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**ANÁLISE GENÔMICA DE *STREPTOMYCES OLINDENSIS* DAUFPE 5622 E DE
SUAS VIAS CRÍPTICAS PARA A OBTENÇÃO DE NOVOS METABÓLITOS
SECUNDÁRIOS DE INTERESSE BIOTECNOLÓGICO**

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RESUMO

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A caracterização de novos clusters biossintéticos tem se tornado uma prioridade dentro da pesquisa com actinobactérias devido ao potencial biotecnológico para produzir novas moléculas com possível uso na indústria e farmácia. Estes produtos naturais têm readquirido interesse pelas características particulares que possuem, por exemplo: biodisponibilidade, especificidade de alvo e diversidade de estruturas químicas. Ademais, a maioria das vias biossintéticas para a produção destas moléculas, descobertas no genoma de bactérias do gênero *Streptomyces*, permanecem crípticas ou silenciadas em condições de cultura no laboratório. Isto tem levado ao desenvolvimento de diversas técnicas e metodologias para a expressão destas vias metabólicas. Uma destas estratégias é a super-expressão de genes reguladores específicos dentro destas vias, como as proteínas reguladoras de antibióticos em *Streptomyces* (SARP, pelas siglas em Inglês). O laboratório de Bio-Produtos no Departamento de Microbiologia no Instituto de Ciências Biomédicas, na Universidade de São Paulo, tem trabalhado com a linhagem *Streptomyces olindensis* DAUFPE 5622, que produz uma molécula do grupo das antraciclinas chamada Cosmomicina D. Esta possui uma alta atividade antitumoral e tem sido de grande interesse devido ao padrão de glicosilação. Utilizando sequenciamento *Shotgun*, foram obtidos 120 contigs representando o genoma de *S. olindensis*, os quais foram anotados e submetidos ao NCBI sob o número de acesso JJOH00000000. Utilizando o software antiSMASH e anotação manual, foram identificados trinta e três clusters envolvidos na produção de metabólitos secundários. Dentro destes, foram encontrados clusters gênicos para a produção de metabólitos previamente descritos, com diversas atividades biológicas, como a cosmomicina D, melanina, pigmento de esporos, fator-A, geosmina, albaflavenona, hopanoides, amiquelina, hidroxiectoina, desferrioxamina e fosacetamicina. Os outros vinte dois clusters possivelmente produzem derivados de compostos já conhecidos ou moléculas novas, motivo pelo qual, foi realizada uma análise de genômica comparativa para identificar, caracterizar e anotar cada via biossintética achada em *S. olindensis*. Subsequentemente, foram encontrados diversos reguladores específicos de vias metabólicas, tomando especial interesse os SARP. Foram escolhidos dois clusters biossintéticos para a super-expressão (Aminociclitol e um Policitídeo Tipo I) destes genes, levando a detecção do composto sob condições de cultura. Estudos posteriores permitirão identificar a estrutura química destes compostos e propor diferentes metodologias para a obtenção de produtos naturais de interesse biotecnológico, sintetizados por *Streptomyces olindensis*.

Palavras-chave: *Streptomyces*. *Genome mining*. Anotação de genoma. *Cluster* biossintético. Cosmomicina D. Super-expressão de genes reguladores

ABSTRACT

FERREIRA-TORRES M.A. **Analysis of *Streptomyces olindensis* DAUFPE 5622 genome and its cryptic pathways to obtain new secondary metabolites of biotechnological interest.** 2015. 103 p. Master thesis (Microbiology) - Instituto de Ciências Biomédicas, Universidade de São Paulo, São Paulo, 2015.

Characterization of new metabolic biosynthetic clusters has become a priority among researchers in actinobacteria because of their biotechnological potential to produce new molecules with possible use in the pharmaceutical and industrial field. These natural products regain interest because of their particular characteristics, for example: bioavailability, target specificity and chemical structure diversity. Moreover, most of the biosynthetic pathways for the production of these molecules that have been discovered on the genomes of *Streptomyces* remain cryptic or silenced under standard laboratory conditions, which lead to the development of different technical approaches to express these new clusters. One of the strategies is the overexpression of the specific regulatory genes within these pathways, such as *Streptomyces* Antibiotic Regulatory Protein (SARP) to detect these new products in different culture media. The Bio products Laboratory, in the Microbiology Department in the Biomedical Science Institute at São Paulo's University, has been working with *Streptomyces olindensis* DAUFPE 5622, the producer of the anthracycline Cosmomycin D, which has an important antitumoral activity and has attracted interest because of its distinctive glycosylation pattern. Using Shotgun sequencing the genome was obtained, organized in 120 contigs, annotated and submitted in the NCBI under the accession number JJOH00000000, and by further prediction were identified thirty-three secondary metabolite clusters using the antiSMASH server and manual annotation. Among them, were found gene clusters for the production of known metabolites with different biological activity such as Cosmomycin D, Melanin, Spore pigment, A-factor, Geosmin, Albaflavenone, Hopene, Amychelin, Hidroxectoine, Desferrioxamine B and Fosacetamycin. The other twenty-two clusters were likely to produce derivatives of known compound or new molecules, which is why, a comparative genomic study was carried out to identify, characterize and annotate every genome cluster found in *S. olindensis*. Subsequently, several pathway-specific regulators in the described biosynthetic clusters were found, arising special interest the SARP, choosing two different pathways (Aminocyclitol and Polyketide Type I), to over express the regulatory genes, in order to detect the biological product under culture conditions. Further studies will allow the identity of the chemical structure of these products to be found and propose different approaches to obtain other natural products of interest synthetized by *Streptomyces olindensis*.

