2010.04.004>.
- Rodrigues, A.M.C., Darnet, S., Silva, L.H.M. (2010). Fatty acid profiles and tocopherol contents of buriti (*Mauritia flexuosa*), patawa (*Oenocarpus bataua*), tucuma (*Astrocaryum vulgare*), mari (*Poraqueiba paraensis*) and inaja (*Maximiliana maripa*) fruits. *Journal of the Brazilian Chemical Society*, 21. <https://doi.org/10.1590/S0103-50532010001000028>.
- Rodrigues, R. B., Lichtenthaler, R., Zimmermann, B. F., Papagiannopoulos, M., Fabricius, H., & Marx, F. (2006). Total oxidant scavenging capacity of *Euterpe oleracea* Mart. (acai) seeds and identification of their polyphenolic compounds. *Journal of Agricultural and Food Chemistry*, 54(12), 4162-4167. <https://doi.org/10.1021/jf058169p>.
- Rojas-Garcia, A., Fuentes, E., Cadiz-Gurrea, M. L., Rodriguez, L., Villegas-Aguilar, M. D. C., Palomo, I., . . . Segura-Carretero, A. (2022). Biological Evaluation of Avocado Residues as a Potential Source of Bioactive Compounds. *Antioxidants (Basel)*, 11(6). <https://doi.org/10.3390/antiox11061049>.
- Rufino, M. D. M., Alves, R. E., de Brito, E. S., Perez-Jimenez, J., Saura-Calixto, F., & Mancini, J. (2010). Bioactive compounds and antioxidant capacities of 18 non-traditional tropical fruits from Brazil. *Food Chemistry*, 121(4), 996-1002. <https://doi.org/10.1016/j.foodchem.2010.01.037>.
- Rufino, M. D. M., Perez-Jimenez, J., Arranz, S., Alves, R. E., de Brito, E. S., Oliveira, M. S. P., & Saura-Calixto, F. (2011). Açaí (*Euterpe oleracea*) 'BRS Para': A tropical fruit source of antioxidant dietary fiber and high antioxidant capacity oil. *Food Research International*, 44(7), 2100-2106. <https://doi.org/10.1016/j.foodres.2010.09.011>.
- Ryan, E. M., Duryee, M. J., Hollins, A., Dover, S. K., Pirruccello, S., Sayles, H., . . . Mikuls, T. R. (2019). Antioxidant properties of citric acid interfere with the uricase-based measurement of circulating uric acid. *Journal of Pharmaceutical and Biomedical Analysis*, 164, 460-466. <https://doi.org/10.1016/j.jpba.2018.11.011>.
- Sánchez-Camargo, A.P., Gutiérrez, L.F., Vargas, S.M., Martinez-Correa, H.A., Parada-Alfonso, F., Narváez-Cuenca, C.E. (2019). Valorisation of mango peel: Proximate composition, supercritical fluid extraction of carotenoids, and application as an antioxidant additive for an edible oil. *The Journal of Supercritical Fluids*, 152. <https://doi.org/10.1016/j.supflu.2019.104574>.

- Santonocito, D., Granata, G., Geraci, C., Panico, A., Siciliano, E. A., Raciti, G., & Puglia, C. (2020). Carob Seeds: Food Waste or Source of Bioactive Compounds? *Pharmaceutics*, 12(11).
- <https://doi.org/10.3390/pharmaceutics12111090>.
- Santos-Burgoa, C., Rios, C., Mercado, L.A., Arechiga-Serrano, R., Cano-Valle, F., Eden-Wynter, R.A., Texcalac-Sangrador, J.L., Villa-Barragan, J.P., Rodriguez-Agudelo, Y., Montes, S. (2001). Exposure to Manganese: Health Effects on the General Population,a Pilot Study in Central Mexico. *Environmental Research Section*, 85, 90-104. <https://doi.org/10.1006/enrs.2000.4108>.
- Sardi, J. D. O., Freires, I. A., Lazarini, J. G., Infante, J., de Alencar, S. M., & Rosalen, P. L. (2017). Unexplored endemic fruit species from Brazil: Antibiofilm properties, insights into mode of action, and systemic toxicity of four *Eugenia* spp. *Microbial Pathogenesis*, 105, 280-287.
