

Universidade de São Paulo
Instituto de Ciências Biomédicas

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**Investigação sobre a ocorrência de reprogramação fetal
no desenvolvimento do pâncreas endócrino em modelo
animal de diabetes *mellitus* tipo 1**

Dissertação apresentada ao Programa de
Pós-graduação em Biologia de Sistemas
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Universidade de São Paulo, para
obtenção do Título de Mestre em
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Área de concentração: Biologia de Sistemas

Orientador: Profa. Dra. Telma Maria Tenório Zorn

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RESUMO

DIAS, C. Investigação sobre a ocorrência de reprogramação fetal no desenvolvimento do pâncreas endócrino em modelo animal de diabetes *mellitus* tipo 1 [dissertação (Mestrado em Biologia de Sistemas)]. São Paulo: Instituto de Ciências Biomédicas; Universidade de São Paulo, São Paulo; 2019.

Várias evidências, incluindo as originadas de estudos anteriores do LBR&MEC, sugerem que condições adversas durante o desenvolvimento intrauterino promovam alterações moleculares e estruturais em órgãos e sistemas vitais podendo comprometer o seu funcionamento no indivíduo adulto. A hiperglicemia é um fator que influencia negativamente o desenvolvimento fetal, modificando processos biológicos importantes, como o padrão de síntese e deposição dos componentes da matriz extracelular (MEC). A MEC participa diretamente do processo de ramificação e morfogênese do pâncreas, e pouco é conhecido a respeito dos efeitos da hiperglicemia materna sobre a MEC desse órgão durante seu desenvolvimento. Investigamos por meio de imuno-histoquímica como a hiperglicemia materna severa modifica a distribuição de panlaminina, das cadeias $\alpha 1$ e $\gamma 1$ das lamininas e da integrina $\alpha 3$, moléculas da MEC que desempenham um papel chave na diferenciação do pâncreas endócrino. Avaliamos o perfil proliferativo das células presentes nas ilhotas ainda, a distribuição das células α e β por meio da marcação de glucagon e insulina no pâncreas de fetos de 19 dias. Analisamos por RT-qPCR a expressão dos fatores de transcrição *Pdx1* e *Pax4* que controlam o desenvolvimento e diferenciação das células β pancreáticas. O modelo utilizado foi o de gestação complicada por diabetes *mellitus* tipo 1 (DM1), desenvolvido por nosso grupo, quimicamente induzido por aloxana sem tratamento de reposição insulínica, em camundongos. Observamos que a marcação de panlaminina e das cadeias $\alpha 1$ e $\gamma 1$ das lamininas é mais fraca no pâncreas endócrino dos fetos de mães hiperglicêmicas, quando comparado ao grupo controle. Por outro lado, vimos um aumento na deposição da integrina $\alpha 3$ na membrana basal das ilhotas pancreáticas dos fetos gerados sob condições de hiperglicemia materna. O índice proliferativo das células endócrinas, observado por imuno-histoquímica para PCNA, também é menor nesse grupo. Observamos um aumento da área de ilhotas fetais imunomarcadas para a insulina, indicando aumento na massa de células β nessas ilhotas, enquanto que a área imunomarcada para glucagon estava com marcação menos intensa no grupo experimental comparado ao controle. Identificamos que a expressão relativa de *Pdx1* é menor no pâncreas do grupo experimental comparado a expressão nos animais do grupo controle, enquanto a expressão de *Pax4* está aumentada. Concluímos por meio de nossas abordagens histoquímicas que a hiperglicemia materna altera a morfogênese do pâncreas endócrino fetal modificando o

padrão de deposição de moléculas da membrana basal peri-ilhotas, promovendo uma diminuição da atividade proliferativa das células endócrinas, associada a alterações na expressão de fatores de crescimento importantes para o estado diferenciado e proliferativo das células β . Essas células apresentam aumento da massa funcional identificada pelo aumento da deposição de insulina no tecido pancreático.

PALAVRAS-CHAVE: Diabetes *Mellitus* tipo 1. Matriz Extracelular. Reprogramação Fetal. Histologia. Biologia do Desenvolvimento.

ABSTRACT

DIAS, C. Research about occurrence of fetal reprogramming in the development of endocrine pancreas in animal model of type 1 diabetes *mellitus* [dissertation (Master's degree in System Biology)], Institute of Biomedical Sciences; University of São Paulo, São Paulo; 2019.

Previous studies from our lab and others have shown that adverse conditions during intrauterine development promotes molecular and structural changes in vital organs and systems which may alter on their function in the adult individuals. Hyperglycemia impacts on fetal development by modifying important biological processes, such as the pattern of synthesis and deposition of extracellular matrix (ECM) components. ECM cooperates in pancreatic branching and morphogenesis and little is known about the effect of maternal hyperglycemia on the pancreas' ECM during development. We investigate through immunohistochemistry, how severe maternal hyperglycemia modifies the distribution of panlaminin, laminins α 1 and γ 1 chains and integrin α 3, ECM molecules that play a key role in the differentiation of the endocrine pancreas. We evaluate the proliferative index of islet cells and, α and β cells distribution, by glucagon and insulin fetal (E19.0) pancreas staining. We analyzed by RT-qPCR the expression of the *Pdx1* and *Pax4* transcription factors that control the development and differentiation of pancreatic β cells. The model used was created by our group, a pregnancy model complicated by type 1 diabetes *mellitus* (T1D) chemically induced by alloxan without treatment of insulin replacement, in mice. We observed that the labeling of panlaminin and laminins α 1 and γ 1 chains is weaker in the endocrine pancreas of the fetuses from hyperglycemic mothers. On the other hand, integrin α 3 deposition increased in the basement membrane of the pancreatic islets of the fetuses generated under maternal hyperglycemia. Immunohistochemistry for PCNA showed lower proliferative index of endocrine cells. There was an increase in the area of immunolabeled fetal islets indicating an increase in β -cell mass in these islets; whereas the glucagon-immunolabeled area was smaller in the experimental group compared to the control group. The relative expression of *Pdx1* was lower in the pancreas from the experimental group, and the *Pax4* expression was increased. We conclude from our histochemical approaches that maternal hyperglycemia alters fetal endocrine pancreas morphogenesis by modifying the pattern of peri-islet basement membrane molecules, promoting a decrease in endocrine cell proliferative activity associated with changes in the expression of important growth factors for the differentiated and proliferative state of the β -cells. These cells have increased functional mass identified by increased insulin deposition in pancreatic tissue.

