

UNIVERSIDADE DE SÃO PAULO
Faculdade de Filosofia, Ciências e Letras de Ribeirão Preto
Departamento de Química
Programa de Pós-Graduação em Química

“Identificação e isolamento de fitotoxinas produzidas por actinobactérias isoladas da Caatinga”

Osvaldo Luiz Ferreira Junior

Orientador: Prof. Dr. Luiz Alberto Beraldo de Moraes

Dissertação apresentada à Faculdade de Filosofia, Ciências e Letras de Ribeirão Preto da Universidade de São Paulo, como parte das exigências para a obtenção do título de Mestre em Ciências, Área: **Química.**

RIBEIRÃO PRETO
2018

RESUMO

FERREIRA JUNIOR, Osvaldo Luiz.: Identificação e isolamento de fitotoxinas produzidas por actinobactérias isoladas da Caatinga. 2018, 109 f. Dissertação de mestrado. Faculdade de Filosofia, Ciências e Letras de Ribeirão Preto, Universidade de São Paulo, Ribeirão Preto, 2018.

Palavras chave: Espectrometria de massas; Produtos naturais; Fitotoxina; Albociclina; Actinobactéria; Caatinga.

As actinobactérias são uma reconhecida fonte de metabólitos secundários bioativos com grande diversidade estrutural e funcional. Dentre elas, o gênero *Streptomyces* é sem dúvida o mais explorado, correspondendo a cerca de 75% dos metabólitos secundários estudados, e por dois terços dos antibióticos conhecidos. Dessa forma, ainda é crescente a necessidade de novos herbicidas com perfil de baixa toxicidade ambiental e novos mecanismos de ação, para a substituição de produtos que são utilizados extensivamente há anos. Grandes avanços tecnológicos têm sido alcançados em termos de instrumentação analítica nas últimas décadas, tornando a espectrometria de massas a técnica mais eficiente e robusta na identificação de metabólitos secundários, principalmente quando acoplada a técnicas cromatográficas de ultra eficiência (UHPLC-MS). Nessa dissertação foi explorada a diversidade metabólica de actinobactérias isoladas do bioma Caatinga para a identificação de metabólitos secundários com atividade fitotóxica. Foram produzidos 166 extratos brutos de actinobactérias, das quais, 27 apresentaram algum nível de atividade fitotóxica contra *Lemna minor*. Devido à pronunciada bioatividade ($MIC = 6,25 \mu\text{g}$) o extrato bruto da actinobactéria, CAAT 7-52, foi selecionado para desreplicação, caracterização e fracionamento guiado por bioensaios, e, ao mesmo tempo, um estudo taxonômico foi realizado para determinar o posicionamento filogenético do novo isolado. A análise da sequência do gene 16S rRNA suportou a classificação da linhagem no gênero *Streptomyces* e mostrou que CAAT 7-52 formou uma linha filética distinta junto com a linhagem tipo de *Streptomyces actinomycinicus*, com um valor de identidade da sequência de 99,0%. No extrato bruto de CAAT 7-52 foi identificada a Albociclina como responsável pela atividade fitotóxica ($MIC = 3,12 \mu\text{g}$), assim como alguns compostos análogos. O monitoramento da massa micelial seca e produção de Albociclina mostrou uma maior produção do composto ativo aos 21 dias de crescimento. Esse extrato foi submetido a ensaios de inibição de germinação, onde apresentou atividade contra sementes de buva (*Conyza canadensis*) ($MIC = 12,5 \mu\text{g}$), alface (*Lactuca sativa*) e rúcula (*Eruca sativa*). Variações no meio de cultivo mostraram uma produção seletiva de Albociclina nos meios de cultivo PMB e Czapecck-GL. Ensaios de fitotoxicidade contra *Lemna minor*, utilizando o fermentado do meio PMB sem extração com solventes orgânicos, mostrou atividade em uma diluição de 20%. Portanto, foi possível demonstrar através desse estudo o grande potencial das actinobactérias na produção de metabólitos secundários bioativos e da espectrometria de massas como técnica analítica na identificação dos compostos ativos.

ABSTRACT

FERREIRA JUNIOR, Osvaldo Luiz.: **Identification and isolation of phytotoxins produced by actinobacteria isolated from Caatinga.** 2018, 109 f. Dissertação de mestrado. Faculdade de Filosofia, Ciências e Letras de Ribeirão Preto, Universidade de São Paulo, Ribeirão Preto, 2018.

Keywords: Mass spectrometry; Natural products; Phytotoxin; Albocycline; Actinobacteria; Caatinga.