- <https://doi.org/10.1016/j.micpath.2017.02.044>.
- Saura-Calixto, F., Serrano, J., & Goni, I. (2007). Intake and bioaccessibility of total polyphenols in a whole diet. *Food Chemistry*, 101(2), 492-501. <https://doi.org/10.1016/j.foodchem.2006.02.006>.
- Scalzo, J., Battino, M., Costantini, E., Mezzetti, B. (2005). Breeding and biotechnology for improving berry nutritional quality. *BioFactors*, 23, 213-220. <https://doi.org/10.1002/biof.5520230406>.
- Schieber, A., Stintzing, F.C., Carle, R. (2001). By-products of plant food processing as a source of functional compounds — recent developments. *Trends in Food Science & Technology*, 11, 401-413. [https://doi.org/10.1016/S0924-2244\(02\)00012-2](https://doi.org/10.1016/S0924-2244(02)00012-2).
- Selani, M.M., Canniatti-Brazaca, S.G., Dias, C.T. dos S., Ratnayake, W.S., Flores, R.A., Bianchini, A. (2014). Characterisation and potential application of pineapple pomace in an extruded product for fibre enhancement. *Food Chemistry*, 163, 23–30. <https://doi.org/10.1016/j.foodchem.2014.04.076>.
- Sette, P., Fernandez, A., Soria, J., Rodriguez, R., Salvatori, D., & Mazza, G. (2020). Integral valorization of fruit waste from wine and cider industries. *Journal of Cleaner Production*, 242. <https://doi.org/10.1016/j.jclepro.2019.118486>.
- Shah, M., Khaliq, F., Nawaz, H., Rahim, F., Ullah, N., Javed, M.S., Amjad, A., Nishan, U., Ullah, S., Ahmed, S., Jalil, N.A.C. (2022). Comparative evaluation of proximate composition and biological activities of peel extracts of three commonly consumed fruits. *Food Science and Technology*, 42. <https://doi.org/10.1590/fst.61021>.
- Shanley, P., Cymerys, M., Serra, M., & Medina, G. (2012). *Frutales y plantas útiles en la vida amazónica*: FAO/CIFOR/PPI.
- Shanley, P., Medina, G. (2005). Frutíferas e Plantas Úteis na Vida Amazônica. Belém (Brasil): Instituto do Homem e Meio Ambiente da Amazônia/Center for International Forestry Research, 300p.
- Shanley, P., Medina, G., Cordeiro, S., & Imbiriba, M. (2005). *Frutíferas e plantas úteis na vida amazônica*: Cifor.
- Shi, J., Zhu, L., Li, Y., Zheng, H., Yu, J., Lu, L., & Liu, Z. (2016). *In Vitro* Study of UGT Metabolism and Permeability of Orientin and Isoorientin, Two Active flavonoid C-glycosides. *Drug Metab Lett*, 10(2), 101-110. <https://doi.org/10.2174/1872312810666160219121217>.
- Silva, A.F.S., Martins, L.C., Moraes, L.M.B., Gonçalves, I.C., Godoy, B.B.R., Erasmus, S.W., Van Ruth, S., Rocha, F.R.P. (2021). Can Minerals Be Used as a Tool to Classify Cinnamon Samples? *Proceedings*, 70. https://doi.org/10.3390/foods_2020-07652.
- Silva, M. R., Freitas, L. G., Souza, A. G., Araújo, R. L., Lacerda, I. C., Pereira, H. V., . . . Melo, J. O. (2019). Antioxidant activity and metabolomic analysis of cagaitas (*Eugenia dysenterica*) using paper spray mass spectrometry. *Journal of the Brazilian Chemical Society*, 30, 1034-1044.
- Simmler, C., Pauli, G. F., & Chen, S. N. (2013). Phytochemistry and biological properties of glabridin. *Fitoterapia*, 90, 160-184. <https://doi.org/10.1016/j.fitote.2013.07.003>.