Keywords: *Streptomyces*. Genome mining. Genome annotation. Biosynthetic Cluster. Cosmomycin D. Overexpression of regulator genes.

1 INTRODUÇÃO

Historicamente, os metabólitos secundários de origem microbiana têm proporcionado uma grande quantidade de moléculas de interesse industrial para uso farmacêutico e outros campos na biotecnologia. Até 2010, tinham sido descritos mais de 500.000 sendo bioativos o 20%. Desta porção bioativa, 35.000 são produzidos por micro-organismos e o 35% é produzida por actinobactérias, sendo o restante produzido por fungos e outras bactérias (BÉRDY, 2012). Embora o desenvolvimento de novos fármacos tenha diminuído na última década, o interesse por novas estruturas químicas se mantém aberto na indústria devido as propriedades intrínsecas dos produtos naturais como a biodisponibilidade das moléculas, a complexidade estrutural e a especificidade de alvo (HARVEY; EDRADA-EBEL; QUINN, 2015). A interdisciplinaridade na pesquisa tem proporcionado diversas ferramentas para a busca de novas biomoléculas, encontrando, assim, diferentes *clusters* biosintéticos no genoma de vários micro-organismos, incluindo actinobactérias (BENTLEY et al., 2002; BÉRDY, 2012). Pode-se detectar domínios conservados típicos de enzimas que produzem moléculas com atividade biológica, caracterizadas quimicamente como policetídeos (PKS), peptídeos não ribossomais (NRPS), terpenos, peptídeos de síntese ribossomal, sideróforos, etc. Estes descobrimentos indicam um grande potencial biotecnológico para obtenção de novos antibióticos, compostos antitumorais e moléculas com outra aplicação biotecnológica (BÉRDY, 2005, 2012; CHALLIS, 2014; NETT; IKEDA; MOORE, 2009).

Em *Streptomyces coelicolor*, a actinobactéria modelo, foram encontrados 29 *clusters* biosintéticos dentro do seu genoma dedicados à produção de metabólitos secundários de interesse industrial, e em *Streptomyces avermitilis* detectaram-se 37 *clusters* para a produção de novas biomoléculas (NETT; IKEDA; MOORE, 2009). Estes dados são um indício do potencial codificado no genoma das actinobactérias, sendo candidatas à busca de novos produtos metabólicos, inicialmente pelo uso de ferramentas de bioinformática para sua caracterização e expressão. A maioria destes novos *clusters*, encontrados em diversos microrganismos, não é detectada em condições de fermentação padrão em laboratório, por isso são chamados de *clusters* ou vias crípticas (BÉRDY, 2012; NETT; IKEDA; MOORE, 2009; OCHI;

HOSAKA, 2013). Este fato leva à busca de novas estratégias para a expressão destes compostos, como a diversificação de meios de cultura e condições de crescimento para a obtenção de novas moléculas (OSMAC approach) (BODE et al., 2002) ou a produção em hospedeiros heterólogos (BALTZ, 2010; STARCEVIC et al., 2012).

No laboratório de Bioproductos vem se trabalhando com a bactéria *Streptomyces olindensis*, isolada da região de Olinda, no estado de Pernambuco, Brasil. Esta bactéria produz o agente antitumoral Cosmomicina D, amplamente estudada, já que a molécula possui um padrão de glicosilação pouco comum, que potencializa a sua atividade contra diversas linhas celulares tumorais. Além disso, possui uma maior quantidade de genes de resistência que ajudam na saída do composto e na sobrevivência da bactéria (FURLAN et al., 2004; GARRIDO et al., 2006).

A sequência do genoma foi descrita previamente por Rojas e colaboradores em 2014, para ganhar um maior entendimento sobre o cluster gênico da Cosmomicina D e expandir as possibilidades na área de biossíntese combinatória para, assim, modificar a atividade biológica da molécula. Neste estudo é apresentada uma análise completa do genoma como também a comparação com outras espécies de *Streptomyces* relevantes no campo dos compostos bioativos. Finalmente é revelado o potencial codificado para uma variedade de novos produtos naturais, além do agente antitumoral Cosmomicina D, o que levará ao desenvolvimento de diferentes estratégias para a expressão e obtenção de novas moléculas.

5 CONCLUSÕES

- A descrição das características físicas permitiu comparar a topologia do cromossomo de *S. olindensis* com genomas de referência, atribuindo à linearidade *in-silico* e identificando proteínas envolvidas na plasticidade e receptividade do cromossomo, encontrando a possível inserção do *cluster* do metabólito híbrido entre melanina e nucleosídeo.
- A classificação funcional das proteínas codificadas no genoma de *Streptomyces olindensis* DAUFPE 5622, atribui mais do 50% das regiões codificadoras à funções do metabolismo primário e 2.8% das proteínas foram relacionadas com metabolismo secundário.
- A análise de genômica comparativa, utilizando a linhagem *S. olindensis* DAUFPE 5622 e quatro espécies próximas altamente conhecidas, permitiu elucidar a conformação do core genoma em bactérias deste gênero, relatando vias de metabolismo primário, e alguns metabólitos secundários amplamente distribuídos em actinobactérias como sideróforos e exopolisacarídeos.
- Utilizando os grupos de ortólogos da base de dado do KEGG, conseguiu-se determinar as funções dos elementos únicos presentes em *S. olindensis*, destacando-se as proteínas envolvidas na produção de metabólitos secundários como policetídeo sintases, NRP sintases, terpeno sintases entre outros.
- A ferramenta antiSMASH permitiu a identificação da maioria dos *clusters* biossintéticos, facilitando a caracterização das vias metabólicas visando a produção de novos produtos naturais bioativos.
- Realizando a anotação funcional e caracterização dos *clusters* metabólicos, logrou-se realizar uma comparação com vias metabólicas já conhecidas para assim, evitar o redescobrimento de moléculas descritas previamente.