Actinobacterias are a well known source of bioactive secondary metabolites with great structural and functional diversity. Among them, the *Streptomyces* genus is certainly the most explored, corresponding to about 75% of studied secondary metabolites, and for two thirds of known antibiotics. Thus, there is still a growing need for new herbicides with low environmental toxicity and new modes of action, for the replacement of products that have been broadly used for years. Great technological progress has been achieved in analytical instrumentation in the last few decades, making mass spectrometry the most efficient and robust technique to identify secondary metabolites, mainly when coupled to ultra-performance chromatographic techniques (UHPLC-MS). In this essay, was explored the metabolic diversity of the actinobacterias isolated from the Caatinga biome to identification of secondary metabolites with phytotoxic activity. Were produced 166 crude extracts from actinobacterias, from this, 27 showed some phytotoxic activity against *Lemna minor*. Due to noticeable bioactivity ($MIC = 6,25 \mu\text{g}$) the crude extract from actinobacteria CAAT 7-52, was selected to dereplication, characterization and bioassay guided fractionation, and, at the same time, a taxonomic study was carried out to determinate the phylogenetic location of the new isolated. The analysis of 16S rRNA gene sequence supported the lineage classification on the genus *Streptomyces* and established that CAAT 7-52 assembled a distinct phyletic line together with the lineage type of *Streptomyces actinomycinicus*, with an identity sequence value of 99,0%. In the crude extract CAAT 7-52, Albocycline was identified as the responsible for the phytotoxic activity ($MIC = 3,12 \mu\text{g}$), so as some analogous compounds. The mycelial dry mass and Albocycline production monitoring revealed a major production of the active compound at 21 days of growth. This extract was subjected to germination inhibition assays, where revealed activity against horseweed (*Conyza canadensis*) ($MIC = 12,5 \mu\text{g}$), lettuce (*Lactuca sativa*) and arugula (*Eruca sativa*) seeds. Variations in the culture media showed a selective production of Albocycline in the PMB and Czapeck-GL culture media. Phytotoxicity assays against *Lemna minor*, applying the fermented PMB media without organic solvents extractions, revealed activity with a 20% dilution. Therefore, through this study was able to demonstrate the great potential of the actinobacterias in the production of bioactive secondary metabolites and the mass spectrometry as analytical technique to identification of the active compounds.

CONCLUSÃO

De um total de 166 extratos brutos obtidos, 27 (16%) apresentaram algum nível de atividade fitotóxica com *Lemna minor*. Dentre eles, o extrato CAAT 7-52 (MIC = 6,25 µg) apresentou atividade fitotóxica mais pronunciada, o qual foi submetido ao fracionamento guiado por bioensaio para desreplicação dos compostos ativos por espectrometria de massas, onde foi observada a presença de dois compostos ativos os quais foram caracterizados como sendo o ácido 3-hidroxibenzóico presente na fração 15, e a Albociclina presente na fração 30 (MIC = 3,12 µg). Ambos os compostos não possuem relatos na literatura sobre possuírem atividade fitotóxica. O extrato CAAT 7-52 também apresentou atividade de inibição de germinação contra sementes de alface, rúcula e buva, todas dicotiledôneas, esta última sendo a mais promissora em razão de seu difícil controle. A actinobactéria CAAT 7-52 foi identificada através do sequenciamento do gene 16S rRNA apresentando um valor de identidade de sequência de 99% com *Streptomyces actinomycinicus*. Também foi verificada a atividade do meio de cultivo fermentado, sem extração com solventes orgânicos, o qual apresentou atividade fitotóxica até uma proporção de 20%, sendo de interesse em razão de possível utilização em teste para controle direto de plantas-daninhas. Ocorre a produção seletiva da albociclina quando utilizado o meio de cultivo PMB e Czapecck com xarope de glicose e a produção seletiva dos análogos da Albociclina em meio de cultivo ISP-2, sendo que ambos os extratos apresentaram atividade fitotóxica significativa. A partir do monitoramento da massa micelial e da produção de Albociclina, foi apresentado um máximo de produção de Albociclina em torno de 21 dias. Foram identificados os compostos indol-3-carboxaldeído, ácido benzacético e benzacetamida nas frações 14, 15 e 22 do extrato AM 33-22, os quais apresentaram atividade fitotóxica em grau moderado contra *Lemna minor*. Ainda que mais experimentos e otimizações necessitem ser realizados, tais resultados mostram o enorme potencial de aplicação biotecnológica das actinobactérias e de seus metabólitos secundários, como os obtidos nesse trabalho.