- Singh, A.K., Cabral, C., Kumar, R., Ganguly, R., Rana, H.K., Gupta, A., Lauro, M.R., Carbone, C., Reis, F., Pandey, A.K. (2019). Beneficial Effects of Dietary Polyphenols on Gut Microbiota and Strategies to Improve Delivery Efficiency. *Nutrients*, 11. <https://doi.org/10.3390/nu11092216>.
- Singh, T.C. (2015). Avaliação dos parâmetros físico-químicos e estabilidade de compostos bioativos em óleos de polpa e amêndoas de frutos amazônicos. Tese (Doutorado). Universidade Estadual Paulista “Júlio de Mesquita Filho”, Instituto de Biociências, Letras e Ciências Exatas, São José do Rio Preto, São Paulo. 158 p.
- Slavin, J. (2013). Fiber and Prebiotics: Mechanisms and Health Benefits. *Nutrients*, 5, 1417-1435. <https://doi.org/10.3390/nu5041417>.
- Soetan, K.O., Olaifa, C.O., Oyewole, O.E. (2010). The importance of mineral elements for humans, domestic animals and plants: A review. *African Journal of Food Science*, 4, 200-222.
- Stachiw, R., Ribeiro, S. B., Jardim, M. A. G., Possimoser, D., Alves, W. D., & Cavalheiro, W. C. S. (2016). Potential of biodiesel production with oil seed native species from Rondonia, Brazil. *Acta Amazonica*, 46(1), 81-90. <https://doi.org/10.1590/1809-4392201501151>.
- Statista. (2022a). Açaí berry crop production in Brazil in 2021, by state. Available: <https://www.statista.com/statistics/1069905/production-acai-brazil-state/> [Accessed 12/10/2022].
- Statista. (2022b). Brazil: açaí berry production 2016-2021. Available: <https://www.statista.com/statistics/1069776/production-acai-brazil/> [Accessed 12/10/2022].
- Taamalli, A., Arraez-Roman, D., Abaza, L., Iswaldi, I., Fernandez-Gutierrez, A., Zarrouk, M., & Segura-Carretero, A. (2015). LC-MS-based metabolite profiling of methanolic extracts from the medicinal and aromatic species *Mentha pulegium* and *Origanum majorana*. *Phytochemical Analysis*, 26(5), 320-330. <https://doi.org/10.1002/pca.2566>.
- Taiz, L., Zeiger, E., Briskin, D., Bloom, A. (2013). Nutrição Mineral. In: Taiz L, Zeiger E. Fisiologia Vegetal. 5ª edição. Porto Alegre: Artmed, 2013, Cap. 5, pp. 107-130. 918p.
- Tena, N., Martin, J., & Asuero, A. G. (2020). State of the Art of Anthocyanins: Antioxidant Activity, Sources, Bioavailability, and Therapeutic Effect in Human Health. *Antioxidants (Basel)*, 9(5). <https://doi.org/10.3390/antiox9050451>.
- Thiyagarajan, P., Chandrasekaran, C. V., Deepak, H. B., & Agarwal, A. (2011). Modulation of lipopolysaccharide-induced pro-inflammatory mediators by an extract of *Glycyrrhiza glabra* and its phytoconstituents. *Inflammopharmacology*, 19(4), 235-241. <https://doi.org/10.1007/s10787-011-0080-x>.
- Torres-León, C., Ramírez-Guzman, N., Londoño-Hernandez, L., Martínez-Medina, G.A., Díaz-Herrera, R., Navarro-Macias, V., Alvarez-Pérez, O.B., Picazo, B., Villarreal-Vázquez, M., Ascacio-Valdes, J., Aguilar, C.N. (2018). Food Waste and Byproducts: An Opportunity to Minimize Malnutrition and Hunger in Developing Countries. *Frontiers in Sustainable Food Systems*, 2. <https://doi.org/10.3389/fsufs.2018.00052>.
- Tremocoldi, M.A., Rosalen, P.L., Franchin, M., Massarioli, A.P., Denny, C., Daiuto, E.R., Paschoal, J.A.R., Melo, P.S., de Alencar, S.M. (2018). Exploration of avocado by-products as natural sources of bioactive compounds. *Plos One*, 13. <https://doi.org/10.1371/journal.pone.0192577>.