KEY WORDS: Type 1 Diabetes *Mellitus*. Extracellular Matrix. Fetal Reprogramming. Histology. Development Biology.

1 INTRODUÇÃO

O pâncreas é uma glândula mista essencial para o metabolismo de nutrientes. Possui uma porção exócrina, composta por acinos que secretam enzimas digestivas e por uma porção endócrina, formada pelas ilhotas pancreáticas que abrigam as células α , β , γ , ϵ e PP responsáveis pela homeostase da glicose (2). Em camundongos, o desenvolvimento pancreático torna-se morfologicamente evidente em torno do 9º dia embrionário, quando a superfície epitelial começa a se formar, alongando-se em ramos. A Matriz extracelular (MEC) participa diretamente do processo de morfogênese do pâncreas promovendo adesão das células precursoras pancreáticas, e pela interação de seus componentes com vias de sinalização importantes para a diferenciação e proliferação celular (2).

O Retardo de crescimento intrauterino é uma alteração frequente em modelos murinos de gestação complicada por diabetes. De modo geral, os fetos apresentam baixo peso corpóreo e menor volume pancreático, embora a porcentagem de tecido endócrino esteja aumentada, o que pode ser considerado um quadro de insuficiência pancreática (3).

Esses fenômenos adaptativos são considerados mecanismos de reprogramação fetal (4). A teoria da reprogramação do desenvolvimento embrionário e fetal foi postulada por David J. Baker no início dos anos 90 (4). Por meio de um estudo *coorte*, Baker mostrou que filhos de mães malnutridas no período perinatal, têm maior predisposição a desenvolver doenças cardiovasculares e metabólicas ao longo da vida adulta, devido a mecanismos adaptativos adotados durante a vida intrauterina para garantir sua sobrevivência (4). Barker também correlacionou o surgimento de hiperinsulinemia e resistência à insulina em indivíduos adultos ao diabetes materno, indicando que essa doença pode estimular a reprogramação nos fetos (4).

O Diabetes *mellitus* (DM) é caracterizado pela hiperglicemia ocasionada por defeitos na secreção e/ou ação da insulina nas células alvo (3). São classificados três tipos principais, sendo eles: Tipo 1 (DM1), Tipo 2 (DM2) e Diabetes Gestacional (DMG) (5)(6)(7). O desenvolvimento e a progressão dos três tipos de diabetes apresentam componentes genéticos, porém, fatores ambientais também estão intimamente relacionados à disfunção e/ou morte das células β pancreáticas produtoras de insulina (7). O DM1 tem baixa prevalência, mas aproximadamente 86.000 indivíduos desenvolvem essa doença a cada ano mundialmente (7). Em decorrência da hiperglicemia crônica podem ocorrer sérias complicações, dentre elas desordens reprodutivas e comprometimento da organogênese em fetos gerados por mães hiperglicêmicas (7)(8). A MEC participa diretamente da morfogênese, ramificação e do comportamento celular do pâncreas em desenvolvimento (9). As lamininas 111, 211, 221, 411

e 421 presentes na MEC pancreática de roedores são essenciais para a adesão e proliferação, como também para a secreção de insulina pelas células β (10). Já a integrina $\alpha 3$ atua como receptor para lamininas e desempenha um papel essencial no desenvolvimento das células β regulando sua sobrevivência e função (11).

Dada a importância da MEC para o desenvolvimento e função adequados do pâncreas (12) e sabendo que o DM1 é uma condição que pode influenciar o desenvolvimento fetal alterando os componentes de MEC (12)(13)(14), hipotetizamos que a hiperglicemia materna poderia modificar a deposição das cadeias $\alpha 1$ e $\gamma 1$ das principais lamininas presentes no pâncreas fetal, assim como da integrina $\alpha 3$ no pâncreas endócrino dos fetos, e ter impacto na expressão de fatores de transcrição importantes para a diferenciação e proliferação de células β pancreáticas fetais, como *Pdx1* e *Pax4*.

Nossa hipótese foi investigada em pâncreas de fetos de camundongos de 19 dias provindos de um modelo de gestação complicada por hiperglicemia, tipo DM1 de curto prazo (30-50 dias) quimicamente induzido sem reposição insulínica (13)(15)(16).