4. REFERÊNCIAS

1. Cragg, G. M. & Newman, D. J. Biodiversity: A continuing source of novel drug leads. *Pure Appl. Chem.* **77**, 7–24 (2005).
2. Dias, D. A., Urban, S. & Roessner, U. A Historical overview of natural products in drug discovery. *Metabolites* **2**, 303–336 (2012).
3. Kaur, P. K., Kaur, J. & Saini, H. S. Antifungal potential of *Bacillus vallismortis* R2 against different phytopathogenic fungi. *Spanish J. Agric. Res.* **13**, 1–11 (2015).
4. Suma, K. L. S., Dev, A. & Florence, E. J. M. Identification and Characterization of Lipopeptides from *Bacillus subtilis* B1 Against Sapstain Fungus of Rubberwood Through MALDI-TOF-MS and RT-PCR. *Curr. Microbiol.* **73**, 46–53 (2016).
5. Zarins-Tutt, J. S. *et al.* Prospecting for new bacterial metabolites: a glossary of approaches for inducing, activating and upregulating the biosynthesis of bacterial cryptic or silent natural products. *Nat. Prod. Rep.* **33**, 54–72 (2016).
6. Terhonen, E., Sipari, N. & Asiegbu, F. O. Inhibition of phytopathogens by fungal root endophytes of Norway spruce. *Biol. Control* **99**, 53–63 (2016).
7. Lu, Y. H. *et al.* Isolation, identification, derivatization and phytotoxic activity of secondary metabolites produced by *Cladosporium oxysporum* DH14, a locust-associated fungus. *J. Integr. Agric.* **15**, 832–839 (2016).
8. Fleming, A. On the antibacterial action of cultures of a penicillium, with special reference to their use in the isolation of *B. influenzae*. 1929. *Br. J. Exp. Pathol.* **10**, 226–236 (1929).
9. Dembitsky, V. M., Savidov, N., Poroikov, V. V, Gloriozova, T. A. & Imbs, A. B. Naturally occurring aromatic steroids and their biological activities. 4663–4674 (2018).
10. Popova, V. *et al.* Biologically active components in seeds of three *Nicotiana* species. *Ind. Crops Prod.* **117**, 375–381 (2018).
11. Liu, X. *et al.* Bioprospecting microbial natural product libraries from the marine environment for drug discovery. *J. Antibiot. (Tokyo)*. **63**, 415–422 (2010).
12. Jensen, P. R. A metabolomics guided exploration of marine natural product chemical

space. *Metabolomics* (2016). doi:10.1007/s11306-016-1087-5

13. Solecka, J., Zajko, J., Postek, M. & Rajnisz, A. Biologically active secondary metabolites from Actinomycetes. *Open Life Sci.* **7**, 373–390 (2012).
14. Katz, L. & Baltz, R. H. Natural product discovery: past, present, and future. *J. Ind. Microbiol. Biotechnol.* **43**, 155–176 (2016).
15. Dougherty, T. J. & Pucci, M. J. *Antibiotic Discovery and Development. Antibiotic Discovery and Development* (Springer Science+Business Media, 2014). doi:10.1007/978-1-4614-1400-1
16. White, R. J. The Early History of Antibiotic Discovery: Empiricism Ruled. in *Antibiotic Discovery and Development* (eds. Dougherty, T. J. & Pucci, M. J.) 3–31 (Springer Science+Business Media, 2014).
17. Bills, G. F. & Gloer, J. B. Biologically Active Secondary Metabolites from the Fungi. *Microbiol. Spectr.* **4**, (2016).
18. CropLife. 4,500 Years of Crop Protection. *Crop Life International* (2017). Available at: <https://croplife.org/news/4500-years-of-crop-protection/>. (Accessed: 6th June 2018)
19. University, C. The History of Integrated Pest Management. *Cornell University* Available at: <https://courses.cit.cornell.edu/ipm444/lec-notes/extra/ipm-history.html>. (Accessed: 6th June 2018)
20. Vats, S. Herbicides: History, Classification and Genetic Manipulation of Plants for Herbicide Resistance. in *Sustainable Agriculture Reviews* (ed. Lichtfouse, E.) **15**, (Springer International Publishing Switzerland, 2015).
21. Zhao, Y. Auxin biosynthesis and its role in plant development. *Annu Rev Plant Biol* **61**, 49–64 (2010).
22. Alterman, M. K. & Neptune, A. M. L. Efeito do Ácido 2,4-Diclorofenoxyacético (2,4-D) na Absorção do Fósforo (32P) pelo Trigo (*Triticum aestivum*, L) e a Sua Distribuição na Planta. *An. da Esc. Super. Agric. 'Luiz Queiroz'* **34**, 10 (1977).
23. Sparks, T. C., Hahn, D. R. & Garizi, N. V. Natural products, their derivatives, mimics and synthetic equivalents: role in agrochemical discovery. *Pest Manag. Sci.* **73**, 700–715 (2017).