- A identificação e anotação das 34 vias biossintéticas permitiu determinar a presença de genes reguladores, selecionando os clusters biossintéticos a serem ativados.

REFERÊNCIAS*

- AHLERT, J. et al. Identification of *stsC*, the gene encoding the L-glutamine:scyllo-inosose aminotransferase from streptomycin-producing Streptomyces. **Archives of Microbiology**, v. 168, n. 2, p. 102–113, 1997.
- AIGLE, B. et al. Genome mining of *Streptomyces ambofaciens*. **Journal of Industrial Microbiology & Biotechnology**, v. 41, n. 2, p. 251–263, 2014.
- AIGLE, B.; CORRE, C. **Waking up Streptomyces secondary metabolism by constitutive expression of activators or genetic disruption of repressors**. [s.l.] Elsevier Inc., 2012. v. 517
- AJITHKUMAR, V.; PRASAD, R. The activator/repressor protein DnrO of *Streptomyces peucetius* binds to DNA without changing its topology. **International Journal of Biological Macromolecules**, v. 46, n. 3, p. 380–384, 2010.
- ALAM, M. T. et al. Genome-based phylogenetic analysis of *Streptomyces* and its relatives. **Molecular Phylogenetics and Evolution**, v. 54, n. 3, p. 763–772, 2010.
- ALAM, M. T. et al. Comparative genome-scale metabolic modeling of actinomycetes: The topology of essential core metabolism. **FEBS Letters**, v. 585, n. 14, p. 2389–2394, 2011.
- ARNISON, P. G. et al. Ribosomally synthesized and post-translationally modified peptide natural products: overview and recommendations for a universal nomenclature. **Natural Product Reports**, v. 30, n. 1, p. 108–160, 2013.
- AZIZ, R. K. et al. The RAST Server: rapid annotations using subsystems technology. **BMC Genomics**, v. 9, n. 1, p. 75, 2008.
- BALTZ, R. H. *Streptomyces* and *Saccharopolyspora* hosts for heterologous expression of secondary metabolite gene clusters. **Journal of Industrial Microbiology & Biotechnology**, v. 37, n. 8, p. 759–772, 2010.
- BARANASIC, D. et al. Draft genome sequence of *Streptomyces rapamycinicus* strain NRRL5491, the producer of the immunosuppressant rapamycin. **Genome Announcements**, v. 1, n. 4, p. 8766, 2013.
- BENTLEY, S. et al. Complete genome sequence of the model actinomycete *Streptomyces coelicolor A3(2)*. **Nature**, v. 417, n. 6885, p. 141–147, 2002.
- BÉRDY, J. Bioactive microbial metabolites. **The Journal of Antibiotics**, v. 58, n. 1, p. 1–26, 2005.

* De acordo com:

ASSOCIAÇÃO BRASILEIRA DE NORMAS TÉCNICAS. **NBR 6023**: informação e documentação: referências: elaboração. Rio de Janeiro, 2002.

- BÉRDY, J. Thoughts and facts about antibiotics: Where we are now and where we are heading. **The Journal of Antibiotics**, v. 65, n. 8, p. 441–441, 2012.
- BIBB, M. J. Regulation of secondary metabolism in streptomycetes. **Current Opinion in Microbiology**, v. 8, n. 2, p. 208–215, 2005.
- BIEBER, L. W. et al. The anthracycline complex retamycin,. Structure. Determination of the major constituents. **Journal of Natural Products**, v. 2, p. 385–388, 1989.
- BLAŽIČ, M. et al. Annotation of the modular polyketide synthase and nonribosomal peptide synthetase gene clusters in the genome of *Streptomyces tsukubaensis* NRRL18488. **Applied and Environmental Microbiology**, v. 78, n. 23, p. 8183–8190, 2012.
- BODE, H. B. et al. Big effects from small changes : possible ways to explore nature ' s chemical diversity. **ChemBioChem**, v. 3, p. 619–627, 2002.
- BORISOVA, S. A. et al. Biosynthesis of rhizococcins, antifungal phosphonate oligopeptides produced by *Bacillus subtilis* ATCC6633. **Chemistry & Biology**, v. 17, n. 1, p. 28–37, 2010.
- BURSY, J. et al. Synthesis and uptake of the compatible solutes ectoine and 5-hydroxyectoine by *Streptomyces coelicolor* A3(2) in Response to Salt and Heat Stresses. **Applied and Environmental Microbiology**, v. 74, n. 23, p. 7286–7296, 2008.
- CANE, D. E. E.; IKEDA, H. Exploration and mining of the bacterial terpenome. **Accounts of Chemical Research**, v. 45, n. 3, p. 463–472, 2012.
- CHALLIS, G. L. Exploitation of the *Streptomyces coelicolor* A3(2) genome sequence for discovery of new natural products and biosynthetic pathways. **Journal of Industrial Microbiology & Biotechnology**, v. 41, n. 2, p. 219–232, 2014.
- CHANDRA, G.; CHATER, K. F. Developmental biology of *Streptomyces* from the perspective of 100 actinobacterial genome sequences. **FEMS Microbiology Reviews**, v. 38, n. 3, p. 345–379, 2014.
- CHEN, D. et al. Improvement of FK506 Production in *Streptomyces tsukubaensis* by genetic enhancement of the supply of unusual polyketide extender units via utilization of two distinct site-specific recombination systems. **Applied and Environmental Microbiology**, v. 78, n. 15, p. 5093–5103, 2012.
- CHOI, W. S.; WU, X.; CHOENG, Y. Genetic organization of the putative salbostatin biosynthetic gene cluster including the 2- epi -5- epi -valiolone synthase gene in *Streptomyces albus* ATCC 21838. **Applied Genetics and Molecular Biotechnology**, p. 637–645, 2008.
- CIMERMANCIC, P. et al. Insights into secondary metabolism from a global analysis of prokaryotic biosynthetic gene clusters. **Cell**, v. 158, n. 2, p. 412–421, 2014.