7 CONCLUSÃO

Concluímos que a hiperglicemia materna 30-50 dias promove alterações no desenvolvimento do pâncreas endócrino fetal de camundongos, modificando diferencialmente o padrão de deposição de moléculas da membrana basal das ilhotas, associado a diminuição da atividade proliferativa das células endócrinas com aumento da massa funcional de células β . Acreditamos que o conjunto dessas alterações seja um indício de reprogramação das células do pâncreas endócrino, que levam a modificações na morfologia do órgão e de seu funcionamento. O estudo aprofundado desses achados iniciais são necessários para comprovar a reprogramação por parte dessas células e os efeitos a longo prazo.

REFERÊNCIAS*

1. Moraes V de, Toquinho. Aquarela. 1983.
2. Shih HP, Wang A, Sander M. Pancreas Organogenesis: From Lineage Determination to Morphogenesis. *Annu Rev Cell Dev Biol*. 2013;29(1):81–105.
3. Aerts L, Vercruyse L, Van Assche FA. The endocrine pancreas in virgin and pregnant offspring of diabetic pregnant rats. *Diabetes Res Clin Pract*. 1997;38(1):9–19.
4. Hales CN, Barker DJP. Type 2 (non-insulin-dependent) diabetes mellitus: the thrifty phenotype hypothesis. *Int J Epidemiol*. 2013;42(5):1215–22.
5. Federation ID. IDF Atlas. 2015. 144 p
6. Steck AK, Rewers MJ. Genetics of type 1 diabetes. *Clin Chem*. 2011;57(2):176–85.
7. Gallen I. Type 1 Diabetes: Clinical Management of the Athlete. *Clin Chem*. 2012;367(9911):194.
8. Stirban A, Peter R, Tschoepe D. Complications of Type 1 Diabetes : New Molecular Findings. 2008;328–51.
9. Beattie GM, Rubins JS, Mally MI, Otonkoski T, Hayek A. Regulation of proliferation and differentiation of human fetal pancreatic islet cells by extracellular matrix, hepatocyte growth factor, and cell-cell contact. *Diabetes*. 1996;45:1223–8.
10. Jiang FX, Cram DS, DeAizpurua HJ, Harrison LC. Laminin-1 promotes differentiation of fetal mouse pancreatic beta-cells. *Diabetes*. 1999;48(4):722–30.
11. Krishnamurthy M, Li J, Fellows GF, Rosenberg L, Goodyer CG, Wang R. Integrin α 3, but not β 1, regulates islet cell survival and function via PI3K/Akt signaling pathways. *Endocrinology*. 2011;152(2):424–35.
12. Jennings RE, Berry AA, Strutt JP, Gerrard DT, Hanley NA. Human pancreas development. *Development*. 2015;
13. Favaro RR, Salgado RM, Covarrubias AC, Bruni F, Lima C, Fortes ZB, et al. Long-term type 1 diabetes impairs decidualization and extracellular matrix remodeling during early embryonic development in mice. *Placenta*. 2013;34(12):1128–35.
14. Sanches JC, Favaro RR, Barrence FC, Bevilacqua E, Fortes ZB, Zorn TMT. Distinct effects of short- and long-term type 1 diabetes to the placental extracellular matrix and fetal development in mice. *Placenta*. 2017;53:1–7.
15. Sanches JC, Favaro RR, Barrence FC, Bevilacqua E, Fortes ZB, Zorn TMT. Distinct effects of short- and long-term type 1 diabetes to the placental extracellular matrix and fetal development in mice. *Placenta*. 2017;53:1–7.
16. Favaro RR, Salgado RM, Raspantini PR, Fortes ZB, Zorn TMT. Effects of long-term diabetes on the structure and cell proliferation of the myometrium in the early

- pregnancy of mice. *Int J Exp Pathol.* 2010;91(5):426–35.
17. Dean L, McEntyre J. The Genetic Landscape of Diabetes. *Genet Landsc Diabetes.* 2004;(Md):1–20.
 18. Wilcox G. Insulin and Insulin. 2005;26(May):19–39.
 19. Brownlee M. Biochemistry and molecular cell biology of diabetic complications. *Nature.* 2001;414(December):813–20.
 20. Brownlee M, Cerami A. The biochemistry of the complications of diabetes mellitus. *Annu Rev Biochem.* 1981;50(1):385–432.
 21. Lomas J, Anderson GM, Dominick-Pierre KA, Vayda E, Enkin MW, Hannah WJ. Type I diabetes mellitus. A chronic autoimmune. *N Engl J Med.* 1989;321(19):1306–11.
 22. Wild S, Roglic G, Green A, Sicree R, King H. Estimates for the year 2000 and projections for 2030. *Diabetes Care.* 2004;27(5):1047–53.
 23. Ministério da Saúde. Cadernos de Atenção Básica Diabetes Mellitus. 2006.
 24. Federation ID. IDF Atlas. 2017. 144 p.
 25. You WP, Henneberg M. Type 1 diabetes prevalence increasing globally and regionally: The role of natural selection and life expectancy at birth. *BMJ Open Diabetes Res Care.* 2016;4(1):1–7.
 26. Hrabovsky M. The Epidemiology of Type 1 Diabetes Mellitus. *Intech open.* 2018;2:64.
 27. Shulman RM, Daneman D. Type 1 diabetes mellitus in childhood. *Medicine (Baltimore).* 2010;38(12):679–85.
 28. Devendra D, Liu E, Eisenbarth GS. Clinical review Type 1 diabetes: recent developments. *Br Med J.* 2004;328(1):750–4.
 29. Jun HS, Yoon JW. The role of viruses in Type I diabetes: Two distinct cellular and molecular pathogenic mechanisms of virus-induced diabetes in animals. *Diabetologia.* 2001;44(3):271–85.
 30. Hviid A, Stellfeld M, Wohlfahrt J, Melbye M. Childhood Vaccination and Type 1 Diabetes. *N Engl J Med.* 2004;350(14):1398–404.
 31. Islam ST, Srinivasan S, Craig ME. Environmental determinants of type 1 diabetes: A role for overweight and insulin resistance. *J Paediatr Child Health.* 2014;50(11):874–9.
 32. Acharjee S, Ghosh B, Al-Dhubiab BE, Nair AB. Understanding type 1 diabetes: Etiology and models. *Can J Diabetes.* 2013;37(4):269–76.
 33. Bruno G, Runzo C, Cavallo-Perin P, Merletti F, Rivetti M, Pinach S, et al. Incidence of type 1 and type 2 diabetes in adults aged 30–49 years: The population-based registry in