24. Heap, I. The International Survey of Herbicide Resistant Weeds. (2018). Available at: www.weedscience.org. (Accessed: 12th June 2018)
25. Mahmood, Q., Bilal, M. & Jan, S. *Herbicides, Pesticides, and Plant Tolerance: An Overview. An Overview. Emerging Technologies and Management of Crop Stress Tolerance: Biological Techniques* **1**, (Elsevier Inc., 2014).
26. Tanaka, Y. & Omura, S. Agroactive Compounds of Microbial Origin. *Annu. Rev. Microbiol.* **47**, 57–87 (1993).
27. Ujváry, I. *Pest Control Agents from Natural Products. Hayes' Handbook of Pesticide Toxicology Volume 1*, (Elsevier Inc., 2010).
28. Hoerlein, G. Glufosinate (phosphinothricin), a natural amino acid with unexpected herbicidal properties. *Rev. Environ. Contam. Toxicol.* **138**, 73–145 (1994).
29. Duke, S. O. The history and current status of glyphosate. *Pest Manag. Sci.* (2017). doi:10.1002/ps.4652
30. Dekker, J. & Duke, S. O. Herbicide-Resistant Field Crops. *Adv. Agron.* **54**, 69–116 (1995).
31. Duke, S. O. & Powles, S. B. Glyphosate: a once-in-a-century herbicide. *Pest Manag. Sci.* **64**, 319–325 (2008).
32. Moss, S. Herbicide Resistance in Weeds. in *Weed Research: Expanding Horizons* (eds. Hatcher, P. E. & Froud-Williams, R. J.) 181–214 (John Wiley & Sons Ltd., 2017).
33. Benbrook, C. M. Trends in glyphosate herbicide use in the United States and globally. *Environ. Sci. Eur.* **28**, 15 (2016).
34. Adegas, F. S. *et al.* Impacto econômico da resistência de plantas daninhas a herbicidas no Brasil Introdução. *Circ. Técnica* **132**, 12 (2017).
35. Lanfranconi, L. E. & Gazziero, D. L. P. Herbicide resistant weeds. *CropLife Latin America* Available at: <https://www.croplifela.org/en/diseases/herbicide-resistant-weeds>. (Accessed: 18th June 2018)
36. Embrapa. Bayer e Embrapa pesquisam plantas daninhas resistentes a herbicidas. *Pesquisa, Desenvolvimento e Inovação* (2018). Available at: <https://www.embrapa.br/busca-de-noticias/-/noticia/31269822/bayer-e-embrapa-pesquisam-plantas-daninhas-resistentes-a-herbicidas>

pesquisam-plantas-daninhas-resistentes-a-herbicidas. (Accessed: 18th June 2018)

37. Heap, I. & Duke, S. O. Overview of glyphosate-resistant weeds worldwide. *Pest Manag. Sci.* **74**, 1040–1049 (2018).
38. Duke, S. O., Owens, D. K. & Dayan, F. E. The growing need for biochemical bioherbicides. *ACS Symp. Ser.* **1172**, 31–43 (2014).
39. Taketani, R. G., Kavamura, V. N. & Santos, S. N. Diversity and Technological Aspects of Microorganisms from Semiarid Environments. in *Diversity and Benefits of Microorganisms from the Tropics* (eds. de Azevedo, J. L. & Quecine, M. C.) 1–439 (Springer International Publishing AG, 2017). doi:10.1007/978-3-319-55804-2
40. Kavamura, V. N., Taketani, R. G., Ferreira, C., de Melo, I. S. & Mendes, R. The role of species turnover in structuring bacterial communities in a local scale in the cactus rhizosphere. *Plant Soil* **425**, 101–112 (2018).
41. Araújo, E. L., Castro, C. C. & Albuquerque, U. P. Dynamics of Brazilian Caatinga – A Review Concerning the Plants , Environment and People. *Funct. Ecosyst. Communities* **1**, 15–28 (2007).
42. Kavamura, V. N. Bactérias associadas às cactáceas da Caatinga: promoção de crescimento de plantas sob estresse hídrico. (Universidade de São Paulo - ESALQ, 2012).
43. Taketani, R. G. *et al.* Dry Season Constrains Bacterial Phylogenetic Diversity in a Semi-Arid Rhizosphere System. *Microb. Ecol.* **73**, 153–161 (2017).
44. Kavamura, V. N. *et al.* Water Regime Influences Bulk Soil and Rhizosphere of Cereus jamacaru Bacterial Communities in the Brazilian Caatinga Biome. *PLoS One* **8**, (2013).
45. Taketani, R. G., Kavamura, V. N., Mendes, R. & Melo, I. S. Functional congruence of rhizosphere microbial communities associated to leguminous tree from Brazilian semiarid region. *Environ. Microbiol. Rep.* **7**, 95–101 (2015).
46. Romagnoli, E. M. & Andreote, F. D. Rizosfera. in *Microbiologia do Solo* (eds. Cardoso, E. J. B. N. & Andreote, F. D.) 47–60 (ESALQ, 2016). doi:10.11606/97858481567
47. Barka, E. A. *et al.* Taxonomy, Physiology, and Natural Products of Actinobacteria. *Microbiol. Mol. Biol. Rev.* **80**, 1–43 (2016).