- CIONI, J. P. et al. Cyanohydrin phosphonate natural product from *Streptomyces regensis*. **Journal of Natural Products**, v. 77, n. 2, p. 243–249, 2014.
- COLE, S. P. et al. Biosynthesis of the antibiotic actinorhodin. Analysis of blocked mutants of *Streptomyces coelicolor*. **Journal of Antibiotics (Tokyo)**, v. 40, n. 3, p. 340–347, 1987.
- CONTRERAS HERNÁNDEZ C. A. **Caracterização de genes biossintéticos do antitumoral cosmomicina D**. 2013. 105 f. Dissertação (Mestrado em Microbiologia) - Instituto de Ciências Biomédicas, Universidade de São Paulo, São Paulo, 2013.
- DELCHER, A. L. et al. Identifying bacterial genes and endosymbiont DNA with Glimmer. **Bioinformatics**, v. 23, n. 6, p. 673–679, 2007.
- DEMAIN, A. L. Importance of microbial natural products and the need to revitalize their discovery. **Journal of Industrial Microbiology & Biotechnology**, v. 41, n. 2, p. 185–201, 2014.
- DEMAIN, A. L.; ADARIO, J. L. Contributions of Microorganisms to Industrial Biology. **Molecular Biotechnology**, v. 38, n. 1, p. 41–55, 2008.
- DEMYDCHUK, Y. et al. Analysis of the tetracycline gene cluster: insights into the biosynthesis of a polyether tetrone antibiotic. **ChemBioChem**, v. 9, n. 7, p. 1136–1145, 2008.
- DOROGHAZI, J. R.; BUCKLEY, D. H. Intraspecies comparison of *Streptomyces pratinus* genomes reveals high levels of recombination and gene conservation between strains of disparate geographic origin. **BMC genomics**, v. 15, n. 1, p. 970, 2014.
- DOROGHAZI, J. R.; METCALF, W. W. Comparative genomics of actinomycetes with a focus on natural product biosynthetic genes. **BMC Genomics**, v. 14, n. 1, p. 611, 2013.
- DUNN, B. J.; KHOSLA, C. Engineering the acyltransferase substrate specificity of assembly line polyketide synthases. **Journal of the Royal Society, Interface / the Royal Society**, v. 10, n. 85, p. 297, 2013.
- DURHAM, A. M. et al. EGene: a configurable pipeline generation system for automated sequence analysis. **Bioinformatics**, v. 21, n. 12, p. 2812–2813, 2005.
- EDDY, S. R. Accelerated profile HMM searches. **PLoS Computational Biology**, v. 7, n. 10, p. e1002195, 2011.
- EDGAR, R. C. MUSCLE: multiple sequence alignment with high accuracy and high throughput. **Nucleic Acid Research**, v. 32, n. 5, p. 1792–1797, 2004.
- ENGEL, R. Phosphonates as analogues of natural phosphates. **Chemical Reviews**, v. 77, p. 349–367, 1977.