- the province of Turin, Italy. *Diabetes Care*. 2005;28(11):2613–9.
34. Lecture KCB. Insulin action, diabetogenes, and the cause of type II diabetes. *Diabetes*. Diabetes. 1994;43:1066–85.
 35. RP1. R. Antagonist: diabetes and insulin resistance—philosophy, science, and the multiplier hypothesis. *J Lab Clin Med*. 1995;125(5):560–4.
 36. Ye J. Mechanisms of insulin resistance in obesity. *Front Med China*. 2013;7(1):14–24.
 37. Spaight C, Gross J, Horsch A, Puder JJ. Gestational diabetes mellitus. *Endocr Dev*. 2016;31:163–78.
 38. Benhalima K, Devlieger R, Van Assché A. Screening and management of gestational diabetes. *Best Pract Res Clin Obstet Gynaecol*. 2015;29(3):339–49.
 39. Wei YM, Yang HX. Diagnosis and management of gestational diabetes mellitus in China. *Chin Med J (Engl)*. 2012;125(7):1206–8.
 40. Casson I, Clarke C, Howard C, McKendrick O, Pennycook S, Pharoah P, et al. Outcomes of pregnancy in insulin dependent diabetic women: results of a five year population cohort study. *Bmj*. 1997;315(7103):275–8.
 41. M Makhseed, V. M. Musini, M. A. Ahmed J A-H. Placental pathology in relation to the White's classification of diabetes mellitus. *Archives of Gynecology and Obstetrics*. 2002;136–40.
 42. Oh W, Gelardi NL, Cha CJ. Maternal hyperglycemia in pregnant rats: Its effect on growth and carbohydrate metabolism in the offspring. *Metabolism*. 1988;37(12):1146–51.
 43. Marco LJ, McCloskey K, Vuillermin PJ, Burgner D, Said J, Ponsonby A. Cardiovascular Disease Risk in the Offspring of Diabetic Women : The Impact of the Intrauterine Environment. *Exp Diabetes Res*. 2012;2012.
 44. Akcakus M, Koklu E, Baykan A, Yikilmaz A, Coskun A, Gunes T, et al. Macrosomic newborns of diabetic mothers are associated with increased aortic intima-media thickness and lipid concentrations. *Horm Res*. 2007;67(6):277–83.
 45. Mason RM. Extracellular Matrix Metabolism in Diabetic Nephropathy. *J Am Soc Nephrol*. 2004;14(5):1358–73.
 46. Bruning JC, Brüning JC, Gautam D, Burks DJ, Gillette J, Schubert M, et al. Role of brain insulin receptor in control of body weight and reproduction. *Science* (80-). 2000;289(5487):2122–5.
 47. Correia-Santos AM, Vicente GC, Suzuki A, Pereira AD, dos Anjos JS, Lenzi-Almeida KC, et al. Maternal use of flaxseed oil during pregnancy and lactation prevents morphological alterations in pancreas of female offspring from rat dams with experimental diabetes. *Int J Exp Pathol*. 2015;96(2):94–102.

48. Mayer C, Joseph KS. Fetal growth: A review of terms, concepts and issues relevant to obstetrics. *Ultrasound Obstet Gynecol.* 2013;41(2):136–45.
49. Barker DJ, Hales CN, Fall CH, Osmond C, Phipps K CP. Type 2 (non-insulin-dependent) diabetes mellitus, hypertension and hyperlipidaemia (syndrome X): relation to reduced fetal growth. *Diabetologia.* 1993;36:5.
50. Tuovinen S, Aalto-Viljakainen T, Eriksson JG, Kajantie E, Lahti J, Pesonen AK, et al. Maternal hypertensive disorders during pregnancy: Adaptive functioning and psychiatric and psychological problems of the older offspring. *BJOG An Int J Obstet Gynaecol.* 2014;121(12):1482–91.
51. Himmelmann K, Himmelmann A, Niklasson A SA. Hypertension in pregnancy and size at birth. *Blood Press.* 1996;5:6.
52. Gray PH, O'Callaghan MJ, Harvey JM, Burke CJ PD. Placental pathology and neurodevelopment of the infant with intrauterine growth restriction. *Dev Med Child Neurol.* 1999;41:16–20.
53. Barker DJP. Mothers, Babies and Health in Later Life. Churchill Livingst. 1998;2.
54. Moritz KM, Evans RG, Hoppe CC, Cullen-McEwen LA, Bertram JF, Dowling J, et al. Combined prenatal and postnatal protein restriction influences adult kidney structure, function, and arterial pressure. *Am J Physiol Integr Comp Physiol.* 2006;292(1):R462–9.
55. Stuart RO, Bush KT, Nigam SK. Changes in global gene expression patterns during development and maturation of the rat kidney. *Proc Natl Acad Sci.* 2002;98(10):5649–54.
56. Woodall SM, Johnston BM, Breier BH GP. Chronic maternal undernutrition in the rat leads to delayed postnatal growth and elevated blood pressure of offspring. *Pediatr Res.* 1996;40:438–43.
57. Longnecker D. Anatomy and Histology of the Pancreas. *Am Pancreat Assoc.* 2017;1–26.
58. Dale BE. Anatomy of the pancreas.pdf. *The Pancreas: Biology, Pathobiology, and Disease.* 1993. p. 1–8.
59. Abraham L. Kierszenbaum, M.D. PD. *Histologia e Biologia Celular Uma Introdução à Patologia.* 2nd ed. Ltda EE, editor. Rio de Janeiro - RJ - Brasil; 2008. 495-498 p.
60. In't Veld, P., & Smeets S. Microscopic Anatomy of the Human Islet of Langerhans. *Islets of Langerhans.* 2014;2:1–18.
61. Carvalho CPF, Barbosa HCL, Britan A, Santos-Silva JCR, Boschero AC, Meda P, et al. Beta cell coupling and connexin expression change during the functional maturation of rat pancreatic islets. *Diabetologia.* 2010;53(7):1428–37.