48. Anandan, R., Dharumadurai, D. & Manogaran, G. P. An Introduction to Actinobacteria. in *Actinobacteria: Basics and Biotechnological Applications* (ed. Dharumadurai, D.) 3–38 (IntechOpen, 2016). doi:10.5772/32009
49. El Euch, I. Z. *et al.* Bioactive secondary metabolites from new terrestrial Streptomyces sp. TN82 strain: Isolation, structure elucidation and biological activity. *Med. Chem. Res.* 1–8 (2018). doi:10.1007/s00044-017-2130-4
50. Adegbeye, M. F. & Babalola, O. O. Actinomycetes : a yet inexhaustive source of bioactive secondary metabolites. *Microb. Pathog. Strateg. Combat. them Sci. Technol. Educ.* 786–795 (2013).
51. Waksman, S. A. & Woodruff, H. B. Bacteriostatic and Bactericidal Substances Produced by a Soil Actinomyces. *Exp. Biol. Med.* **45**, 609–614 (1940).
52. Kominek, L. A. Biosynthesis of novobiocin by Streptomyces niveus. *Antimicrob. Agents Chemother.* **1**, 123–134 (1972).
53. Umezawa, H. *et al.* Production and isolation of a new antibiotic: kanamycin. *J Antibiot* **10**, 181–188 (1957).
54. Dulmage, H. T. The production of neomycin by Streptomyces fradiae in synthetic media. *Appl. Microbiol.* **1**, 103–6 (1953).
55. Margalith, P. & Beretta, G. Rifomycin. IX. Taxonomic study on Streptomyces mediterranei nov. sp. *Mycopathol. Mycol. Appl.* **13**, 321–330 (1960).
56. Schatz, A. & Waksman, S. A. Effect of Streptomycin and Other Antibiotic Substances upon Mycobacterium tuberculosis and Related Organisms,. *Exp. Biol. Med.* **57**, 244–248 (1944).
57. Linke, H. A. B., Mechlinski, W. & Schaffner, C. P. Production of amphotericin B-14c by Streptomyces nodosus fermentation, and preparation of the amphotericin B-14c-methyl-ester. *J. Antibiot. (Tokyo)*. **27**, 155–160 (1974).
58. Matsuoka, M., Yagishita, K. & Umezawa, H. Studies on the intermediate metabolism of chloramphenicol production. *Japanese J. Med. Sci. Biol.* **6**, 161–169 (1953).
59. Waksman, S. A. & Woodruff, H. B. Selective Antibiotic Action of Various Substances of Microbial Origin. *J. Bacteriol.* **44**, 373–84 (1942).