- EVANS, B. S. et al. Discovery of the antibiotic phosacetamycin via a new mass spectrometry-based method for phosphonic acid detection. **ACS chemical biology**, v. 8, n. 5, p. 908–913, 2013.
- FELNAGLE, E. A. et al. Nonribosomal peptide synthetases involved in the production of medically relevant natural products. **Molecular Pharmaceutics**, v. 5, n. 2, p. 191–211, 2008.
- FINKING, R.; MARAHIEL, M. A. Biosynthesis of Nonribosomal Peptides. **Annual Review of Microbiology**, v. 58, n. 1, p. 453–488, 2004.
- FINN, R. D.; CLEMENTS, J.; EDDY, S. R. HMMER web server: interactive sequence similarity searching. **Nucleic Acids Research**, v. 39, p. W29–37, 2011.
- FISCHBACH, M. A; WALSH, C. T. Assembly-line enzymology for polyketide and nonribosomal peptide antibiotics: logic, machinery, and mechanisms. **Chemical Reviews**, v. 5, p. 3468–3496, 2006.
- FUNA, N. et al. Biosynthesis of hexahydroxyperylenequinone melanin via oxidative aryl coupling by cytochrome P-450 in *Streptomyces griseus*. **Journal of Bacteriology**, v. 187, n. 23, p. 8149–8155, 2005.
- FURLAN, R. L. A et al. DNA-binding properties of cosmomycin D, an anthracycline with two trisaccharide chains. **The Journal of Antibiotics**, v. 57, n. 10, p. 647–54, 2004.
- GAN, H. et al. Comparative genomic analysis of six bacteria belonging to the genus *Novosphingobium*: insights into marine adaptation, cell-cell signaling and bioremediation. **BMC Genomics**, v. 14, n. 1, p. 431, 2013.
- GARRIDO, L. M. et al. Insights in the glycosylation steps during biosynthesis of the antitumor anthracycline cosmomycin: characterization of two glycosyltransferase genes. **Applied Microbiology and Biotechnology**, v. 73, n. 1, p. 122–131, 2006.
- GETSIN, I. et al. Comparative genomics of transport proteins in developmental bacteria: *Myxococcus xanthus* and *Streptomyces coelicolor*. **BMC Microbiology**, v. 13, n. 1, p. 279, 2013.
- GOMEZ-ESCRIBANO, J. P.; BIBB, M. J. Heterologous expression of natural product biosynthetic gene clusters in *Streptomyces coelicolor*: from genome mining to manipulation of biosynthetic pathways. **Journal of Industrial Microbiology & Biotechnology**, v. 41, n. 2, p. 425–431, 2014.
- GOODFELLOW, M.; WILLIAMS, S. T. Ecology of actinomycetes. **Annual Review of Microbiology**, v. 37, n. 41, p. 189–216, 1983.
- GUO, J. et al. High-level production of melanin by a novel isolate of *Streptomyces kathirae*. **FEMS Microbiology Letters**, v. 357, n. 1, p. 85–91, 2014.

- GUYET, A. et al. Regulation of the clpP1-clpP2 operon by the pleiotropic regulator AdpA in *Streptomyces lividans*. **Archives of Microbiology**, v. 195, n. 12, p. 831–841, 2013.
- HARVEY, A. L.; EDRADA-EBEL, R.; QUINN, R. J. The re-emergence of natural products for drug discovery in the genomics era. **Nature Reviews Drug Discovery**, v. 14, n. 2, p. 111–129, 2015.
- HERTWECK, C. The biosynthetic logic of polyketide diversity. **Angewandte Chemie International edition**, v. 48, n. 26, p. 4688–4716, jan. 2009.
- HOPWOOD, D. A. Forty years of genetics with *Streptomyces*: from in vivo through in vitro to in silico. **Microbiology**, v. 145, p. 2183–2202, 1999.
- HOPWOOD, D. A. Soil To Genomics : The Streptomyces Chromosome. **Annual Review of Genetics**, v. 40, p. 1–23, 2006.
- HUANG, C. H. et al. The telomeres of *Streptomyces* chromosomes contain conserved palindromic sequences with potential to form complex secondary structures. **Molecular Microbiology**, v. 28, n. 5, p. 905–916, 1998.
- HUTCHINGS, M. I. et al. The E cell envelope stress response of *Streptomyces coelicolor* is influenced by a novel lipoprotein, CseA. **Journal of Bacteriology**, v. 188, n. 20, p. 7222–7229, 2006.
- HWANG, K.-S. et al. Systems biology and biotechnology of *Streptomyces* species for the production of secondary metabolites. **Biotechnology Advances**, v. 32, n. 2, p. 255–268, 2014.
- IKEDA, H. et al. Complete genome sequence and comparative analysis of the industrial microorganism *Streptomyces avermitilis*. **Nature Biotechnology**, v. 21, n. 5, p. 526–531, 2003.
- JANDA, J. M.; ABBOTT, S. L. 16S rRNA gene sequencing for bacterial identification in the diagnostic laboratory: pluses, perils, and pitfalls. **Journal of Clinical Microbiology**, v. 45, n. 9, p. 2761–2764, 2007.
- JIANG, J.; HE, X.; CANE, D. E. Geosmin biosynthesis. *Streptomyces coelicolor* germacradienol/germacrene D synthase converts farnesyl diphosphate to geosmin. **Journal of the American Chemical Society**, v. 128, n. 25, p. 8128–8129, 2006.
- JOHNSON, M. et al. NCBI BLAST: a better web interface. **Nucleic Acids Research**, v. 36, p. W5–W9, 2008.
- JU, K.-S.; DOROGHAZI, J. R.; METCALF, W. W. Genomics-enabled discovery of phosphonate natural products and their biosynthetic pathways. **Journal of Industrial Microbiology & Biotechnology**, v. 41, n. 2, p. 345–356, 2014.