62. Wong HY, Ahrén B, Lips CJM, Höppener JWM, Sundler F. Postnatally disturbed pancreatic islet cell distribution in human islet amyloid polypeptide transgenic mice. *Regul Pept.* 2003;113(1–3):89–94.
63. Hong EG, Noh HL, Lee SK, Chung YS, Lee KW, Kim HM. Insulin and glucagon secretions, and morphological change of pancreatic islets in OLETF rats, a model of type 2 diabetes mellitus. *J Korean Med Sci.* 2002;17(1):34–40.
64. Bosco D, Armanet M, Morel P, Niclauss N, Sgroi A, Muller YD, et al. Unique arrangement of α - and β -cells in human islets of Langerhans. *Diabetes.* 2010;59(5):1202–10.
65. Martin F, Soria B. Glucose-induced in single human. *Imaging.* 1996;20:409–14.
66. Brissova M, Fowler MJ, Nicholson WE, Chu A, Hirshberg B, Harlan DM, et al. Assessment of human pancreatic islet architecture and composition by laser scanning confocal microscopy. *J Histochem Cytochem.* 2005;53(9):1087–97.
67. Fowler JL, Lee SSY, Wesner ZC, Olehnik SK, Kron SJ, Hara M. Three-Dimensional Analysis of the Human Pancreas. *Endocrinology.* 2018;159(3):1393–400.
68. El-Gohary Y, Sims-Lucas S, Lath N, Tulachan S, Guo P, Xiao X, et al. Three-Dimensional Analysis of the Islet Vasculature. *Anat Rec.* 2012;295(9):1473–81.
69. Konstantinova I, Lammert E. Microvascular development: Learning from pancreatic islets. *BioEssays.* 2004;26(10):1069–75.
70. Lara R, Nyman, 1 K, Sam Wells, 2 W, Steve Head, 2 Michael McCaughey 2, Eric Ford, 1 Marcela Brissova, 1 David W, Piston 2 and Alvin C, Powers1. Real time, multidimensional in vivo imaging used to investigate blood flow in mouse pancreatic islets. *J Clin Invest.* 2008;118(11):3790–7.
71. Nyman LR, Ford E, Powers AC, Piston DW. Glucose-dependent blood flow dynamics in murine pancreatic islets in vivo. *Am J Physiol Metab.* 2010;298(4):E807–14.
72. Ahrén B. Autonomic regulation of islet hormone secretion - Implications for health and disease. *Diabetologia.* 2000;43(4):393–410.
73. Havel, P. J., & Ahren B. Activation of Autonomic Nerves and the Adrenal Medulla Contributes to Increased Glucagon Secretion During Moderate Insulin-Induced Hypoglycemia in Women. *Diabetes.* 1997;5:801–7.
74. Gilliam LK, Palmer JP, Jr GJT. Tyramine-mediated activation of sympathetic nerves inhibits insulin secretion in humans. *2014;92(10):4035–8.*
75. van Deijnen JHM, Hulstaert CE, Wolters GHJ, van Schilfgaarde R. Significance of the peri-insular extracellular matrix for islet isolation from the pancreas of rat, dog, pig, and man. *Cell Tissue Res.* 1992;267(1):139–46.