60. Omura, S. *et al.* Herbimycin, a new antibiotic produced by a strain of *Streptomyces*. *J. Antibiot. (Tokyo)*. **32**, 255–261 (1979).
61. Ōmura, S. & Crump, A. The life and times of ivermectin — a success story. *Nat. Rev. Microbiol.* **2**, 984–989 (2004).
62. Box, S. J., Cole, M. & Yeoman, G. H. Prasinons A and B: potent insecticides from *Streptomyces prasinus*. *Appl Microbiol* **26**, 699–704 (1973).
63. Burg, R. W. *et al.* Avermectins, New Family of Potent Anthelmintic Agents: Producing Organism and Fermentation. *Antimicrob. Agents Chemother.* **15**, 361–367 (1979).
64. Cerdeño, A. M., Bibb, M. J. & Challis, G. L. Analysis of the prodiginine biosynthesis gene cluster of *Streptomyces coelicolor* A3(2): new mechanisms for chain initiation and termination in modular multienzymes. *Chem. Biol.* **8**, 817–829 (2001).
65. Maskey, R. P. *et al.* Anti-cancer and Antibacterial Trioxacarcins with High Anti-malaria Activity from a Marine Streptomycete and their Absolute Stereochemistry. *J. Antibiot. (Tokyo)*. **57**, 771–779 (2004).
66. Vino, S. & Lokesh, K. R. Borrelidin: a promising anticancer agent from *Streptomyces* species. *Adv Biotech* 22–26 (2008).
67. Arcamone, F. *et al.* Adriamycin, 14-hydroxydaimomycin, a new antitumor antibiotic from *S. Peucetius* var.*caesius*. *Biotechnol. Bioeng.* **11**, 1101–1110 (1969).
68. Uyeda, M., Mizukami, M., Yokomizo, K. & Suzuki, K. Pentalenolactone I and Hygromycin A, Immunosuppressants produced by *Streptomyces filipinensis* and *Streptomyces hygroscopicus*. *Biosci. Biotechnol. Biochem.* **65**, 1252–1254 (2001).
69. Shapiro, S. *Regulation of Secondary Metabolism in Actinomycetes*. (CRC Press, 1989).
70. Hynes, R. K. & Boyetchko, S. M. Research initiatives in the art and science of biopesticide formulations. *38*, 845–849 (2006).
71. Chavan, D. V, Mulaje, S. S. & Mohalkar, R. Y. A Review on actinomycetes and their biotechnological applications. *Int. J. Pharm. Sci. Res.* **4**, 1730–1742 (2013).
72. Katz, L. & Baltz, R. H. Natural product discovery : past , present , and future. *J. Ind. Microbiol. Biotechnol.* (2016). doi:10.1007/s10295-015-1723-5
73. Chervin, J. *et al.* Targeted Dereplication of Microbial Natural Products by High-

- Resolution MS and Predicted LC Retention Time. *J. Nat. Prod.* **80**, 1370–1377 (2017).
- 74. Crevelin, E. J., Crotti, A. E. M., Zucchi, T. D., Melo, I. S. & Moraes, L. A. B. Dereplication of Streptomyces sp. AMC 23 polyether ionophore antibiotics by accurate-mass electrospray tandem mass spectrometry. *J. Mass Spectrom.* **49**, 1117–1126 (2014).
 - 75. Carnevale Neto, F. *et al.* Dereplication of Natural Products Using GC-TOF Mass Spectrometry: Improved Metabolite Identification by Spectral Deconvolution Ratio Analysis. *Front. Mol. Biosci.* **3**, 1–13 (2016).
 - 76. Niessen, W. M. A. State-of-the-art in liquid chromatography–mass spectrometry. *J. Chromatogr. A* **856**, 179–197 (1999).
 - 77. Bouslimani, A., Sanchez, L. M., Garg, N. & Dorrestein, P. C. Mass spectrometry of natural products: Current, emerging and future technologies. *Nat. Prod. Rep.* **31**, 718–729 (2014).
 - 78. Gilbert, J. R., Lewer, P., Duebelbeis, D. O. & Carr, A. W. The Central Role of Mass Spectrometry in Natural Products Discovery. *Integr. Strateg. drug Discov. using mass Spectrom.* 149–188 (2005). doi:10.1002/0471721034.ch6
 - 79. De Hoffmann, E. Tandem mass spectrometry: A primer. *J. Mass Spectrom.* **31**, 129–137 (1996).
 - 80. Gross, J. H. *Mass Spectrometry*. Springer (2011). doi:10.1201/9781420040340.axa
 - 81. Hoffmann, E. De & Stroobant, V. *Mass Spectrometry - Principles and Applications. Mass spectrometry reviews* **29**, (2007).
 - 82. States, U. & Substances, T. Ecological Effects Test Guidelines Aquatic Plant Toxicity Test Using Lemna spp ., Tiers I and II. *Environ. Prot.* (1996).
 - 83. Crevelin, E. J. *et al.* Isolation and characterization of phytotoxic compounds produced by streptomyces sp. AMC 23 from red mangrove (Rhizophora mangle). *Appl. Biochem. Biotechnol.* **171**, 1602–1616 (2013).
 - 84. Souza, D. T. *et al.* Saccharopolyspora spongiae sp. nov., a novel actinomycete isolated from the marine sponge scopalina ruetzleri (wiedenmayer, 1977). *Int. J. Syst. Evol. Microbiol.* **67**, 2019–2025 (2017).
 - 85. Lane, D. J. 16S/23S rRNA Sequencing. in *Nucleic Acid Techniques in Bacterial*