- KÄLL, L.; KROGH, A.; SONNHAMMER, E. L. A combined transmembrane topology and signal peptide prediction method. **Journal of Molecular Biology**, v. 338, n. 5, p. 1027–1036, 2004.
- KATO, J. et al. Biosynthesis of gamma-butyrolactone autoregulators that switch on secondary metabolism and morphological development in *Streptomyces*. **Proceedings of the National Academy of Sciences of the United States of America**, v. 104, p. 2378–2383, 2007.
- KIESER, T.; BIBB, M. J.; BUTTNER, M. J.; CHATER, K. F.; HOPWOOD, D. A. **Practical Streptomyces Genetics**. Norwich: The John Innes Foundation, 2002. p
- KIM, J.-N. et al. Comparative genomics reveals the core accessory genomes of *Streptomyces* species. **Journal of Microbiology and Biotechnology**, 2015.
- KOTOWSKA, M.; PAWLIK, K. Roles of type II thioesterases and their application for secondary metabolite yield improvement. **Applied Microbiology and Biotechnology**, p. 7735–7746, 2014.
- KROGH, A. et al. Predicting transmembrane protein topology with a hidden markov model: application to complete genomes. **Journal of Molecular Biology**, v. 305, n. 3, p. 567–580, 2001.
- KUDO, F. et al. Cloning of the pactamycin biosynthetic gene cluster and characterization of a crucial glycosyltransferase prior to a unique cyclopentane ring formation. **The Journal of Antibiotics**, v. 60, n. 8, p. 492–503, 2007.
- KUDO, F.; EGUCHI, T. Biosynthetic genes for aminoglycoside antibiotics. **The Journal of Antibiotics**, v. 62, n. 9, p. 471–481, 2009.
- LEADLAY, P. F. et al. Engineering of complex polyketide biosynthesis--insights from sequencing of the monensin biosynthetic gene cluster. **Journal of Industrial Microbiology & Biotechnology**, v. 27, n. 6, p. 360–367, 2001.
- LEZHAVA, A et al. Physical map of the linear chromosome of *Streptomyces griseus*. **Journal of Bacteriology**, v. 177, n. 22, p. 6492–8, 1995.
- LIU, G. et al. Molecular regulation of antibiotic biosynthesis in *Streptomyces*. **Microbiology and Molecular Biology Reviews**, v. 77, n. 1, p. 112–143, 2013.
- MAHMUD, T. Progress in aminocyclitol biosynthesis. **Current Opinion in Chemical Biology**, v. 13, n. 2, p. 161–170, 2009.
- MAHMUD, T.; LEE, S.; FLOSS, H. G. The biosynthesis of acarbose and validamycin. **Chemical record**, v. 1, n. 4, p. 300–310, 2001.
- MAO, X. et al. Automated genome annotation and pathway identification using the KEGG Orthology (KO) as a controlled vocabulary. **Bioinformatics**, v. 21, n. 19, p. 3787–3793, 2005.

MARTÍN, J. F.; LIRAS, P. Engineering of regulatory cascades and networks controlling antibiotic biosynthesis in *Streptomyces*. **Current Opinion in Microbiology**, v. 13, n. 3, p. 263–273, 2010.

MEDEMA, M. H. The minimum information about a biosynthetic gene cluster (MIBiG) specification. **Nature Chemical Biology**, p. 625–631, 2015.

MENCHER, R. J.; HEIM, A. H. Melanin biosynthesis by *Streptomyces lavendulae*. **Journal of Genetic Microbiology**, n. 28, p. 665–670, 1962.

METCALF, W. W.; VAN DER DONK, W. A. Biosynthesis of phosphonic and phosphinic acid natural products. **Annual Review of Biochemistry**, v. 78, p. 65–94, 2009.

MITCHELL, A. et al. The InterPro protein families database: the classification resource after 15 years. **Nucleic Acids Research**, v. 43, n. D1, p. D213–D221, 2015.

MOTAMEDI, H.; SHAFIEE, A. The biosynthetic gene cluster for the macrolactone ring of the immunosuppressant FK506. **European Journal of Biochemistry**, v. 256, n. 3, p. 528–534, 1998.

NAKANO, C.; HORINOUCHI, S.; OHNISHI, Y. Characterization of a novel sesquiterpene cyclase involved in (+)-caryolan-1-ol biosynthesis in *Streptomyces griseus*. **Journal of Biological Chemistry**, v. 286, n. 32, p. 27980–27987, 2011.

NETT, M.; IKEDA, H.; MOORE, B. S. Genomic basis for natural product biosynthetic diversity in the actinomycetes. **Natural Product Reports**, v. 26, n. 11, p. 1362, 2009.

NEWMAN, D. J.; CRAGG, G. M. Natural products as sources of new drugs over the 30 years from 1981 to 2010. **Journal of Natural Products**, v. 75, n. 3, p. 311–335, 2012.

NINDITA, Y. et al. The *tap-tpg* gene pair on the linear plasmid functions to maintain a linear topology of the chromosome in *Streptomyces rochei*. **Molecular Microbiology**, v. 95, n. 5, p. 846–858, 2015.

NOVAKOVA, R.; BISTAKOVA, J.; KORMANEC, J. Characterization of the polyketide spore pigment cluster whiESa in *Streptomyces aureofaciens* CCM3239. **Archives of Microbiology**, v. 182, n. 5, p. 388–395, 2004.

OCHI, K.; HOSAKA, T. New strategies for drug discovery: activation of silent or weakly expressed microbial gene clusters. **Applied Microbiology and Biotechnology**, v. 97, n. 1, p. 87–98, 2013.

OLANO, C.; MÉNDEZ, C.; SALAS, J. A. Antitumor Compounds from Marine Actinomycetes. **Marine Drugs**, v. 7, n. 2, p. 210–248, 2009.