76. Ziolkowski AF, Parish CR, Sado Y, Ninomiya Y, Simeonovic CJ, Rodgers RJ. Molecular composition of the peri-islet basement membrane in NOD mice : a barrier against destructive insulitis. 2008;1680–8.
77. Arous C, Wehrle-Haller B. Role and impact of the extracellular matrix on integrin-mediated pancreatic β -cell functions. *Biol Cell*. 2017;109(6):223–37.
78. Korpos E, Simeonovic CJ, Wight TN, Bogdani M, Sorokin L, Parish CR. Extracellular Matrix Components in the Pathogenesis of Type 1 Diabetes. *Curr Diab Rep*. 2014;14(12).
79. Rahier J, Wallon J, Henquin JC. Cell populations in the endocrine pancreas of human neonates and infants. *Diabetologia*. 1981;20(5):540–6.
80. Hara M, Miller K, Wojcik P, Kim A, Jo J, Kilimnik G. Islet architecture: A comparative study. *Islets*. 2010;1(2):129–36.
81. Steiner DJ, Kim A, Miller K, Hara M. Pancreatic islet plasticity: Interspecies comparison of islet architecture and composition. *Islets*. 2010;2(3):135–45.
82. Li W-C, Ouziel-Yahalom L, Sharma A, Weir GC, Bonner-Weir S, Guo L. -Cell Growth and Regeneration: Replication Is Only Part of the Story. *Diabetes*. 2010;59(10):2340–8.
83. Jung H. On the origin of the Glagolitic alphabet. *Proc Scr*. 2012;10(8–10):89–120.
84. Häring H-U, Aichler M, Irmler M, Wang-Sattler R, Leitzinger C, Gerdes J, et al. Identification of proliferative and mature β -cells in the islets of Langerhans. *Nature*. 2016;535(7612):430–4.
85. H E. Developmental biology of the pancreas. *Diabetes* 50. Suppl 1. 2001;1580:S5–S9.
86. Georgia S, Bhushan A. B Cell Replication Is the Primary Mechanism for Maintaining Postnatal B Cell Mass. *J Clin Invest*. 2004;114(7):963–8.
87. Dor Y, Brown J, Martinez OI, Melton DA. Adult pancreatic. *Nature*. 2004;(model 3):41–6.
88. Zajicek G , Arber N , Schwartz-Arad D AI. Streaming pancreas: islet cell kinetics. *Diabetes Res*. 1990;13:121–5.
89. Kodama S, Kühtreiber W, Fujimura S, Dale EA, Faustman DL. Islet Regeneration during the Reversal of Autoimmune Diabetes in NOD Mice. *Science* (80-). 2003;302(5648):1223–7.
90. Liew CG, Andrews PW. Stem cell therapy to treat diabetes mellitus. *Rev Diabet Stud*. 2008;5(4):203–19.
91. Gradwohl G, Dierich A, LeMeur M, Guillemot F. Neurogenin3 Is Required for the Development of the Four Endocrine Cell Lineages of the Pancreas. *Proc Natl Acad Sci*

U S A. 2000;97(4):1607–11.

92. Tsai M-J, Huang H-P, Naya FJ, Leiter AB, Qiu Y, DeMayo FJ, et al. Diabetes, defective pancreatic morphogenesis, and abnormal enteroendocrine differentiation in BETA2/NeuroD-deficient mice. *Genes Dev.* 2008;11(18):2323–34.
93. Thorens B, Guillam MT, Beermann F, Burcelin R, Jaquet M. Transgenic reexpression of GLUT1 or GLUT2 in pancreatic β cells rescues GLUT2-null mice from early death and restores normal glucose-stimulated insulin secretion. *J Biol Chem.* 2000;275(31):23751–8.
94. Mansouri A, St-Onge L, Gruss P. Role of Pax genes in endoderm-derived organs. *Trends Endocrinol Metab.* 1999;10(4):164–7.
95. Kallman F, Sciences B. Structure Pancreatic of Differentiating Culture Mouse in Transfilter. *J Cell Biol.* 1960;
96. Offield MF, Jetton TL, Labosky PA, Ray M, Stein RW, Magnuson MA, et al. PDX-1 is required for pancreatic outgrowth and differentiation of the rostral duodenum. *1996;995:983–95.*
97. Fujitani Y. Transcriptional regulation of pancreas development and β -cell function. *Endocr J.* 2017;64(5):477–86.
98. Gelaye B, Rondon M, Araya PR, A PM. ECM signaling regulates collective cellular dynamics to control pancreas branching morphogenesis. *2016;3(10):973–82.*
99. Pan FC, Wright C. Pancreas organogenesis: From bud to plexus to gland. *Dev Dyn.* 2011;240(3):530–65.
100. Pictet RL, Clark WR, Williams RH, Rutter WJ. An ultrastructural analysis of the developing embryonic pancreas. *Dev Biol.* 1972;29(4):436–67.
101. Jiang FX, Georges-Labouesse E, Harrison LC. Regulation of laminin 1-induced pancreatic beta-cell differentiation by alpha6 integrin and alpha-dystroglycan. *Mol Med.* 2001;7(2):107–14.
102. Krishnamurthy M, Wang R. Integrins and extracellular matrices in pancreatic tissue engineering. *Front Biosci Sch Ed.* 2009;1(February 2009):477–91.
103. Hynes RO. The extracellular matrix: not just pretty fibrils. *2013;326(5957):1216–9.*
104. Frantz C, Stewart KM, Weaver VM. The extracellular matrix at a glance. *J Cell Sci.* 2010;123(24):4195–200.
105. Lu P, Weaver VM, Werb Z. The extracellular matrix: A dynamic niche in cancer progression. *J Cell Biol.* 2012;196(4):395–406.
106. Yue B. Biology of the Extracellular Matrix: An Overview. *J Glaucoma.* 2014;1–8.
107. Hynes RO YK. *Extracellular Matrix Biology.* Cold Spring Harb Lab Press. 2011;