- Velderrain-Rodriguez, G. R., Palafox-Carlos, H., Wall-Medrano, A., Ayala-Zavala, J. F., Chen, C. Y. O., Robles-Sánchez, M., . . . Gonzalez-Aguilar, G. A. (2014). Phenolic compounds: their journey after intake. *Food & Function*, 5(2), 189-197. <https://doi.org/10.1039/c3fo60361j>.
- Wang, G., Lei, Z., Zhong, Q., Wu, W., Zhang, H., Min, T., . . . Lai, F. (2017). Enrichment of caffeic acid in peanut sprouts and evaluation of its *in vitro* effectiveness against oxidative stress-induced erythrocyte hemolysis. *Food Chemistry*, 217, 332-341. <https://doi.org/10.1016/j.foodchem.2016.07.126>.

- Wojtunik-Kulesza, K., Oniszczuk, A., Oniszczuk, T., Combrzynski, M., Nowakowska, D., & Matwijczuk, A. (2020). Influence of *In Vitro* Digestion on Composition, Bioaccessibility and Antioxidant Activity of Food Polyphenols-A Non-Systematic Review. *Nutrients*, 12(5). <https://doi.org/10.3390/nu12051401>.
- Yamaguchi, K. K. D., Pereira, L. F. R., Lamarao, C. V., Lima, E. S., & da Veiga, V. F. (2015). Amazon acai: Chemistry and biological activities: A review. *Food Chemistry*, 179, 137-151. <https://doi.org/10.1016/j.foodchem.2015.01.055>.
- Yang, X., Wang, T., Guo, J., Sun, M., Wong, M. W., & Huang, D. (2019). Dietary Flavonoids Scavenge Hypochlorous Acid via Chlorination on A- and C-Rings as Primary Reaction Sites: Structure and Reactivity Relationship. *J Agric Food Chem*, 67(15), 4346-4354. <https://doi.org/10.1021/acs.jafc.8b06689>.
- Yokota, T., Nishio, H., Kubota, Y., & Mizoguchi, M. (1998). The inhibitory effect of glabridin from licorice extracts on melanogenesis and inflammation. *Pigment Cell Res*, 11(6), 355-361. <https://doi.org/10.1111/j.1600-0749.1998.tb00494.x>.
- Yuyama, L.K.O., Aguiar, J.P.L., Filho, D.F.S., Yuyama, K., Varejão, M.J., Fávaro, D.I.T., Vasconcellos, M.B.A., Pimentel, A.S., Caruso, M.S.F. (2011) Caracterização físico-química do suco de açaí de *Euterpe precatoria* Mart. oriundo de diferentes ecossistemas amazônicos. *Acta Amazonica*, 41, 545-552.
- Zahid, H. F., Ali, A., Ranadheera, C. S., Fang, Z. X., Dunshea, F. R., & Ajlouni, S. (2022). *In vitro* bioaccessibility of phenolic compounds and alpha-glucosidase inhibition activity in yoghurts enriched with mango peel powder. *Food Bioscience*, 50. <https://doi.org/10.1016/j.fbio.2022.102011>.
- Zhang, H., & Tsao, R. (2016). Dietary polyphenols, oxidative stress and antioxidant and anti-inflammatory effects. *Current Opinion in Food Science*, 8, 33-42. <https://doi.org/10.1016/j.cofs.2016.02.002>.
- Zhang, Y., Xiong, H., Xu, X. F., Xue, X., Liu, M. N., Xu, S. Y., . . . Li, X. R. (2018). Compounds Identification in Semen Cuscutae by Ultra-High-Performance Liquid Chromatography (UPLCs) Coupled to Electrospray Ionization Mass Spectrometry. *Molecules*, 23(5). <https://doi.org/10.3390/molecules23051199>.
- Zhu, L., Um, T., Ma, M., Sun, H., Zhao, G. (2022). Nutritional composition, antioxidant activity, volatile compounds, and stability properties of sweet potato residues fermented with selected lactic acid bacteria and bifidobacterial. *Food Chemistry*, 374, 131500. <https://doi.org/10.1016/j.foodchem.2021.131500>.