OMURA, S. et al. Genome sequence of an industrial microorganism *Streptomyces avermitilis*: deducing the ability of producing secondary metabolites. **Proceedings of**

the National Academy of Sciences of the United States of America, v. 98, n. 21, p. 12215–20, 2001.

OVERBEEK, R. et al. The SEED and the rapid annotation of microbial genomes using subsystems technology (RAST). **Nucleic Acids Research**, v. 42, n. D1, p. D206–D214, 2014.

PAN, J.-J. et al. Biosynthesis of squalene from farnesyl diphosphate in bacteria: three steps catalyzed by three enzymes. **ACS Central Science**, v. 1, n. 2, p. 77–82, 2015.

PETERSEN, T. N. et al. SignalP 4.0: discriminating signal peptides from transmembrane regions. **Nature Methods**, v. 8, n. 10, p. 785–786, 2011.

PHELAN, R. M. et al. Engineering terpene biosynthesis in *Streptomyces* for production of the advanced biofuel precursor bisabolene. **ACS Synthetic Biology**, v. 4, n. 4, p. 393–399, 2014.

PORALLA, K.; MUTH, G.; HÄRTNER, T. Hopanoids are formed during transition from substrate to aerial hyphae in *Streptomyces coelicolor* A3(2). **FEMS Microbiology Letters**, v. 189, p. 93–95, 2000.

POWELL, S. et al. eggNOG v4.0: nested orthology inference across 3686 organisms. **Nucleic Acids Research**, v. 42, n. D1, p. D231–D239, 2014.

PRASEUTH, A P. et al. Complete sequence of biosynthetic gene cluster responsible for producing triostin A and evaluation of quinomycin-type antibiotics from *Streptomyces triostinicus*. **Biotechnol Prog**, v. 24, n. 6, p. 1226–1231, 2008.

RABYK, M. V.; OSTASH, B. O.; FEDORENKO, V. O. Gene networks regulating secondary metabolism in actinomycetes: Pleiotropic regulators. **Cytology and Genetics**, v. 48, n. 1, p. 55–67, 2014.

ROJAS, J. D. et al. Genome sequence of *Streptomyces olindensis* DAUFPE 5622, producer of the antitumoral anthracycline cosmomycin D. **Genome Announcements**, v. 2, n. 3, p. 4–5, 2014.

SADEGHI, A. et al. Diversity of the ectoines biosynthesis genes in the salt tolerant *Streptomyces* and evidence for inductive effect of ectoines on their accumulation. **Microbiological Research**, v. 169, n. 9–10, p. 699–708, 2014.

SALAS, J. A.; MÉNDEZ, C. Engineering the glycosylation of natural products in actinomycetes. **Trends in Microbiology**, v. 15, n. 5, p. 219–232, 2007.

SCHMIDT, E. W. The hidden diversity of ribosomal peptide natural products. **BMC biology**, v. 8, p. 83, 2010.

SCHMOOCK, G. et al. Functional Cross-talk between Fatty Acid Synthesis and Nonribosomal Peptide Synthesis in Quinoxaline Antibiotic-producing Streptomycetes. **Journal of Biological Chemistry**, v. 280, n. 6, p. 4339–4349, 2005.

- SCHWECKE, T. et al. The biosynthetic gene cluster for the polyketide immunosuppressant rapamycin. **Proceedings of the National Academy of Sciences of the United States of America**, v. 92, n. 17, p. 7839–7843, 1995.
- SCHWIENTEK, P. et al. The complete genome sequence of the acarbose producer *Actinoplanes* sp. SE50/110. **BMC Genomics**, v. 13, n. 1, p. 112, 2012.
- SEIPKE, R. F.; KALTENPOTH, M.; HUTCHINGS, M. I. *Streptomyces* as symbionts: An emerging and widespread theme? **FEMS Microbiology Reviews**, v. 36, n. 4, p. 862–876, 2012.
- SIEBER, S. A; MARAHIEL, M. A. Learning from nature's drug factories: nonribosomal synthesis of macrocyclic peptides. **Journal of Bacteriology**, v. 185, n. 24, p. 7036–7043, 2003.
- SIVAPERUMAL, P. et al. Melanin from marine *Streptomyces* sp. (MVCS13) with potential effect against ornamental fish pathogens of *Carassius auratus* (Linnaeus, 1758). **Biocatalysis and Agricultural Biotechnology**, v. 3, n. 4, p. 134–141, 2014.
- SOUSA, T. D. S. et al. Anthracyclones from *Micromonospora* sp. **Journal of Natural Products**, v. 75, n. 3, p. 489–493, 2012.
- STARCEVIC, A. et al. Recombinatorial biosynthesis of polyketides. **Journal of Industrial Microbiology & Biotechnology**, v. 39, n. 3, p. 503–511, 2012.
- TAMURA, K. et al. MEGA6: Molecular evolutionary genetics analysis version 6.0. **Molecular Biology and Evolution**, v. 30, n. 12, p. 2725–2729, 2013.
- UDWARY, D. et al. Genome sequencing reveals complex secondary metabolome in the marine actinomycete *Salinispora tropica*. **Proceedings of the National Academy of Sciences** v. 104, n. 25, p. 10376–10381, 2007.
- UNTERGASSER, A. et al. Primer3-new capabilities and interfaces. **Nucleic Acids Research**, v. 40, n. 15, p. 1–12, 2012.
- VAN WEZEL, G. P.; McDOWALL, K. J. The regulation of the secondary metabolism of *Streptomyces*: new links and experimental advances. **Natural Product Reports**, v. 28, n. 7, p. 1311, 2011.
- VASANTHAKUMAR, A.; KATTUSAMY, K.; PRASAD, R. Regulation of daunorubicin biosynthesis in *Streptomyces peucetius* - feed forward and feedback transcriptional control. **Journal of Basic Microbiology**, v. 53, n. 8, p. 636–644, 2013.
- VENTURA, M. et al. Genomics of Actinobacteria: tracing the evolutionary history of an ancient phylum. **Microbiology and Molecular Biology Reviews**, v. 71, n. 3, p. 495–548, 2007.
- VERNIKOS, G. S.; PARKHILL, J. Interpolated variable order motifs for identification of horizontally acquired DNA: revisiting the *Salmonella* pathogenicity islands. **Bioinformatics**, v. 22, n. 18, p. 2196–2203, 2006.