108. Star VL, Kleinman HK, Cannon FB, Hassell JR, Laurie GW, Martin GR, et al. Basement membrane complexes with biological activity. *Biochemistry*. 2005;25(2):312–8.
109. Carter W, Edgar D, Hohenester E, E E, Jones J, Aumailley M, et al. A simplified laminin nomenclature. *Matrix Biol*. 2005;24(5):326–32.
110. Yurchenco, Holly Colognato PD. Form and Function: The Laminin Family of Heterotrimers. *Dev Dyn*. 2000;213–34.
111. Yurchenco PD, Amenta PS, Patton BL. Basement membrane assembly, stability and activities observed through a developmental lens. *Matrix Biol*. 2004;22(7):521–38.
112. Miner JH, Yurchenco PD. Laminin Functions in Tissue Morphogenesis. *Annu Rev Cell Dev Biol*. 2004;20(1):255–84.
113. Timpl R, Brown JC. Supramolecular assembly of basement membranes. *BioEssays*. 1996;18(2):123–32.
114. Humphries JD. Integrin ligands at a glance. *J Cell Sci*. 2006;119(19):3901–3.
115. Horwitz AR, Huttenlocher A. Integrins in Cell Migration. *Cold Spring Harb Perspect Biol*. 2011;1–17.
116. Sitterley G. Laminin. *BioFiles*. 2008;3(8):11.
117. Miao G, Zhao Y, Li Y, Xu J, Gong H, Qi R, et al. Basement membrane extract preserves islet viability and activity in vitro by up-regulating $\alpha 3$ integrin and its signal. *Pancreas*. 2013;42(6):971–6.
118. Bouvard D, Bengtsson T, Gustafsson E, Fässler R, Berna A, Aszódi A, et al. Functional Consequences of Integrin Gene Mutations in Mice. *Circ Res*. 2007;89(3):211–23.
119. Kesavan G, Sand FW, Greiner TU, Johansson JK, Kobberup S, Wu X, et al. Cdc42-Mediated Tubulogenesis Controls Cell Specification. *Cell*. 2009;139(4):791–801.
120. Li Z, Manna P, Kobayashi H, Spilde T, Bhatia A, Preuett B, et al. Multifaceted pancreatic mesenchymal control of epithelial lineage selection. *Dev Biol*. 2004;269(1):252–63.
121. O’Neil JJ, Weir GC, Taneja M, Bonner-Weir S, Sharma A, Tatarkiewicz K, et al. In vitro cultivation of human islets from expanded ductal tissue. *Proc Natl Acad Sci*. 2002;97(14):7999–8004.
122. Lundin K, Ustinov J, Otonkoski T, Korsgren O, Pulkkinen M-A, Gao R. Characterization of Endocrine Progenitor Cells and Critical Factors for Their Differentiation in Human Adult Pancreatic Cell Culture. *Diabetes*. 2007;52(8):2007–15.

123. Kantengwa S, Baetens D, Sadoul K, Buck CA, Halban PA, Rouiller DG. Identification and characterization of $\alpha 3\beta 1$ integrin on primary and transformed rat islet cells. *Exp Cell Res.* 1997;237(2):394–402.
124. Jiang F-X, Harrison LC. Extracellular Signals and Pancreatic β -cell Development: A Brief Review. *Mol Med.* 2018;8(12):763–70.
125. Yebra M, Montgomery AMP, Diaferia GR, Kaido T, Silletti S, Perez B, et al. Recognition of the neural chemoattractant Netrin-1 by integrins $\alpha 6\beta 4$ and $\alpha 3\beta 1$ regulates epithelial cell adhesion and migration. *2003;5:695–707.*
126. Zorn TMT, Bevilacqua E, Abrahamsohn P. Collagen remodeling during decidualization in the mouse. *Cell Tissue Res.* 1986;244(2):443–8.
127. San Martin S, Soto-Suazo M, Ferreira de Oliveira S, Aplin JD, Abrahamsohn P, Zorn TMT. Small leucine-rich proteoglycans (SLRPs) in uterine tissues during pregnancy in mice. *Reproduction.* 2003;125(4):585–95.
128. Salgado RM, Favaro RR, San Martin S, Zorn TMT. The estrous cycle modulates small leucine-rich proteoglycans expression in mouse uterine tissues. *Anat Rec.* 2009;292(1):138–53.
129. Favaro RR, Aplin JD, Zorn TM, Capelo LP, Salgado RM, Glazier JD. Hormone-regulated expression and distribution of versican in mouse uterine tissues. *Reprod Biol Endocrinol.* 2009;7(1):60.
130. Salgado RM, Favaro RR, Zorn TMT. Modulation of small leucine-rich proteoglycans (SLRPs) expression in the mouse uterus by estradiol and progesterone. *Reprod Biol Endocrinol.* 2011;9(1):22.
131. Favaro RR, Salgado RM, Raspantini PR, Fortes ZB, Zorn TMT. Effects of long-term diabetes on the structure and cell proliferation of the myometrium in the early pregnancy of mice. *Int J Exp Pathol.* 2010;91(5):426–35.
132. Favaro RR, Salgado RM, Covarrubias AC, Bruni F, Lima C, Fortes ZB, et al. Long-term type 1 diabetes impairs decidualization and extracellular matrix remodeling during early embryonic development in mice. *Placenta.* 2013;34(12):1128–35.
133. Livak KJ, Schmittgen TD. Analysis of Relative Gene Expression Data Using Real-Time Quantitative PCR and the $2^{-\Delta\Delta CT}$ Method. *Methods.* 2001;25(4):402–8.
134. Chen YW, Chenier I, Tran S, Scotcher M, Chang SY, Zhang SL. Maternal diabetes programs hypertension and kidney injury in offspring. *Pediatr Nephrol.* 2010;25(7):1319–29.
135. Holemans K, Aerts L, Van Assche F a. Lifetime consequences of abnormal fetal pancreatic development. *J Physiol.* 2003;547(Pt 1):11–20.
136. Cnop M, Welsh N, Jonas JC, Jorns A, Lenzen S, Eizirik DL. Mechanisms of pancreatic beta-cell death in type 1 and type 2 diabetes: many differences, few similarities.