- Systematic* (eds. Stackebrandt, E. & Goodfellow, M.) 115–175 (John Wiley and Sons, 1991).
86. Wright, E. S., Yilmaz, L. S. & Noguera, D. R. DECIPHER, a Search-Based Approach to Chimera Identification for 16S rRNA Sequences. *Appl. Environ. Microbiol.* **78**, 717–725 (2012).
 87. Kim, O.-S. *et al.* Introducing EzTaxon-e: a prokaryotic 16S rRNA gene sequence database with phylotypes that represent uncultured species. *Int. J. Syst. Evol. Microbiol.* **62**, 716–721 (2012).
 88. Thompson, J. D., Higgins, D. G. & Gibson, T. J. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res.* **22**, 4673–80 (1994).
 89. Tamura, K. *et al.* MEGA5: Molecular Evolutionary Genetics Analysis Using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Methods. *Mol. Biol. Evol.* **28**, 2731–2739 (2011).
 90. Felsenstein, J. Evolutionary trees from DNA sequences: A maximum likelihood approach. *J. Mol. Evol.* **17**, 368–376 (1981).
 91. Fitch, W. M. Toward Defining the Course of Evolution: Minimum Change for a Specific Tree Topology. *Syst. Zool.* **20**, 406 (1971).
 92. SAITOU, N. & NEI, M. The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.* **4**, 406–425 (1987).
 93. Felsenstein, J. Confidence Limits On Phylogenies: An Approach Using The Bootstrap. *Evolution (N. Y.)* **39**, 783–791 (1985).
 94. Havlíková, L., Matysová, L., Hájková, R., Šatínský, D. & Solich, P. Advantages of pentafluorophenylpropyl stationary phase over conventional C18 stationary phase—Application to analysis of triamcinolone acetonide. *Talanta* **76**, 597–601 (2008).
 95. Corporation, W. MassLynx 4.1: Getting Started Guide. *ReVision* 94 (2005).
 96. Vechia, J. F. Della, Cruz, C., Silva, A. F., Cerveira Jr., W. R. & Garlich, N. Macrophyte Bioassay Applications for Monitoring Pesticides in the Aquatic Environment. *Planta Daninha* **34**, 597–603 (2016).

97. Wang, W. Literature review on duckweed toxicity testing. *Environ. Res.* **52**, 7–22 (1990).
98. Brain, R. A. & Solomon, K. R. PROTOCOL A protocol for conducting 7-day daily renewal tests with *Lemna gibba*. **2**, (2007).
99. Zache, S. Uso de tecnologias diminui infestação de buva na lavoura de soja. *Embrapa* (2011). Available at: <https://www.embrapa.br/busca-de-noticias/-/noticia/18153589/uso-de-tecnologias-diminui-infestacao-de-buva-na-lavoura-de-soja->.
100. Holmes, J. L. & Benoit, F. The mass spectra of carboxylic acids—III: The structures of molecular and fragment ions in benzoic acid and related molecules. *Org. Mass Spectrom.* **4**, 97–107 (1970).
101. Dictionary of Natural Products. *Chapman & Hall Chemical Dictionaries* Available at: <http://dnp.chemnetbase.com>.
102. Gao, Y. *et al.* Comparative phytotoxicity of usnic acid, salicylic acid, cinnamic acid and benzoic acid on photosynthetic apparatus of *Chlamydomonas reinhardtii*. *Plant Physiol. Biochem.* **128**, 1–12 (2018).
103. War, A. R., Paulraj, M. G., War, M. Y. & Ignacimuthu, S. Role of salicylic acid in induction of plant defense system in chickpea (*Cicer arietinum* L.). *Plant Signal. Behav.* **6**, 1787–1792 (2011).
104. Zucchi, T. D., Almeida, L. G., Moraes, L. A. B. & Cônsoli, F. L. Albocycline, the main bioactive compound from *Propionicimonas* sp. ENT-18 against *Sclerotinia sclerotiorum*. *Ind. Crops Prod.* **52**, 264–268 (2014).
105. Taddei, A. & Zeeck, A. Biosynthesis of Albocycline: Origin of the Carbon Skeleton. *J. Antibiot. (Tokyo)*. **50**, 1–3 (1997).
106. Chan, Y. A., Podevels, A. M., Kevany, B. M. & Thomas, M. G. Biosynthesis of polyketide synthase extender units. *Nat. Prod. Rep.* **26**, 90–114 (2009).
107. Harborne, J. B. Dictionary of Natural Products. *Phytochemistry* **38**, 279 (1995).
108. Reusser, F. Mode of Action of Albocycline , an Inhibitor of Nicotinate Biosynthesis. **100**, 11–13 (1969).
109. Taguchi, H., Nishitani, H., Okumura, K., Shimabayashi, Y. & Iwai, K. Biosynthesis and Metabolism of Trigonelline in *Lemna paucicostata* 151. *Agric. Biol. Chem.* **53**, 2867–

2871 (1989).