- VOLFF, J.-N.; ALTENBUCHNER, J. MicroReview Genetic instability of the. **Molecular Microbiology**, v. 27, p. 239–246, 1998.
- WALSH, C. T.; FISCHBACH, M. A. Natural products version 2.0: Connecting genes to molecules. **Journal of the American Chemical Society**, v. 132, n. 8, p. 2469–2493, 2010.
- WANG, H. et al. Genetic screening strategy for rapid access to polyether ionophore producers and products in Actinomycetes. **Applied and Environmental Microbiology**, v. 77, n. 10, p. 3433–3442, 2011.
- WANG, H. et al. Atlas of nonribosomal peptide and polyketide biosynthetic pathways reveals common occurrence of nonmodular enzymes. **Proceedings of the National Academy of Sciences**, v. 111, n. 25, p. 9259–9264, 2014.
- WANG, H.; VAN DER DONK, W. A. Biosynthesis of the class III lantipeptide catenulipeptin. **ACS Chemical Biology**, v. 7, p. 1529–1535, 2012.
- WANG, K. C.; OHNUMA S. Isoprenyl diphosphate synthases. **Biochimica et Biophysica Acta**, v. 1529, p. 33–49, 2000.
- WANG, X. et al. Characterization and analysis of an industrial strain of *Streptomyces bingchengensis* by genome sequencing and gene microarray. **NRC research press**, v. 689, p. 677–689, 2013.
- WEBER, T. et al. antiSMASH 3.0--a comprehensive resource for the genome mining of biosynthetic gene clusters. **Nucleic Acids Research**, v. 43, n. W1, p. W237–W243, 2015.
- WEISSMAN, K. J. Introduction to polyketide biosynthesis. In: HOPWOOD, D.A. (Ed) **Methods in Enzymology: Complex Enzymes in Microbial Natural Product Biosynthesis, Part B: Polyketides, Aminocoumarins and Carbohydrates**. 2009, v. 459. p. 3–16.
- WOLAŃSKI, M.; JAKIMOWICZ, D.; ZAKRZEWSKA-CZERWIŃSKA, J. AdpA, key regulator for morphological differentiation regulates bacterial chromosome replication. **Open Biology**, v. 2, n. 7, p. 120097, 2012.
- WRIGHT, F.; BIBB, M. J. Codon usage in the G+C-rich *Streptomyces* genome. **Gene**, v. 113, n. 1, p. 55–65, 1992.
- XU, L. et al. Complete genome sequence and comparative genomic analyses of the vancomycin-producing *Amycolatopsis orientalis*. **BMC Genomics**, p. 1–18, 2014.
- YAMADA, Y. et al. Terpene synthases are widely distributed in bacteria. **Proceedings of the National Academy of Sciences of the United States of America**, v. 112, n. 3, p. 857–862, 2015.

YANG, C.-C. et al. Mutational Analysis of the Terminal Protein Tpg of *Streptomyces* Chromosomes: Identification of the Deoxynucleotidylation Site. **PLoS ONE**, v. 8, n. 2, p. e56322, 2013.

YU, T. W.; HOPWOOD, D. A. Ectopic expression of the *Streptomyces coelicolor* whiE genes for polyketide spore pigment synthesis and their interaction with the act genes for actinorhodin biosynthesis. **Microbiology**, v. 141, n. 11, p. 2779–2791, 1995.

YU, X. et al. Diversity and abundance of phosphonate biosynthetic genes in nature. **Proceedings of the National Academy of Sciences**, v. 110, n. 51, p. 20759–20764, 2013.

ZABURANNYI, N. et al. Insights into naturally minimised *Streptomyces albus* J1074 genome. **BMC Genomics**, v. 15, n. 1, p. 97, 2014.

ZHAO, B. et al. Biosynthesis of the Sesquiterpene Antibiotic Albaflavenone in *Streptomyces coelicolor* A3(2). **Journal of Biological Chemistry**, v. 283, n. 13, p. 8183–8189, 2008.

ZHOU, Z. et al. Genome plasticity and systems evolution in *Streptomyces*. **BMC Bioinformatics**, v. 13 Suppl 1, n. Suppl 10, p. S8, 2012.

ZOU, Z. et al. A γ -butyrolactone-sensing activator/repressor, JadR3, controls a regulatory mini-network for jadomycin biosynthesis. **Molecular Microbiology**, v. 94, n. 3, p. 490–505, 2014.