Diabetes. 2005;54 Suppl 2(6):S97-107.

137. Wewer Albrechtsen NJ, Kuhre RE, Pedersen J, Knop FK, Holst JJ. The biology of glucagon and the consequences of hyperglucagonemia. *Biomark Med*. 2016;10(11):1141–51.
138. Campbell JE, Drucker DJ. Islet α cells and glucagon-critical regulators of energy homeostasis. *Nat Rev Endocrinol*. 2015;11(6):329–38.
139. Kervran A, Randon J. Development of insulin release by fetal rat pancreas in vitro. Effects of glucose, amino acids, and theophylline. *Diabetes*. 1980;29(9):673–8.
140. Gittes GK, Galante P, Hanahan D, Rutter WJ, Debase HT. Lineage-specific morphogenesis in the developing pancreas: role of mesenchymal factors. *Development*. 1996;122(2):439–47.
141. Kim SK, Hebrok M. Intercellular signals regulating pancreas development and function. *Genes Dev*. 2001;15(2):111–27.
142. Guldager Kring Rasmussen D, Karsdal MA. Laminins. Biochemistry of Collagens, Laminins and Elastin: Structure, Function and Biomarkers. 2016. 163-196 p.
143. Daoud J, Petropavlovskaya M, Rosenberg L, Tabrizian M. The effect of extracellular matrix components on the preservation of human islet function in vitro. *Biomaterials*. 2010;31(7):1676–82.
144. Selg M, Horn N, Pausch F, Hallmann R, Wendler O, Sorokin LM. Expression and Function of Laminins in the Embryonic and Mature Vasculature. *Physiol Rev*. 2005;85(3):979–1000.
145. Rodgers KD, Barritt L, Miner JH, Cosgrove D. The laminins in the murine inner ear: Developmental transitions and expression in cochlear basement membranes. *Hear Res*. 2001;158(1–2):39–50.
146. Llacua A, De Haan BJ, Smink SA, De Vos P. Extracellular matrix components supporting human islet function in alginate-based immunoprotective microcapsules for treatment of diabetes. *J Biomed Mater Res - Part A*. 2016;104(7):1788–96.
147. Wahab NA, Schaefer L, Weston BS, Yiannikouris O, Wright A, Babelova A, et al. Glomerular expression of thrombospondin-1, transforming growth factor beta and connective tissue growth factor at different stages of diabetic nephropathy and their interdependent roles in mesangial response to diabetic stimuli. *Diabetologia*. 2005;48(12):2650–60.
148. Tarsio JF, Reger LA, Furcht LT. Molecular mechanisms in basement membrane complications of diabetes. Alterations in heparin, laminin, and type IV collagen association. *Diabetes*. 1988;37(5):532–9.

- ¹149. Reger LA, Tsilibary EC, Furcht LT, Wohlhueter RM, Charonis AS, Kouzi-Koliakos K, et al. Laminin alterations after in vitro nonenzymatic glycosylation. *Diabetes.* 2007;39(7):807–14.
150. Santos SAA Dos, Porto Amorim EM, Ribeiro LM, Rinaldi JC, Delella FK, Justulin LA, et al. Hyperglycemic condition during puberty increases collagen fibers deposition in the prostatic stroma and reduces MMP-2 activity. *Biochem Biophys Res Commun.* 2017;493(4):1581–6.
151. Gao T, McKenna B, Li C, Reichert M, Nguyen J, Singh T, et al. Pdx1 maintains β cell identity and function by repressing an α cell program. *Cell Metab.* 2014;19(2):259–71.
152. Schisler JC, Becker TC, Mirmira RG, Nesher R, Iype T, Griffen SC, et al. Mechanism of insulin Gene Regulation by the Pancreatic Transcription Factor Pdx-1. *J Biol Chem.* 2005;280(17):16798–807.
153. Robertson RP, Oseid EA, Tanaka Y, Hunter-Berger KK, Harmon JS, Gleason CE. In vivo prevention of hyperglycemia also prevents glucotoxic effects on PDX-1 and insulin gene expression. *Diabetes.* 2007;48(10):1995–2000.
154. Montminy M, Johnson T, Ferreri K, Leonard J, Sharma S, Jhala US. Hormonal regulation of an islet-specific enhancer in the pancreatic homeobox gene STF-1. *Mol Cell Biol.* 2015;17(5):2598–604.
155. Guerra S Del, Lupi R, Marselli L, Masini M, Bugiani M, Sbrana S, et al. Functional and molecular defects of pancreatic islets in human type 2 diabetes in Human Type 2 Diabetes. 2014;54(March 2005):727–35.
156. Smith SB, Ee HC, Conners JR, German MS. Paired-Homeodomain Transcription Factor PAX4 Acts as a Transcriptional Repressor in Early Pancreatic Development. *Mol Cell Biol.* 2015;19(12):8272–80.
157. Brun T, He KHH, Lupi R, Boehm B, Wojtusciszyn A, Sauter N, et al. The diabetes-linked transcription factor Pax4 is expressed in human pancreatic islets and is activated by mitogens and GLP-1. *Hum Mol Genet.* 2008;17(4):478–89.

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