110. Sasamoto, H. & Ashihara, H. Effect of nicotinic acid, nicotinamide and trigonelline on the proliferation of lettuce cells derived from protoplasts. *Phytochem. Lett.* **7**, 38–41 (2014).
111. Mizuno, K. *et al.* Conversion of nicotinic acid to trigonelline is catalyzed by N-methyltransferase belonged to motif B??? methyltransferase family in Coffea arabica. *Biochem. Biophys. Res. Commun.* **452**, 1060–1066 (2014).
112. Disaccharide. *Encyclopædia Britannica, inc.* (2015). Available at: <https://www.britannica.com/science/disaccharide>.
113. Monosaccharide. *Encyclopædia Britannica, inc.* (2018). Available at: <https://www.britannica.com/science/monosaccharide>.
114. Kim, H. J. *et al.* A single module type I polyketide synthase directs de Novo macrolactone biogenesis during galbonolide biosynthesis in *Streptomyces galbus*. *J. Biol. Chem.* **289**, 34557–34568 (2014).
115. Nguyen, H. C., Darbon, E., Thai, R., Pernodet, J. L. & Lautru, S. Post-PKS tailoring steps of the spiramycin macrolactone ring in *streptomyces ambofaciens*. *Antimicrob. Agents Chemother.* **57**, 3836–3842 (2013).
116. Tang, L., Fu, H., Betlach, M. C. & McDaniel, R. Elucidating the mechanism of chain termination switching in the picromycin/methymycin polyketide synthase. *Chem. Biol.* **6**, 553–558 (1999).
117. Mitchison, J. M. *The Biology of the Cell Cycle*. (Cambridge University Press, 1972).
118. Ruiz, B. *et al.* Production of microbial secondary metabolites: Regulation by the carbon source. *Crit. Rev. Microbiol.* **36**, 146–167 (2010).
119. Pu, X. *et al.* A new cyclododeca[d]oxazole derivative from *Streptomyces* spp. CIBYL1. *Nat. Prod. Res.* **27**, 603–608 (2013).
120. Hwang, B. K. *et al.* Isolation and In Vivo and In Vitro Antifungal Activity of Phenylacetic Acid and Sodium Phenylacetate from *Streptomyces humidus* Isolation and In Vivo and In Vitro Antifungal Activity of Phenylacetic Acid and Sodium Phenylacetate from *Streptomyces humidus*. *Appl. Environ. Microbiol.* **67**, 3739–3745 (2001).

121. Sri Andayani, D. G., Sukandar, E. Y., Sukandar, U. & Ketut Adnyana, I. Isolation, identification of phenylacetic acid from *Streptomyces galbus* TP2 strain and its toxicity. *Int. J. Pharm. Pharm. Sci.* **6**, 643–646 (2014).
122. Sajid, I., Shaaban, K. A. & Hasnain, S. Identification, Isolation and optimization of antifungal metabolites from the *Streptomyces malachitofuscus* CTF9. *Brazilian J. Microbiol.* **42**, 592–604 (2011).
123. Yang, S. W. & Cordell, G. A. Metabolism studies of indole derivatives using a staurosporine producer, *Streptomyces staurosporeus*. *J. Nat. Prod.* **60**, 44–48 (1997).
124. Li, Z. H. *et al.* Biological activity and quantification of potential autotoxins from *Picea schrenkiana* leaves. *Allelopath. J.* **27**, 245–262 (2011).
125. Anaya, A. L., Hernandez-Bautista, B. E., Jimenez-Estrada, M. & Velasco-Ibarra, L. Phenylacetic acid as a phytotoxic compound of corn pollen. *J. Chem. Ecol.* **18**, 897–905 (1992).
126. Bartz, F. E., Glassbrook, N. J., Danehower, D. a. & Cubeta, M. a. Elucidating the role of the phenylacetic acid metabolic complex in the pathogenic activity of *Rhizoctonia solani* anastomosis group 3. *Mycologia* **104**, 793–803 (2012).
127. Cardellina, J. H., Nigh, D. & Van Wagenen, B. C. Plant Growth Regulatory Indoles from the Sponges *Dysidea Etheria* and *Ulosa Ruetzleri*. *J. Nat. Prod.* **49**, 1065–1067 (1986).
128. Kaufman, D. D. Biodegradation and persistence of several organophosphate pesticide combinations. *Soil Biol. Biochem.* **9**, 49–57 (1966).
129. Wang, C. *et al.* Enantioselective Phytotoxicity and the Relative Mechanism of Current Chiral Herbicides. *Curr. Protein Pept. Sci.* **18**, 15–21 (2016).